

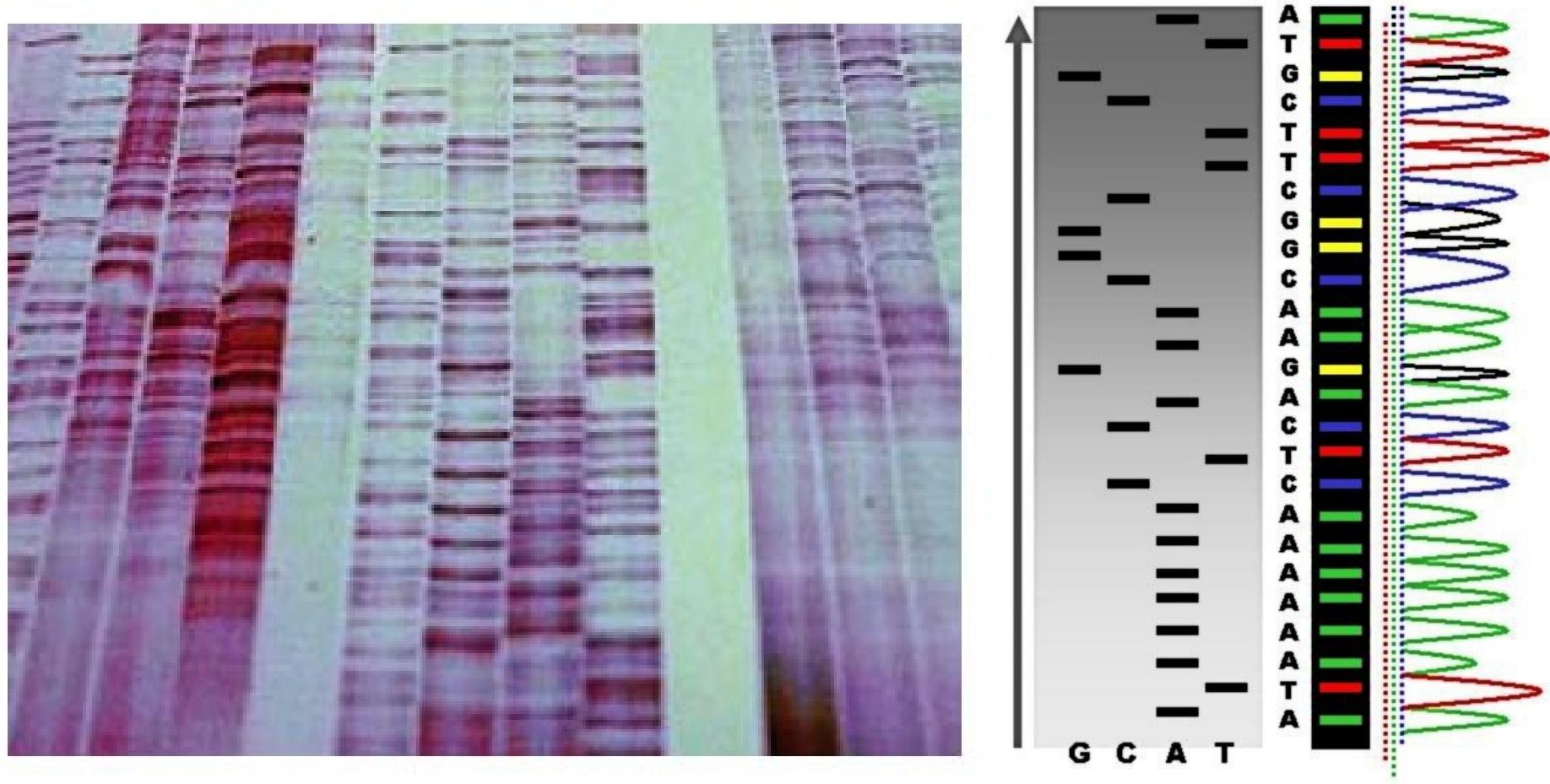
Introduction to next-generation sequencing

Simon van Heeringen
November 3, 2014

So, you want to do sequencing...?

3'Seq 3-seq 3P-seq AHT-ChIP-seq ARS-seq
ATAC-seq BOINC-seq BS-seq Bar-seq BisChIP-
seq Bru-seq Bubble-seq CAB-seq CAGE-seq
CHART-seq CLASH-seq CNV-seq CRE-seq
Capture-C-seq Cel-seq ChIA-PET-seq ChIP-seq
ChIRP-seq Chem-seq Chip-exo-seq Cir-seq
DMS-seq DNase-seq DNasel-seq Dup-seq
FAIRE-seq FRAG-seq FRT-seq Frac-seq Freq-
seq GRO-seq GTI-seq HELP-seq HITs-KIN-seq
Hi-C-seq HiTS-Flip-seq IMS-MDA-seq IN-seq Ig-
seq Immuno-seq MeDI-seq MeDIP-seq Methyl-seq Mu-seq
NET-seq NOME-seq Nascent-seq Novel-seq
Nucleo-seq PAI-seq PAR-Clip-seq PARS-seq

Sanger sequencing



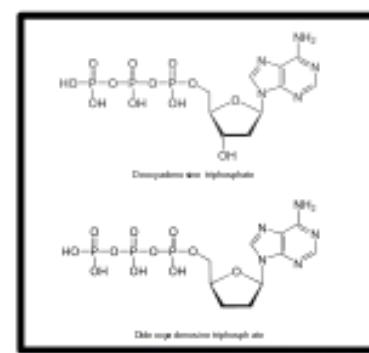
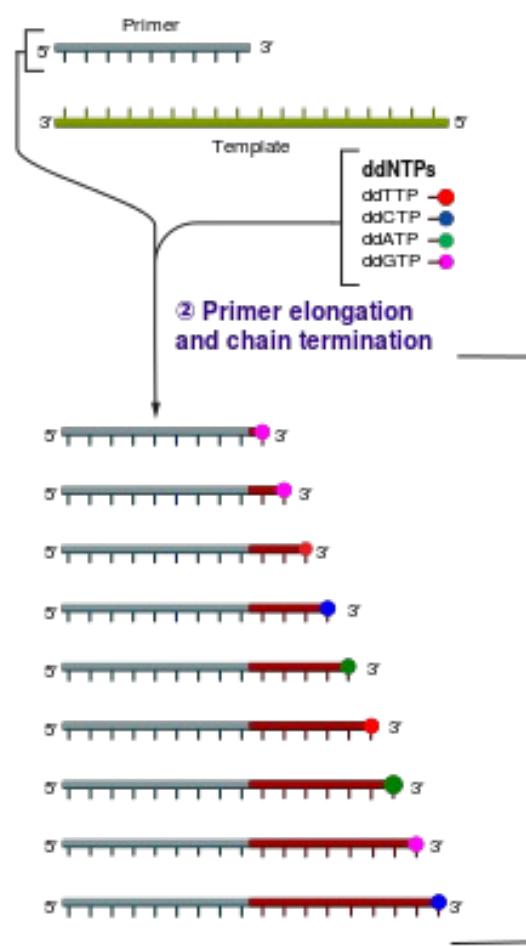
Nature Methods, 2008

Wikipedia

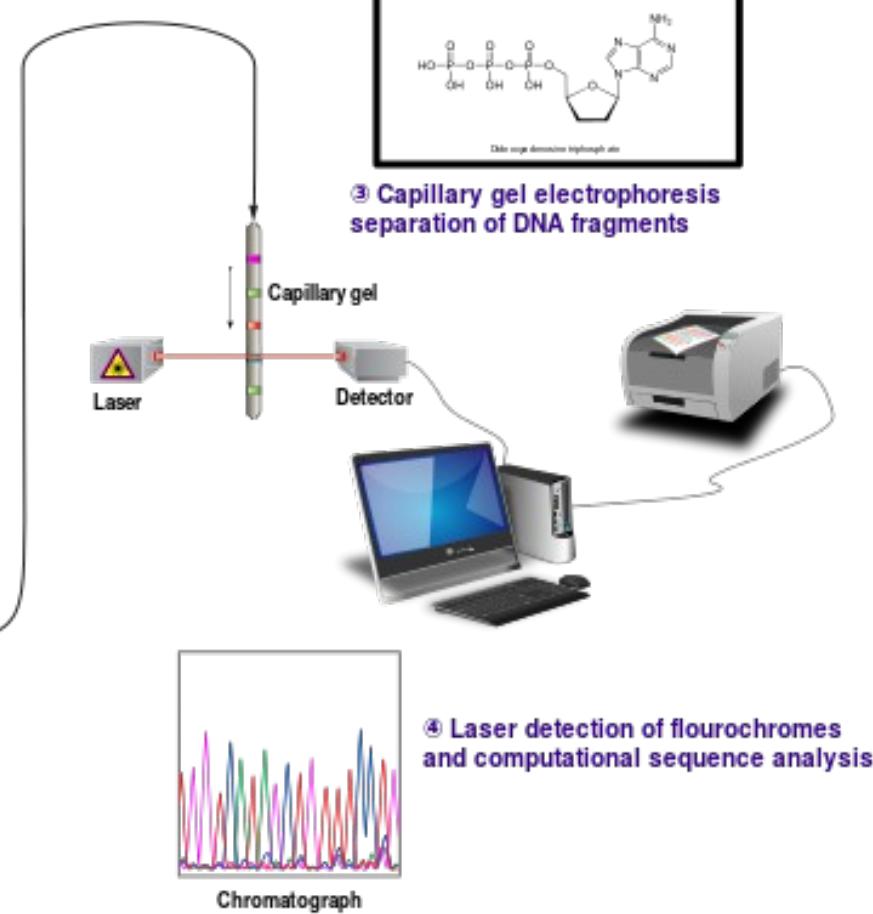
Sanger sequencing

① Reaction mixture

- Primer and DNA template → DNA polymerase
- ddNTPs with flourochromes → dNTPs (dATP, dCTP, dGTP, and dTTP)

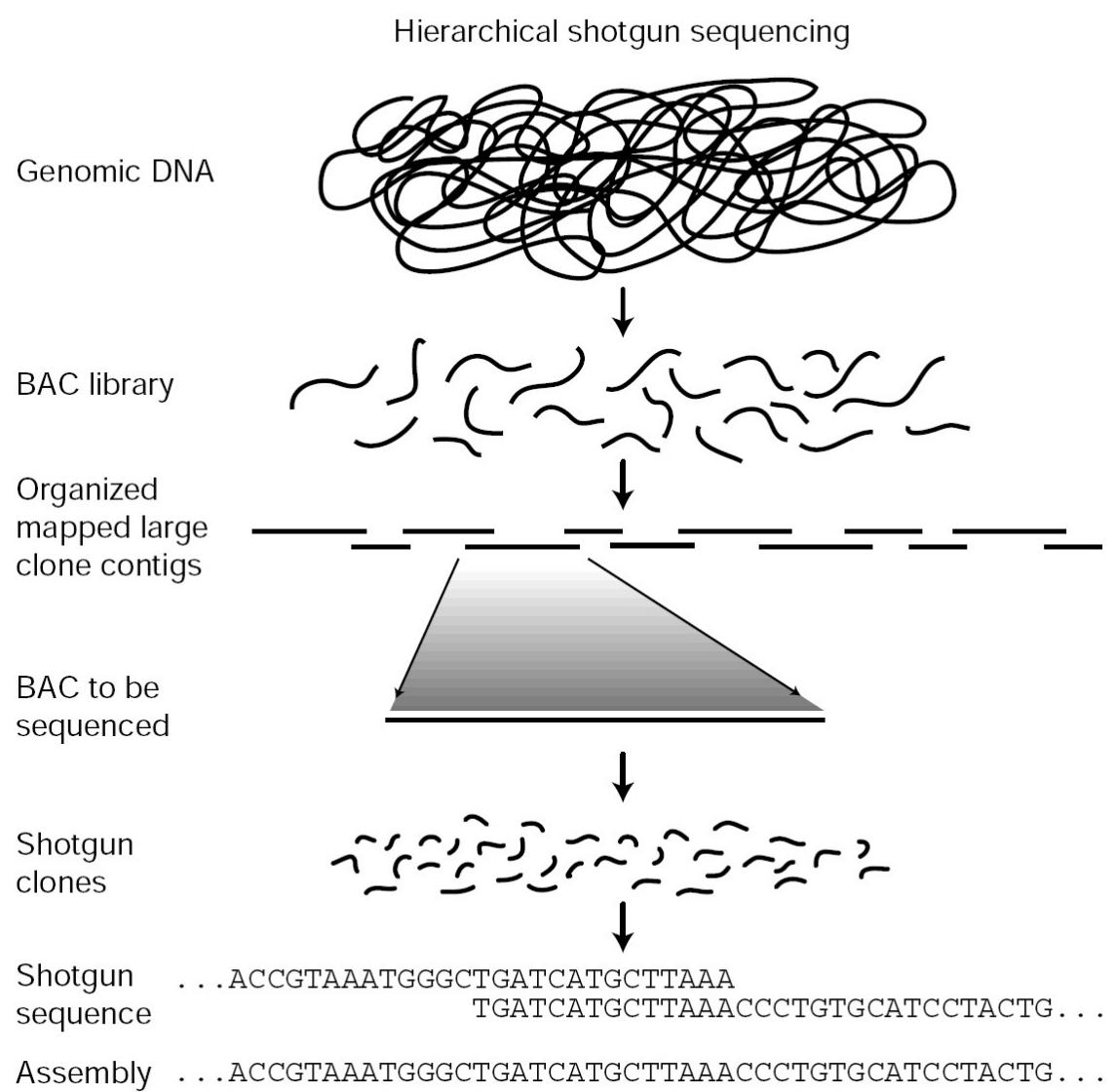


③ Capillary gel electrophoresis separation of DNA fragments



CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGCACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTATCCTCAAGTAATCCGCCCGCTGGCCTCCAAAGTGCAGGGGTGAGGCCAC
AAAAATGGTTATGGAGATCAAAATAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

Shotgun sequencing



Nature, 2001

2001: draft of the human genome



“Next-generation” sequencing

“Next-generation” sequencing

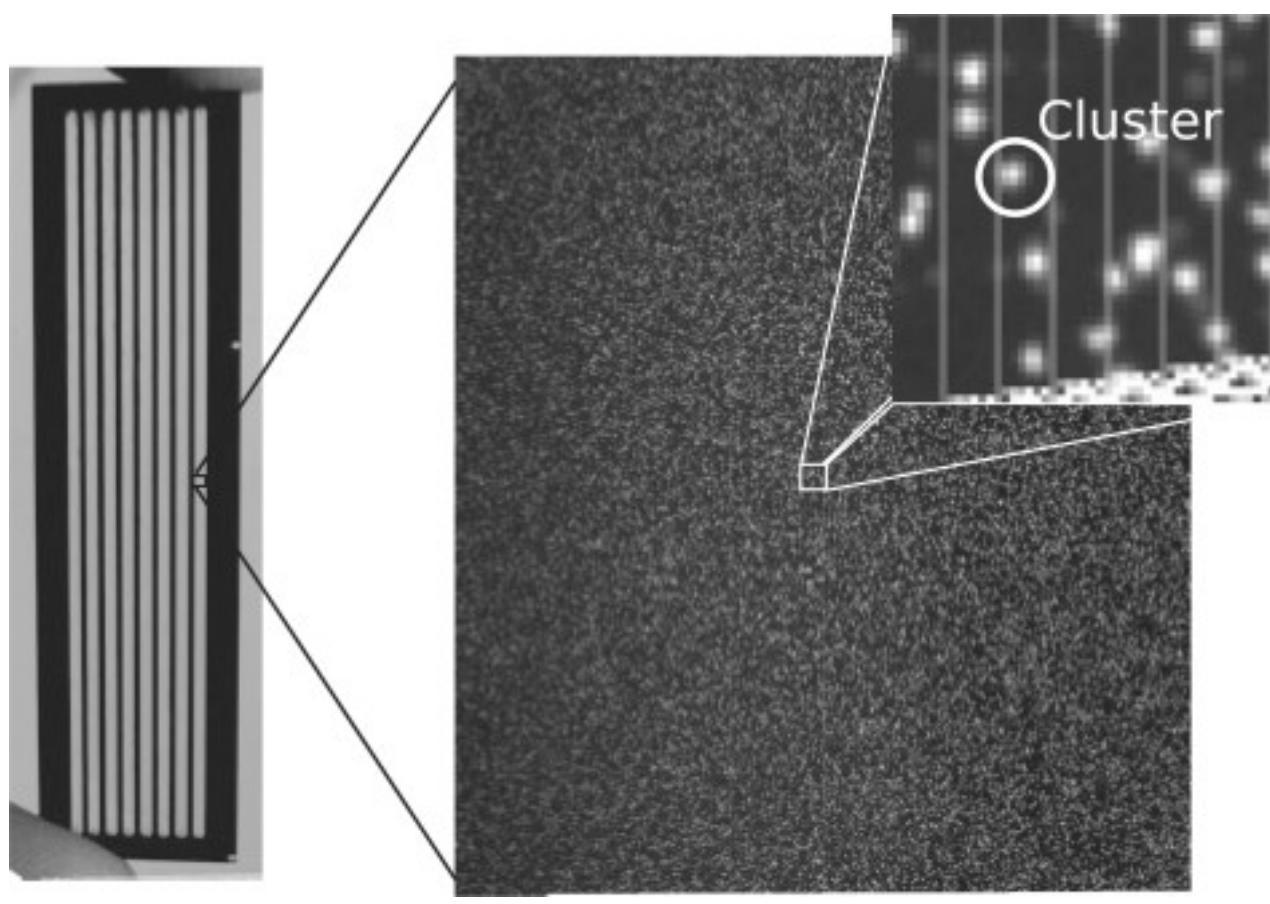
or

.. a whole lotta reads..

Illumina sequencing

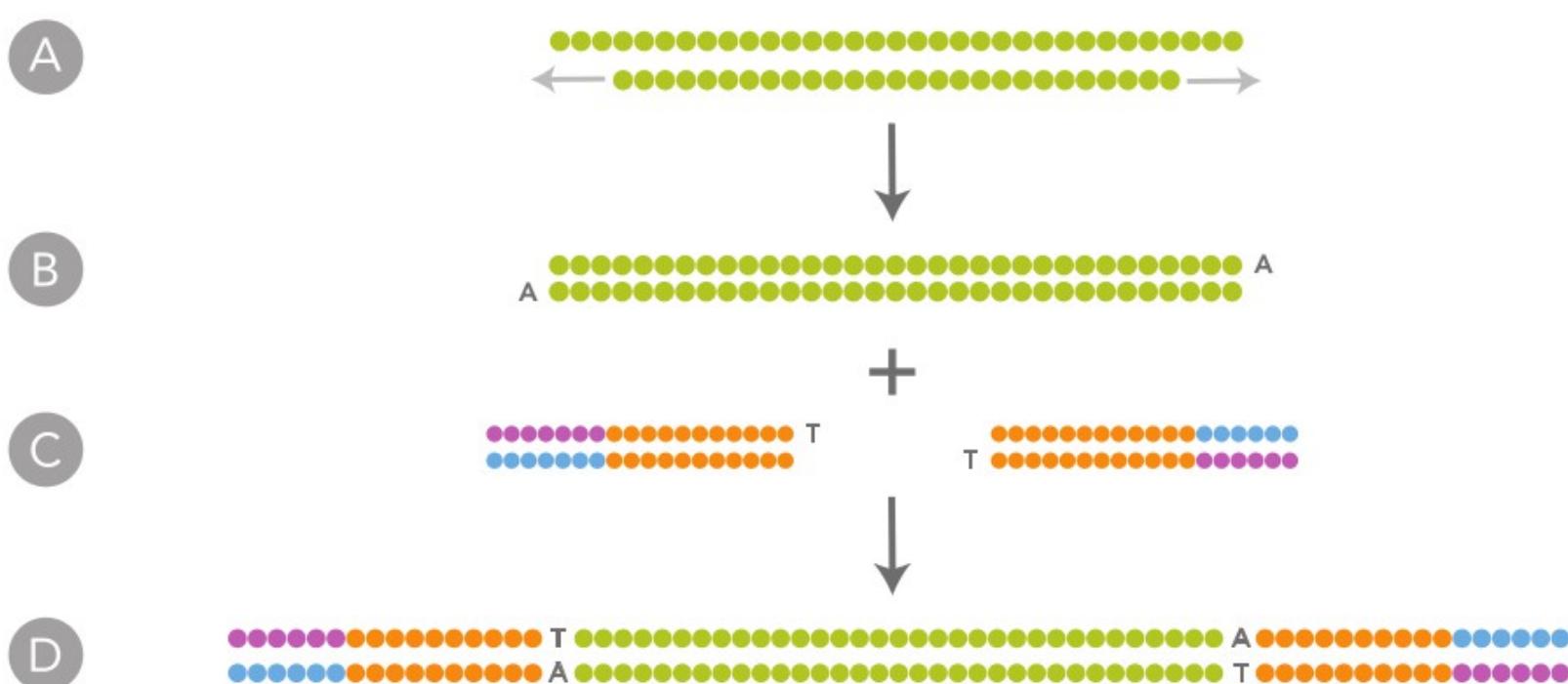


An Illumina flowcell

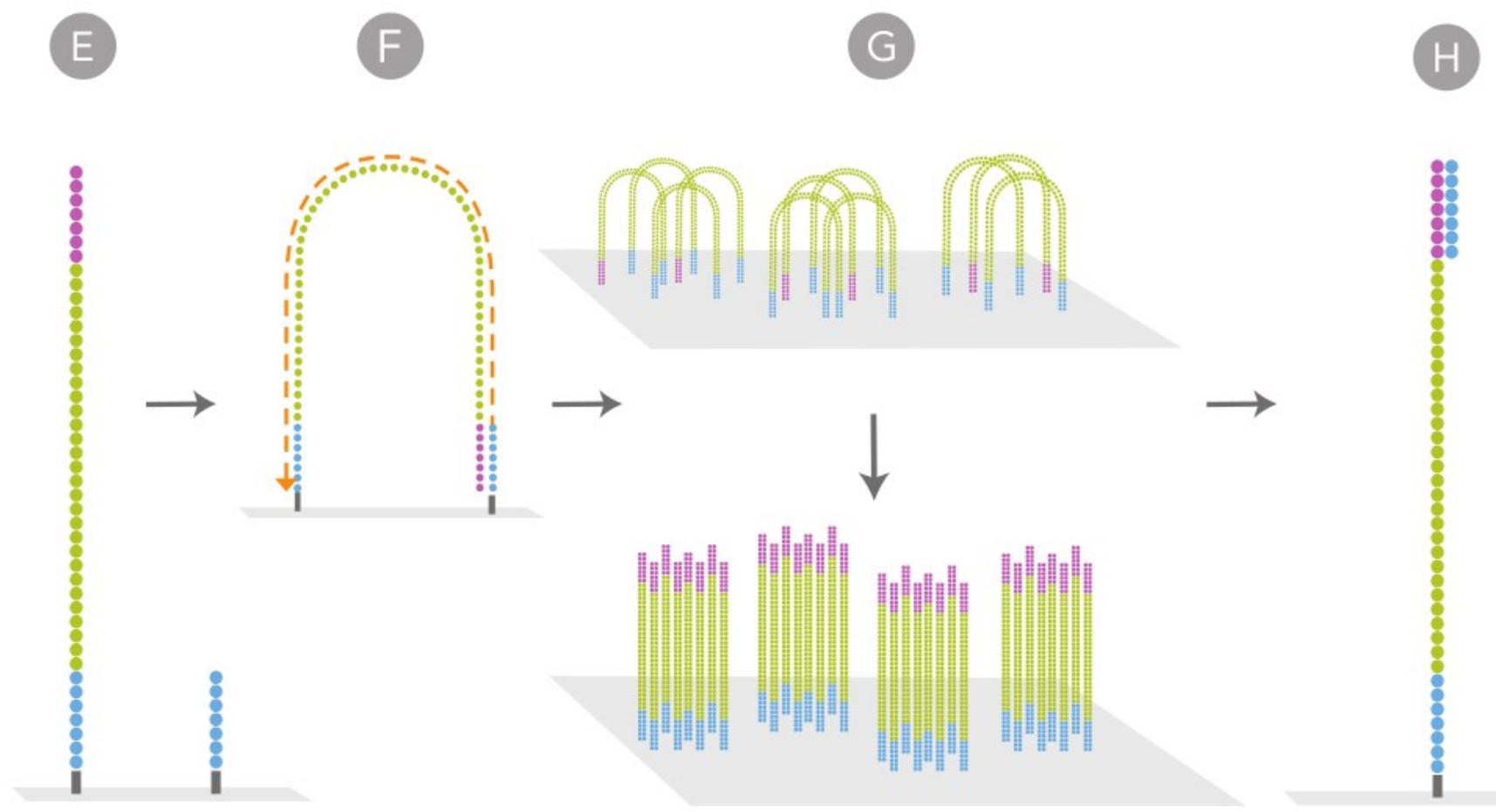


Whiteford et al., Bioinformatics, 2009

Sequencing by synthesis

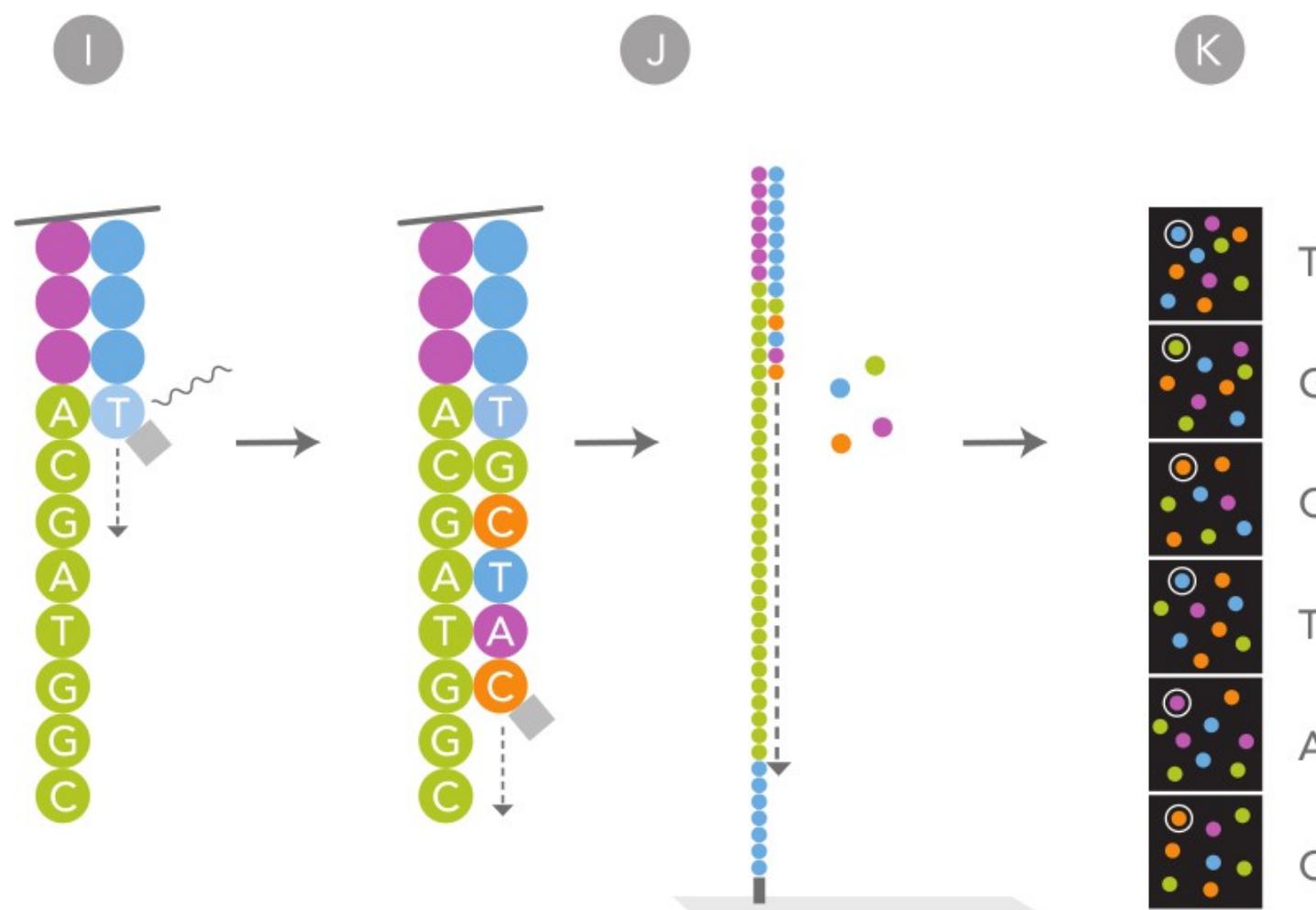


Sequencing by synthesis



CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGACTCCTCAAGTAATCCGCCCGCTCGGCCTCCAAAGTGCAGGGGTGAGGCCAC
AAAAATGGTTATGGAGATCAAAATAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

Sequencing by synthesis



CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTATCCTCAAGTAATCCGCCCGCCTGGCCTCCAAAGTGCAGGGGTGAGCCAC
AAAAATGGTTATGGAGATCAAAATAAGGTGGGGTGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

Illumina sequencing systems



MiSeq

Focused power. Speed and simplicity for targeted and small genome sequencing.

NextSeq 500

Flexible power. Speed and simplicity for everyday genomics.

HiSeq 2500

Production power. Power and efficiency for large-scale genomics.

HiSeq X*

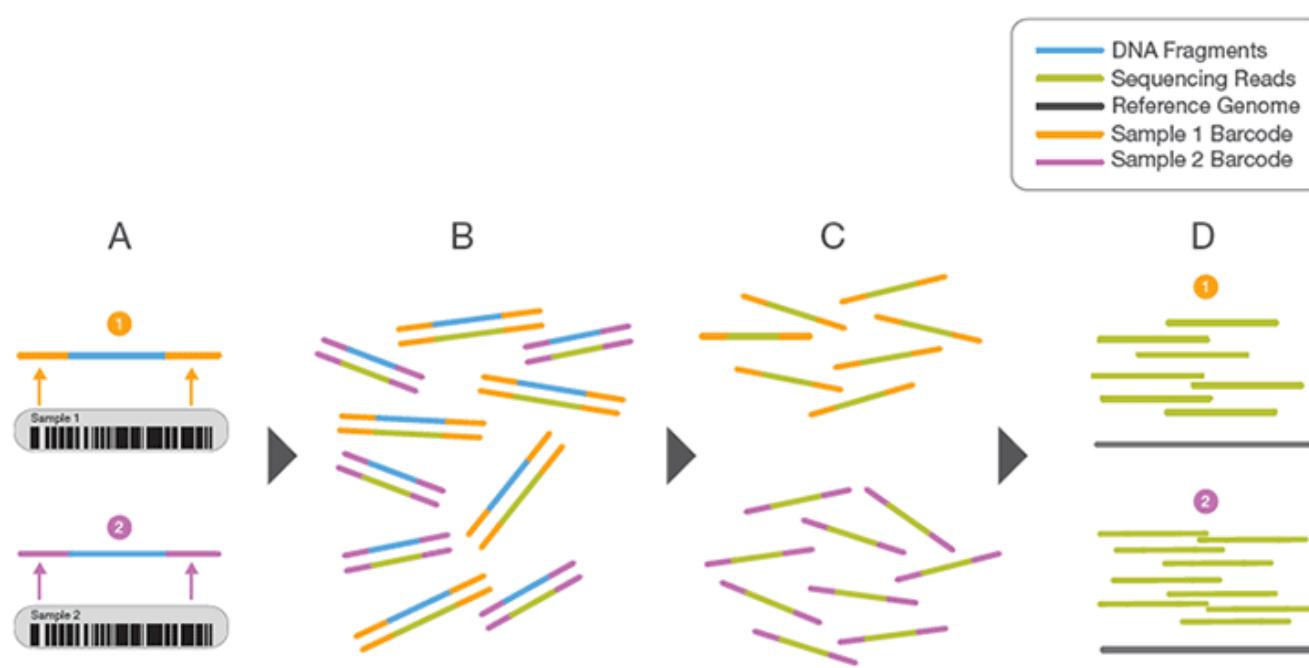
Population power. \$1,000 human genome and extreme throughput for population-scale sequencing.

Key applications	Small genome, amplicon, and targeted gene panel sequencing.	Everyday genome, exome, transcriptome sequencing, and more.	Production-scale genome, exome, transcriptome sequencing, and more.	Population-scale human whole-genome sequencing.	
Run mode	N/A	Mid-Output	High-Output	Rapid Run	High-Output
Flow cells processed per run	1	1	1	1 or 2	1 or 2
Output range	0.3-15 Gb	20-39 Gb	30-120 Gb	10-300 Gb	50-1000 Gb
Run time	5-55 hours	15-26 hours	12-30 hours	7-60 hours	< 1 day - 6 days
Reads per flow cell†	25 Million‡	130 Million	400 Million	300 Million	2 Billion
Maximum read length	2 × 300 bp	2 × 150 bp	2 × 150 bp	2 × 250 bp	2 × 125 bp

CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTATCCTCAAGTAATCCGCCGCCCTGGCCTCCAAAGTGCAGGGGTGAGGCCAC
AAAAATGGTTATGGAGATCAAAATAAGGTGGGGTGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCTG

Multiplexing

Figure 2: Conceptual Overview of Sample Multiplexing



- Two representative DNA fragments from two unique samples, each attached to a specific barcode sequence that identifies the sample from which it originated.
- Libraries for each sample are pooled and sequenced in parallel. Each new read contains both the fragment sequence and its sample-identifying barcode.
- Barcode sequences are used to de-multiplex, or differentiate reads from each sample.
- Each set of reads is aligned to the reference sequence.

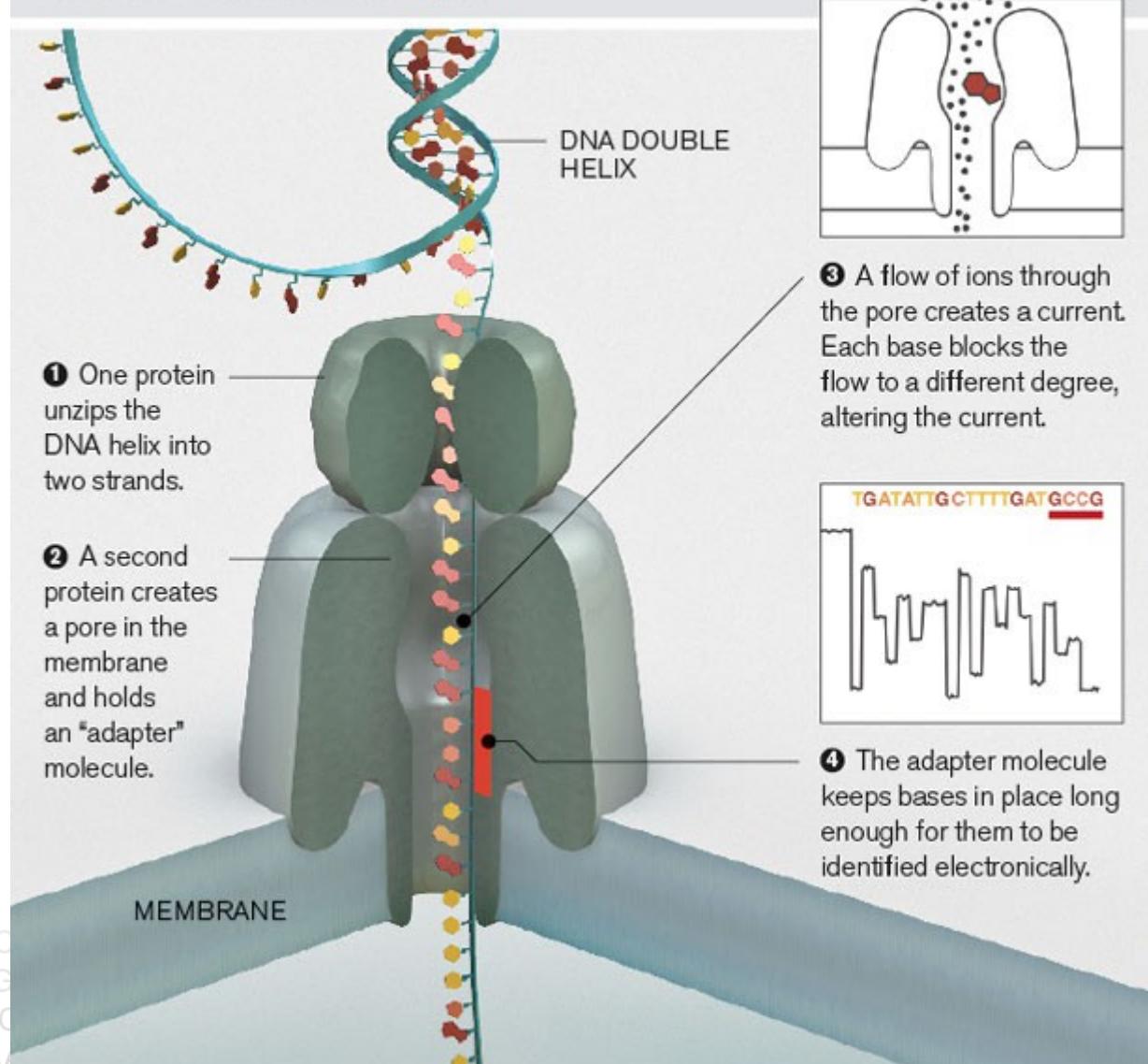
Illumina

Other technologies...



Nanopore sequencing

DNA can be sequenced by threading it through a microscopic pore in a membrane. Bases are identified by the way they affect ions flowing through the pore from one side of the membrane to the other.



The MinION



CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGCACAGAAGCCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGACTCCTCAAGTAATCCGCCCGCTCGGCCTCCAAAGTGCAGGGCGTGAGGCCAC
AAAAATGGTTATGGAGATCAAAATAAGGTGGGGTGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

Pacific Biosciences



What's next?

- It's a rapidly evolving field
- Existing technologies continually improve
- Illumina started out as a small player 10 years ago (Solexa)
- Sequencing as a service?
- Benchtop (USB-sized?) sequencers?

The FASTQ format

```
@D256N5M1:31:C1B42ACXX:4:2305:3881:47
605
ACCCCCCACAGGGACCCCTTGTCACGTCCCCCTAACTC
CCTGC
+
@?
@FDFFFDFFFDBGIIIIIGDGHIG@GHIIIGEF@@@DFH
GGI
```

FASTQ quality scores

Phred quality score

$$Q_{\text{sanger}} = -10 \log_{10} p$$

Q10	1 in 10	90%
Q20	1 in 100	99%
Q30	1 in 1000	99.9%
Q40	1 in 10000	99.99%

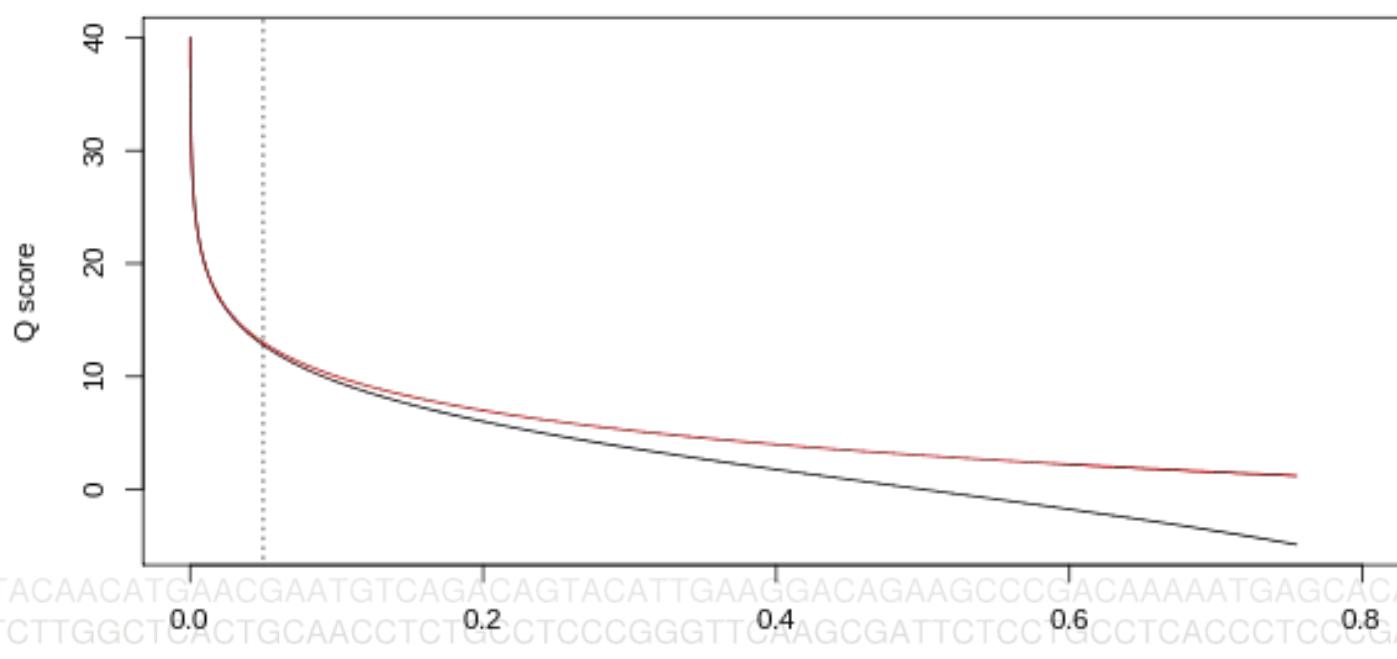
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GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTCAAGTAATCCGCCGCCTGGCCTCCAAAGTGCAGGGGTGAGGCCAC
AAAAATGGTTATGGAGATCAAAATAAGGTGGGGTGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

FASTQ quality scores

Phred quality score

$$Q_{\text{sanger}} = -10 \log_{10} p$$

Q10	1 in 10	90%
Q20	1 in 100	99%
Q30	1 in 1000	99.9%
Q40	1 in 10000	99.99%



FASTQ quality scores

Table 1 ASCII Characters Encoding Q-scores 0–40

Symbol	ASCII Code	Q-Score	Symbol	ASCII Code	Q-Score	Symbol	ASCII Code	Q-Score
!	33	0	/	47	14	=	61	28
"	34	1	0	48	15	>	62	29
#	35	2	1	49	16	?	63	30
\$	36	3	2	50	17	@	64	31
%	37	4	3	51	18	A	65	32
&	38	5	4	52	19	B	66	33
'	39	6	5	53	20	C	67	34
(40	7	6	54	21	D	68	35
)	41	8	7	55	22	E	69	36
*	42	9	8	56	23	F	70	37
+	43	10	9	57	24	G	71	38
,	44	11	:	58	25	H	72	39
-	45	12	;	59	26	I	73	40
.	46	13	<	60	27			

CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGACTCCTATCCTCAAGTAATCCGCCCGCTGGCCTCCAAAGTGCAGGGGTGAGGCCAC
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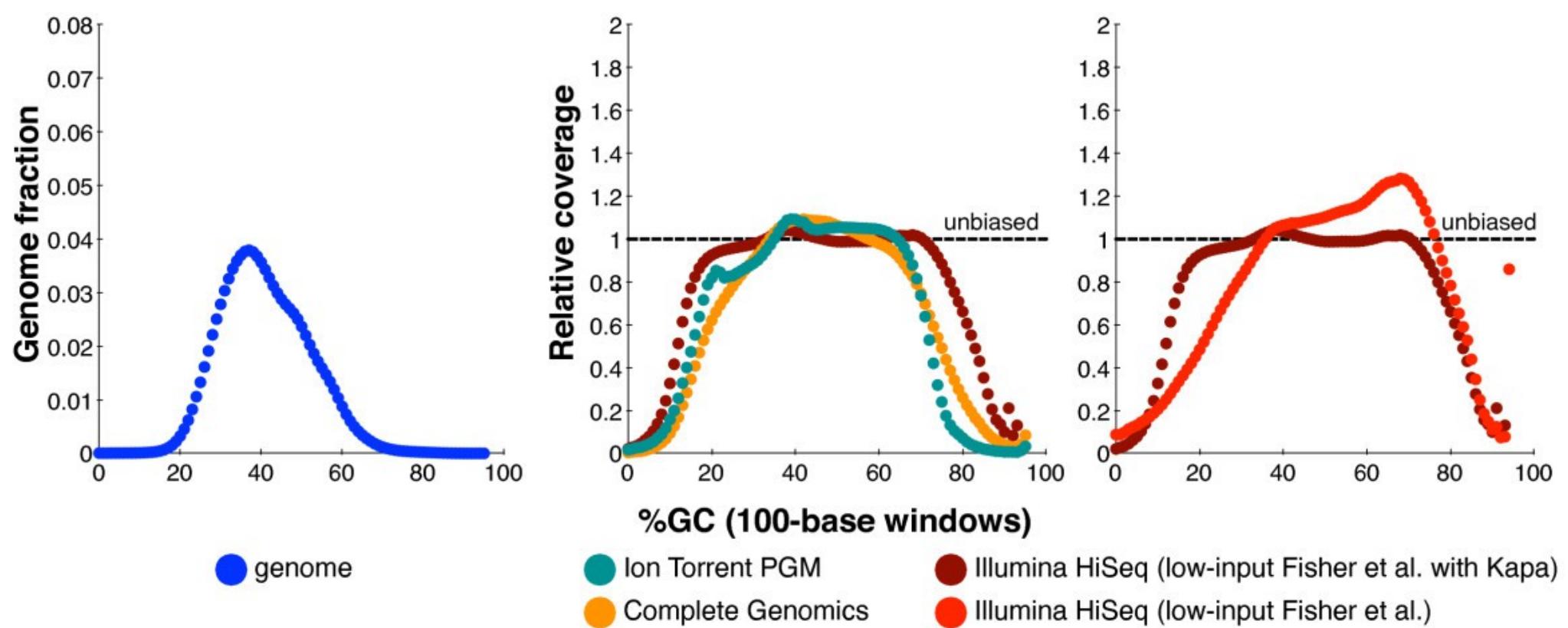
FASTQonfusion!

- Illumina:
 - CASAVA <= 1.3 Solexa
 - CASAVA 1.3 – 1.7 Illumina
 - CASAVA >= 1.8 Sanger (the “standard”)

Data quality and bias

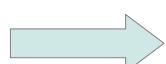
- Sequencing errors
 - Different for different techniques!
- Amplification in sample prep => duplicate reads
- GC bias
 - Sample prep
 - Bridge amplification

GC bias



Ross et al, Genome Biology, 2013

Workflow



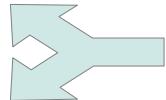
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CATGATGTGTAGGTAATGATTCTGAGACAAATTGCAATTGGTTTCATT
ATATTGGGTTGGATAAAACTGTTAACGAGATTGTCTTCCTGAAACACT
TGTCAGTAACATTAAAAACAGTACAAGACATAATAGTGCCATTGGC
ATAACTGTCAAATGAAACAATCATCAAGTGAATTGAGTTTAGTAGGAAT
CAGGGGTCTGCCTGACAGAAGTGGATTACAGGATTCTAAAAAAGCT
TTACAGATGCCAAACCACAACAATAACAAAGGCATGGATAGGGATCC
TTAAAAAGCTTATCCAATATGAATTGTTCCATATGGACCCTGTCAAGGG
GATTTCCCGGGCTTAGGAAGGGGAGGAGCGAGCAAGACAGCCTACCTTT

Identification of duplicates

Quantification

Peak calling

....



Mapping to reference

Workflow



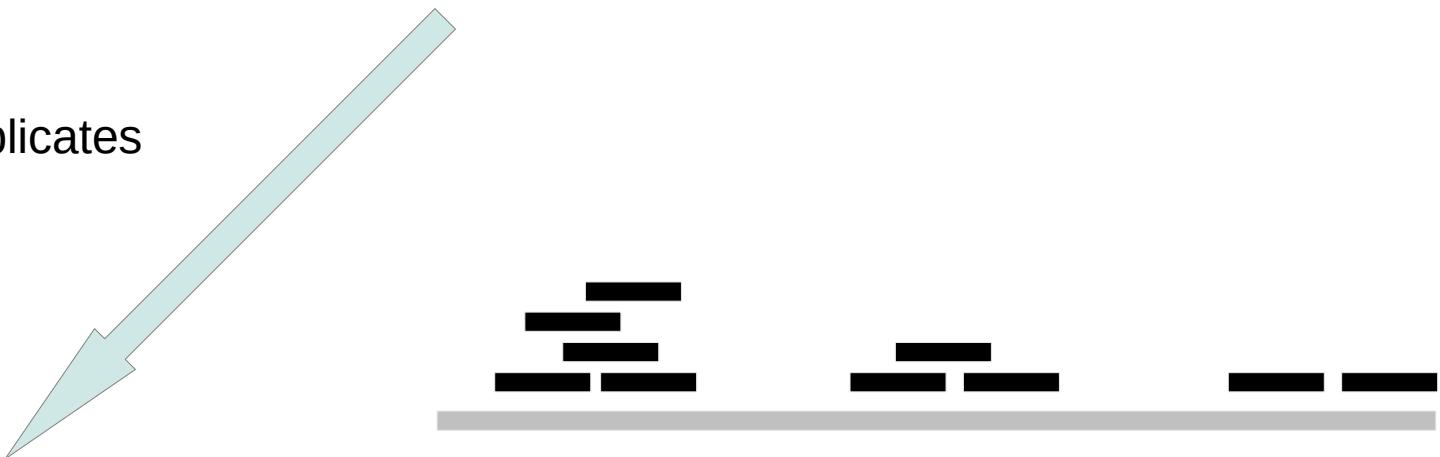
CTAGTGATTATCATCTAGGCCAGTGAATACCACTGGGTGGCAACCCCTACC
GAATGCTCGAGCGTTCATGCGAACGATCGAGCGCATTTGGCGCACGAC
CATGATGTGTAGGTAATGATTCTGAGACAAATTGCAATTGGTTTCATT
ATATTGGGTTGGATAAAACTGTTAACGAGATTGTCTTCCTGAAACACT
TGTCACTAACATTAAAAACAGTACAAGACATAATAGTGCCATTGGC
ATAACTGTCAAATGAAACAATCATCAAGTGAATTGAGTTTAGTAGGAAT
CAGGGTCTGCCTGACAGAAGTGGATTACAGGATTCTAAAAAAAGCT
TTACAGATGCCAAACCACAACAATAACAAAGGCATGGATAGGGATCC
TTAAAAAGCTTATCCAATATGAATTGTTCCATATGGACCCTGTCAAGGG
GATTTCCCGGGCTTAGGAAGGGGAGGAGCGAGCAAGACAGCCTACCTTT

Identification of duplicates

Quantification

Peak calling

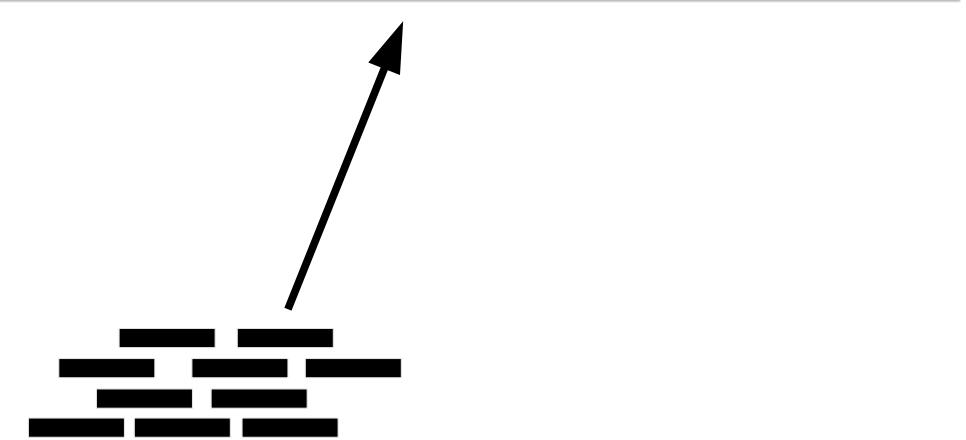
Assembly



...
CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACCTCCTCAAGTAATCCGCCGCCTGGCCTCCAAAGTGCAGGGCGTGAGGCCAC
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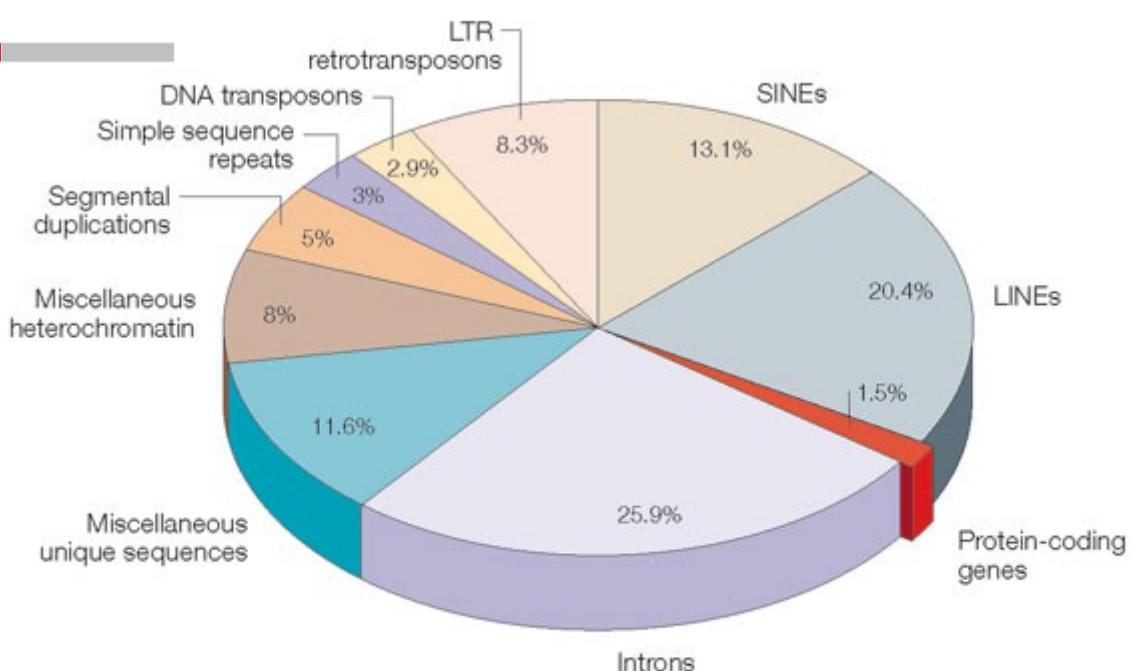
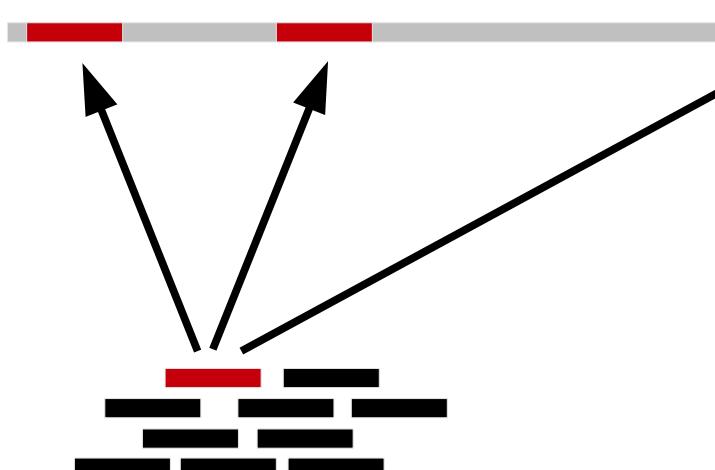
Mapping to reference

- Genome or transcriptome
 - Any other set of sequences



CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTCAAGTAATCCGCCGCCTGGCCTCCAAAGTGCAGGGGTGAGGCCAC
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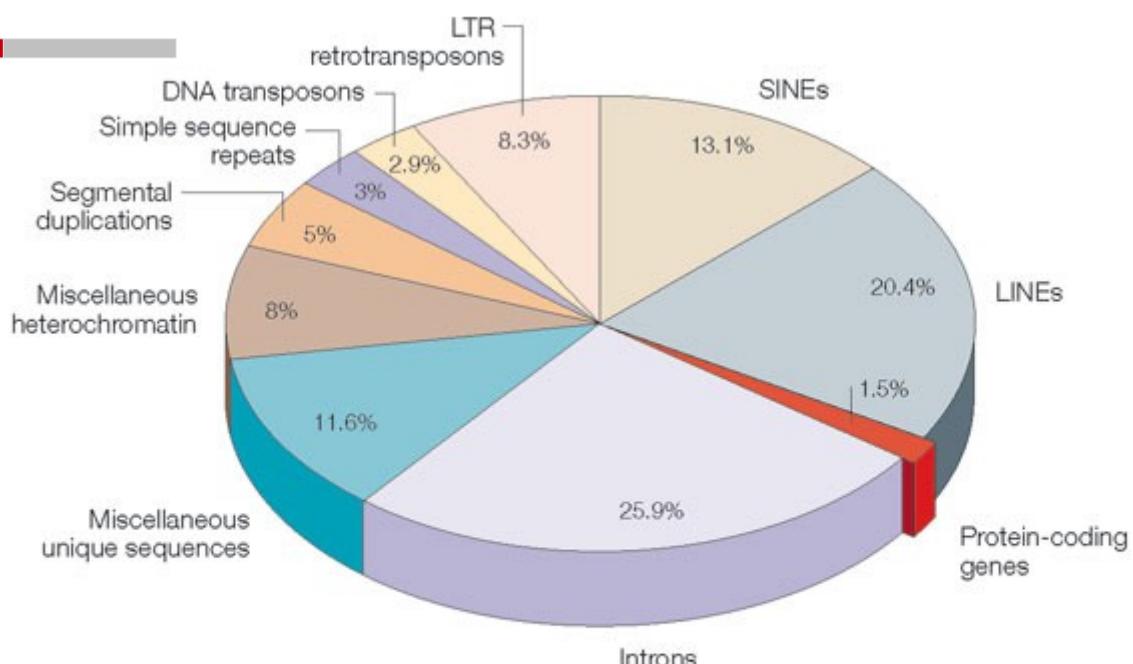
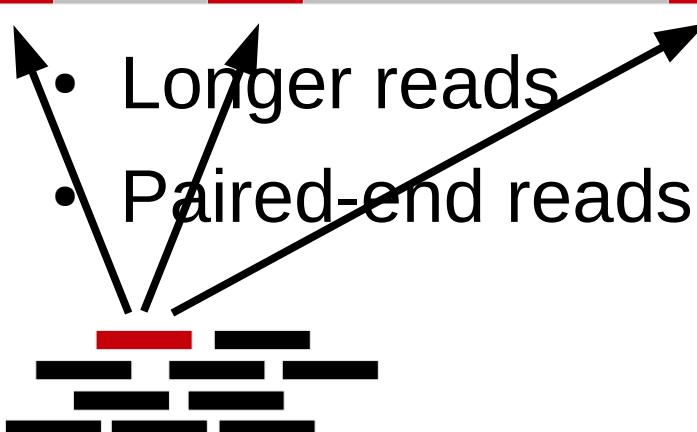
Repetitive sequence



CAAAATACTGATATATAACATGAACGAATGTCAGACAGTA
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCC
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACCTCCTAT
AAAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATL

Repetitive sequence

- How to deal with repeats during mapping?
- What can be done experimentally?

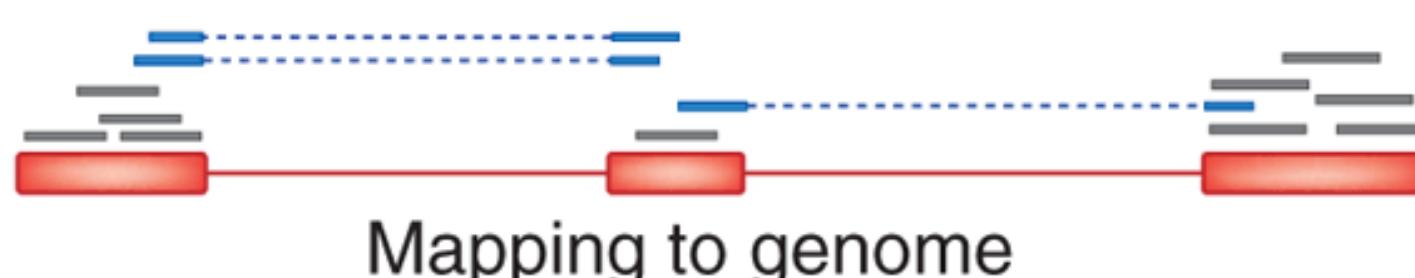
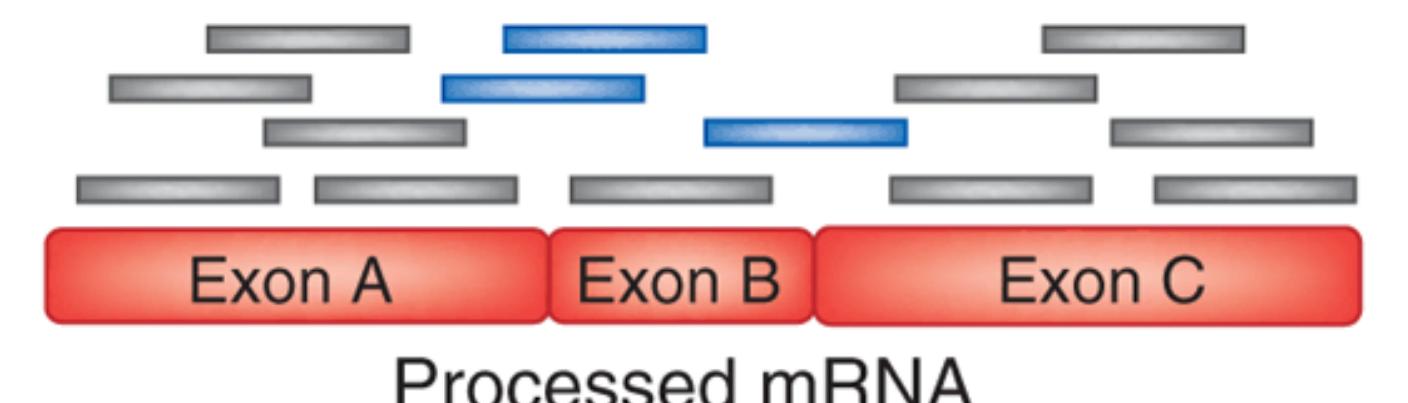


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GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTAT
AAAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATC

Difficulties

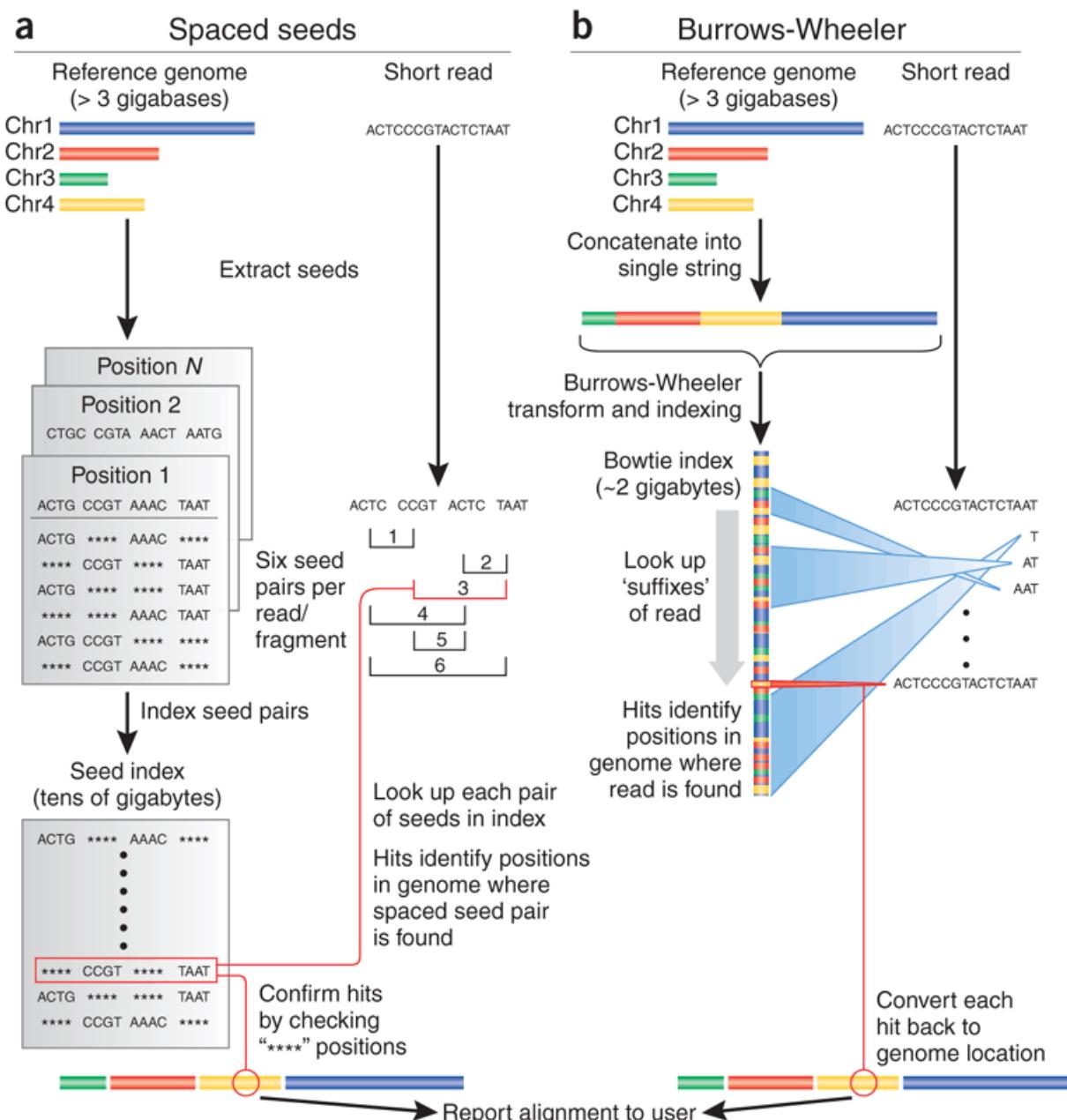
- Sequencing errors
- Errors in reference genome
- Polymorphisms
 - Insertions
 - Deletions
 - SNPs
- Spliced alignment

Mapping RNA-seq / spliced alignment

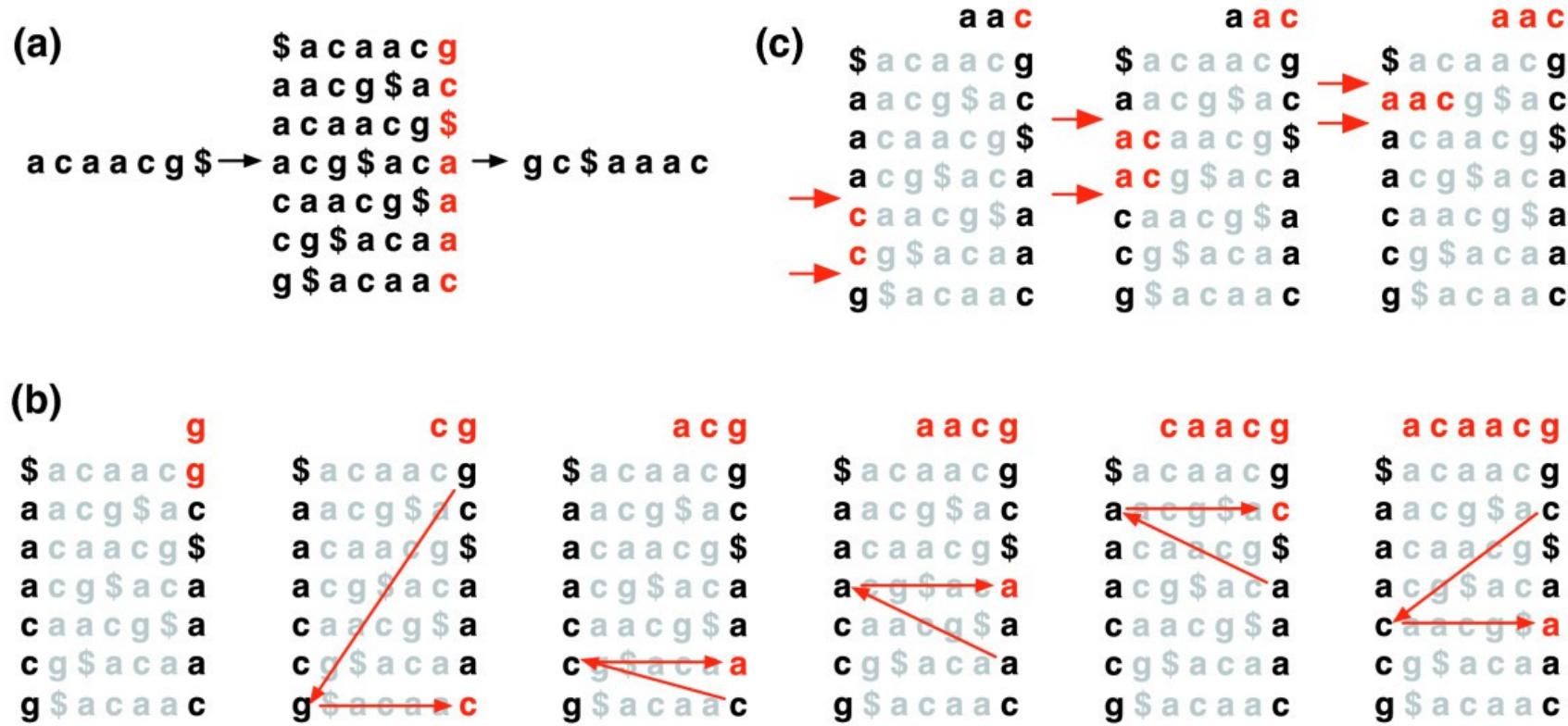


CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCCCGTGTAGTGGCACGATCTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAGGGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTCAAGTAATCCGCCCGCCTGGCCTCCAAAGTGCAGGGGTGAGGCCACAAAAATGGTTATGGAGATCAAAATAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCTG

Approaches to mapping

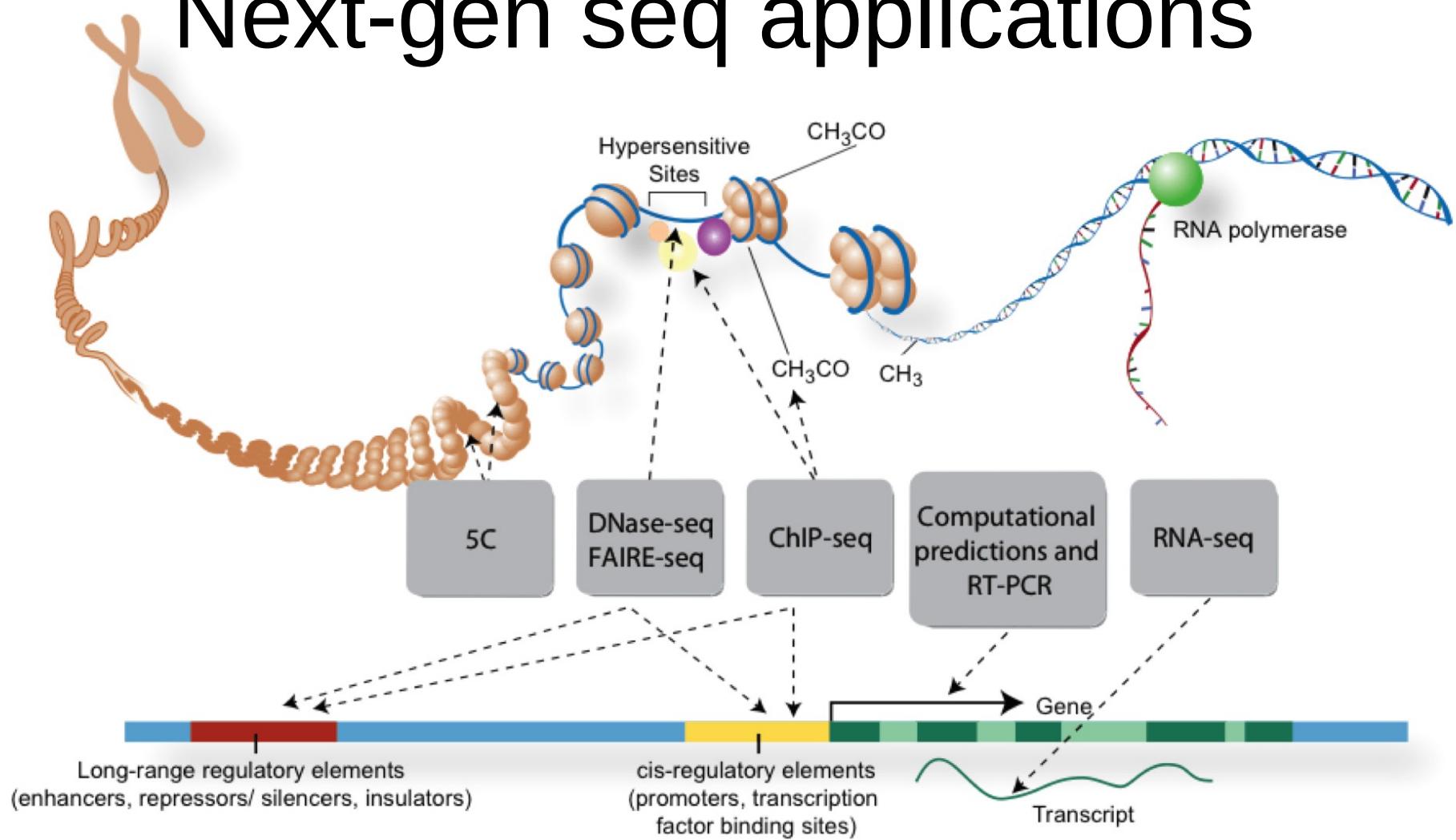


Burrows-Wheeler transform



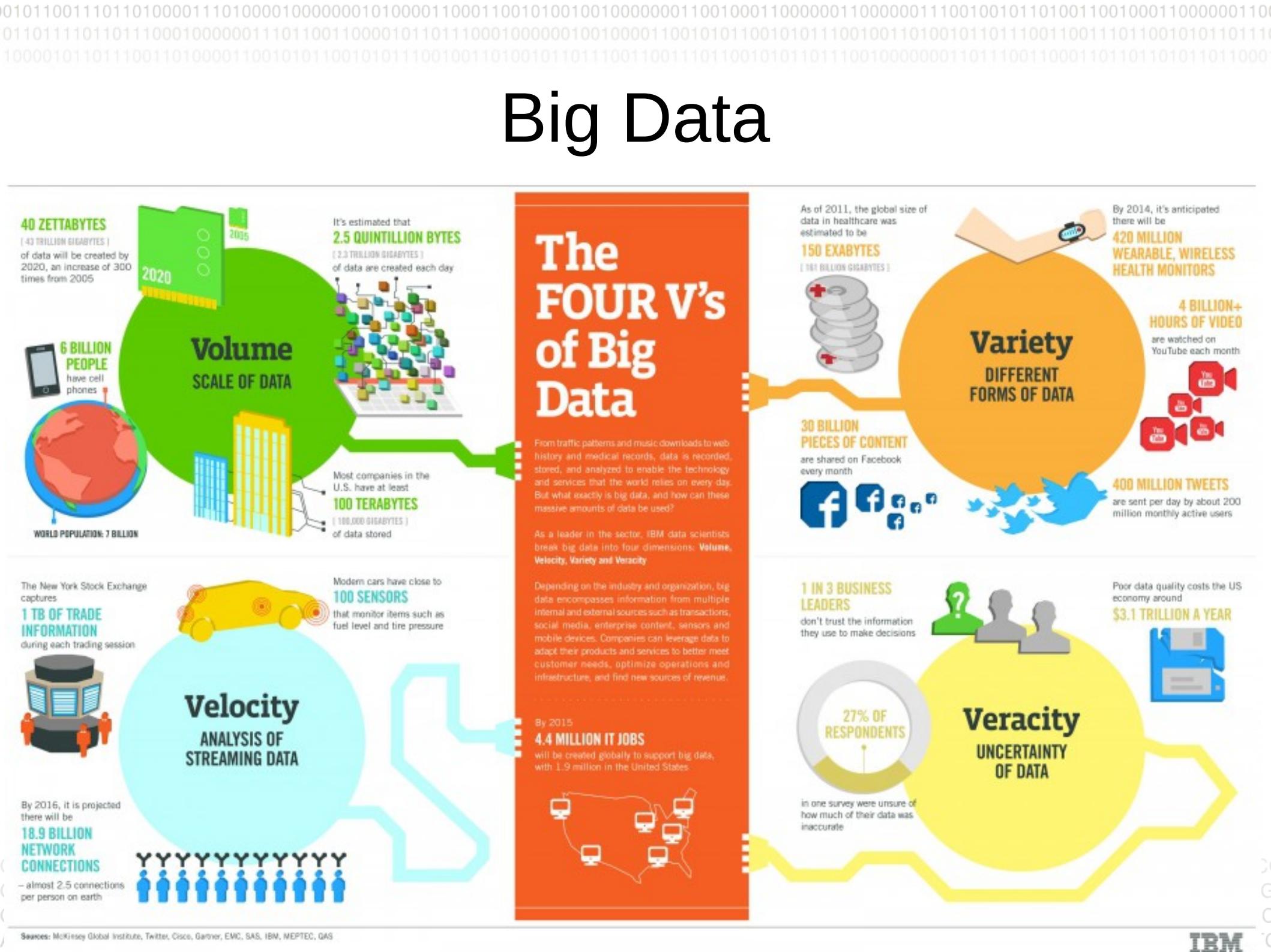
Langmead et al, 2009

Next-gen seq applications



The ENCODE Consortium

Big Data



Big Data

five

Value

The FOUR V's of Big Data

40 ZETTABYTES
(43 TRILLION GIGABYTES)
of data will be created by 2020, an increase of 300 times from 2005



The New York Stock Exchange captures
1 TB OF TRADE INFORMATION during each trading session



By 2016, it is projected there will be
18.9 BILLION NETWORK CONNECTIONS – almost 2.5 connections per person on earth

Velocity

ANALYSIS OF STREAMING DATA



It's estimated that
2.5 QUINTILLION BYTES
(2.3 TRILLION GIGABYTES) of data are created each day

Most companies in the U.S. have at least
100 TERABYTES
(100,000 GIGABYTES) of data stored

Modern cars have close to
100 SENSORS that monitor items such as fuel level and tire pressure

From traffic patterns and music downloads to web history and medical records, data is recorded, stored, and analyzed to enable the technology and services that we depend on every day. But what is the value of all this data?

By 2015
4.4 MILLION IT JOBS will be created globally to support big data, with 1.9 million in the United States.



As of 2011, the global size of data in healthcare was estimated to be

150 EXABYTES
(151 BILLION GIGABYTES)

30 BILLION PIECES OF CONTENT are shared on Facebook every month



Variety

DIFFERENT FORMS OF DATA

By 2014, it's anticipated there will be
420 MILLION WEARABLE, WIRELESS HEALTH MONITORS

4 BILLION+ HOURS OF VIDEO are watched on YouTube each month



400 MILLION TWEETS are sent per day by about 200 million monthly active users

1 IN 3 BUSINESS LEADERS don't trust the information they use to make decisions



27% OF RESPONDENTS

In one survey were unsure of how much of their data was inaccurate

Veracity

UNCERTAINTY OF DATA



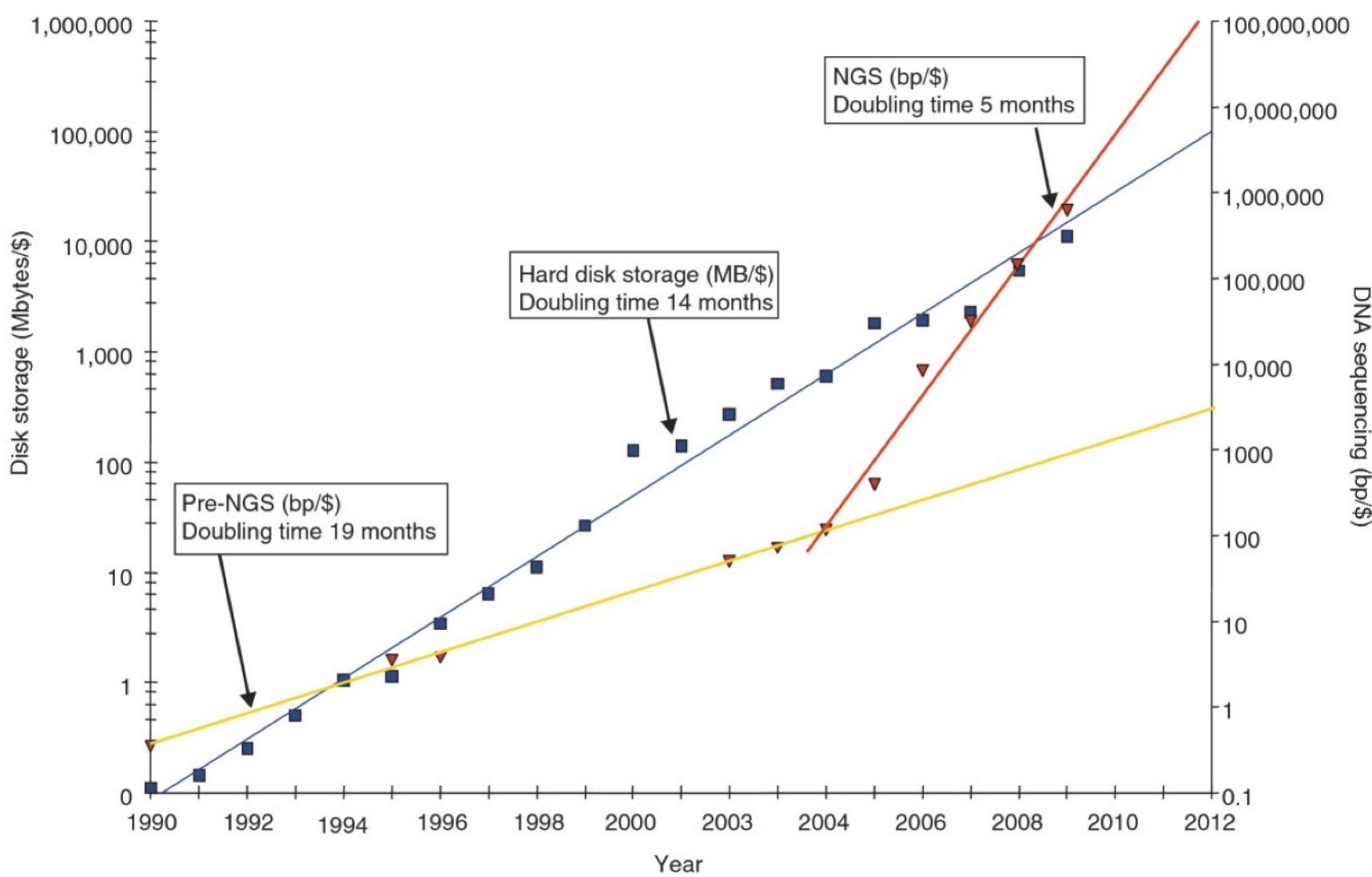
Poor data quality costs the US economy around
\$3.1 TRILLION A YEAR

IBM

Challenges

- Storage
 - From terabytes to petabytes
- Analysis
 - Computational resources
 - Memory
 - CPU
- Sharing
 - Bandwidth

NextGen Sequencing a Game-Changer



Lincoln Stein (via C. Titus Brown)

Opportunities

- A wealth of data in public databases
- Computational analysis:
 - Reproducible
 - Not dependent on lab / materials
- Cloud-based analysis
 - Amazon AWS
 - National HPC and e-Science resources
 - The Netherlands: SURFsara

ARTICLE

Received 9 Apr 2014 | Accepted 25 Jun 2014 | Published 24 Jul 2014

DOI: 10.1038/ncomms5498

OPEN

A highly abundant bacteriophage discovered in the unknown sequences of human faecal metagenomes

Bas E. Dutilh^{1,2,3,4}, Noriko Cassman^{3,†}, Katelyn McNair², Savannah E. Sanchez³, Genivaldo G.Z. Silva⁵, Lance Boling³, Jeremy J. Barr³, Daan R. Speth⁶, Victor Seguritan³, Ramy K. Aziz^{2,7}, Ben Felts⁸, Elizabeth A. Dinsdale^{3,5}, John L. Mokili³ & Robert A. Edwards^{2,4,5,9}

Research

Highly accessed

Open Access

DNA methylation age of human tissues and cell types

Steve Horvath

Correspondence: Steve Horvath shorvath@mednet.ucla.edu

▼ Author Affiliations

Human Genetics, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, CA 90095, USA

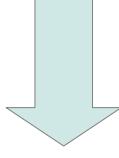
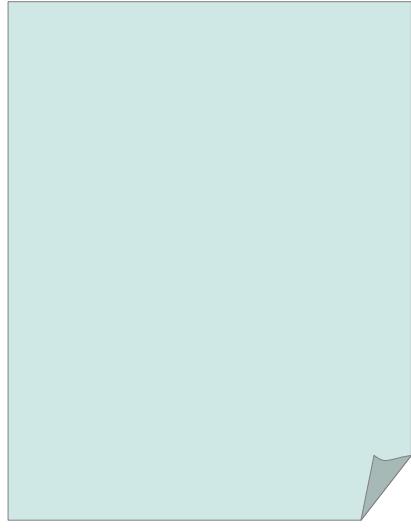
Biostatistics, School of Public Health, University of California Los Angeles, Los Angeles, CA 90095, USA

Human Genetics, Gonda Research Center, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, CA 90095-7088, USA

AATAGCTGGGATTACAG
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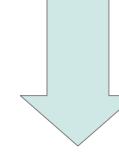
Bioinformatic analysis in a nutshell

“Do something”



Mostly text:

- Sequences
- Genomic coordinates
- Etc.



Mostly text:

- Sequences
- Genomic coordinates
- Etc.

A side note...

Bioinformatician? Computational
Biologist? Data analyst? Data
curator? Database developer?
Statistician? Mathematical
Modeler? Software Developer?
Ontologist? Programmer?

So you want to be a computational biologist? Loman & Watson, 2013

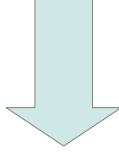
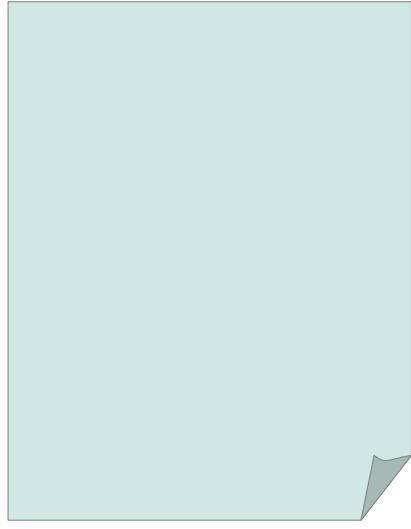


Computational Biology

- It's about the biology
- It's research
- The computer is just the tool used to answer interesting questions
- Iterative, collaborative process between wet-lab and dry-lab

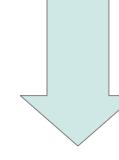
Bioinformatic analysis in a nutshell

“Do something”



Mostly text:

- Sequences
- Genomic coordinates
- Etc.



Mostly text:

- Sequences
- Genomic coordinates
- Etc.

185,750

46,485,800

46,485,850

46,485,900

46,485,950

46,486,000

46,486,050

46,486,100

Homo sapiens (Gene)
Gene annotations (3,048)

Predicted Binding Sites SRF
Binding site annotations (89)

49
SRF (Read coverage)
Graph

0
0
SRF
1,064,027 reads

ChIP peak

RAD54L

36
24
SRF background (Read coverage)
Graph

0
0
SRF background
1,407,191 reads

Predicted Bin... X

Chromosome	Region	Name	p-value	Score	FDR	note
chr1	45224888..45224947	ChIP peak	0.00	0.00	4.43E-108	# forward reads : 196, # reverse reads : 222, Region containing reads : 45224467..45225
chr1	45578365..45578433	ChIP peak	3.82E-6	3.82E-6	3.63E-6	# forward reads : 43, # reverse reads : 63, Region containing reads : 45577983..4557871
chr1	46024004..46024099	ChIP peak	9.20E-6	9.20E-6	6.18E-4	# forward reads : 18, # reverse reads : 13, Region containing reads : 46023761..4602445
chr1	46485912..46485968	ChIP peak	5.02E-14	5.02E-14	1.54E-34	# forward reads : 85, # reverse reads : 80, Region containing reads : 46485517..4648639
chr1	46855203..46855264	ChIP peak	0.00	0.00	3.36E-86	# forward reads : 162, # reverse reads : 160, Region containing reads : 46854793..46855
chr1	51539515..51539583	ChIP peak	4.56E-5	4.56E-5	4.10E-22	# forward reads : 53, # reverse reads : 44, Region containing reads : 51539124..5153995

► NCBI BLAST Home

BLAST finds regions of similarity between biological sequences. [more...](#)

New

DELTABLAST, a more sensitive protein-protein search

Go

BLAST Assembled RefSeq Genomes

Choose a species genome to search, or [list all genomic BLAST databases](#).

- [Human](#)
- [Mouse](#)
- [Rat](#)
- [Cow](#)
- [Pig](#)
- [Dog](#)
- [Rabbit](#)
- [Chimp](#)
- [Guinea pig](#)
- [Sheep](#)

- [Fruit fly](#)
- [Honey bee](#)
- [Chicken](#)
- [Zebrafish](#)
- [Clawed frog](#)

- [Arabidopsis](#)
- [Rice](#)
- [Yeast](#)
- [Neurospora crassa](#)
- [Microbes](#)

Basic BLAST

Choose a BLAST program to run.

nucleotide blast Search a nucleotide database using a nucleotide query <i>Algorithms: blastn, megablast, discontiguous megablast</i>	protein blast Search protein database using a protein query <i>Algorithms: blastp, psi-blast, phi-blast, delta-blast</i>
blastx Search protein database using a translated nucleotide query	tblastn Search translated nucleotide database using a protein query
tblastx Search translated nucleotide database using a translated nucleotide query	

Specialized BLAST

Choose a type of specialized search (or database name in parentheses.)

- Make specific primers with [Primer-BLAST](#)
- Search [trace archives](#)
- Find [conserved domains](#) in your sequence (cds)
- Find sequences with similar [conserved domain architecture](#) (cdart)
- Search sequences that have [gene expression profiles](#) (GEO)
- Search [immunoglobulins and T cell receptor sequences](#) (IgBLAST)
- Screen sequence for [vector contamination](#) (vecscren)
- Align two (or more) sequences using BLAST (bl2seq)
- Search [protein](#) or [nucleotide](#) targets in PubChem BioAssay
- Search [SRA by experiment](#)

Your Recent Results [New!](#)[All Recent results...](#)

News

[New gap costs available for PAM30 and PAM70](#)

The BLAST webpage now offers additional, more stringent, gap costs for PAM30 and PAM70.

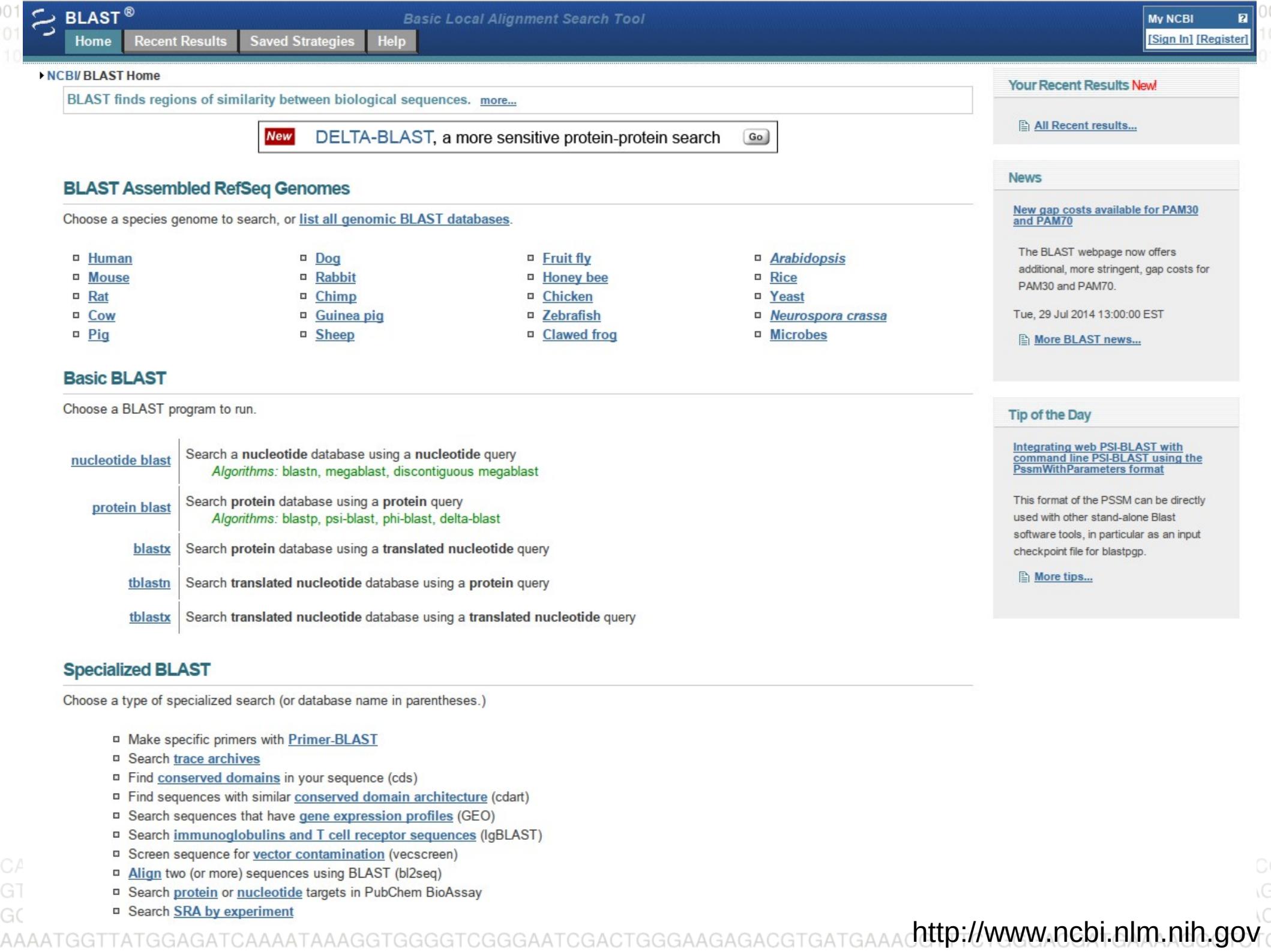
Tue, 29 Jul 2014 13:00:00 EST

[More BLAST news...](#)

Tip of the Day

[Integrating web PSI-BLAST with command line PSI-BLAST using the PssmWithParameters format](#)

This format of the PSSM can be directly used with other stand-alone Blast software tools, in particular as an input checkpoint file for blastpgp.

[More tips...](#)

Data intensive biology *for everyone.*

Galaxy is an open, web-based platform for data intensive biomedical research. Whether on the free public server or your own instance, you can perform, reproduce, and share complete analyses.

Use Galaxy



Use project's free server or other public servers

Get Galaxy



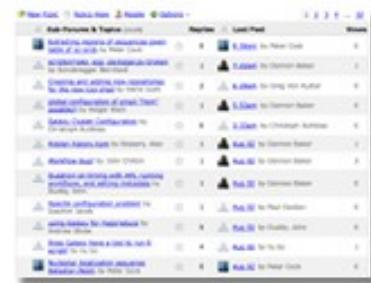
Install locally or in the cloud or get Galaxy on SlipStream

Learn Galaxy



Screencasts, Galaxy 101, ...

Get Involved



Mailing lists, Tool Shed, wiki

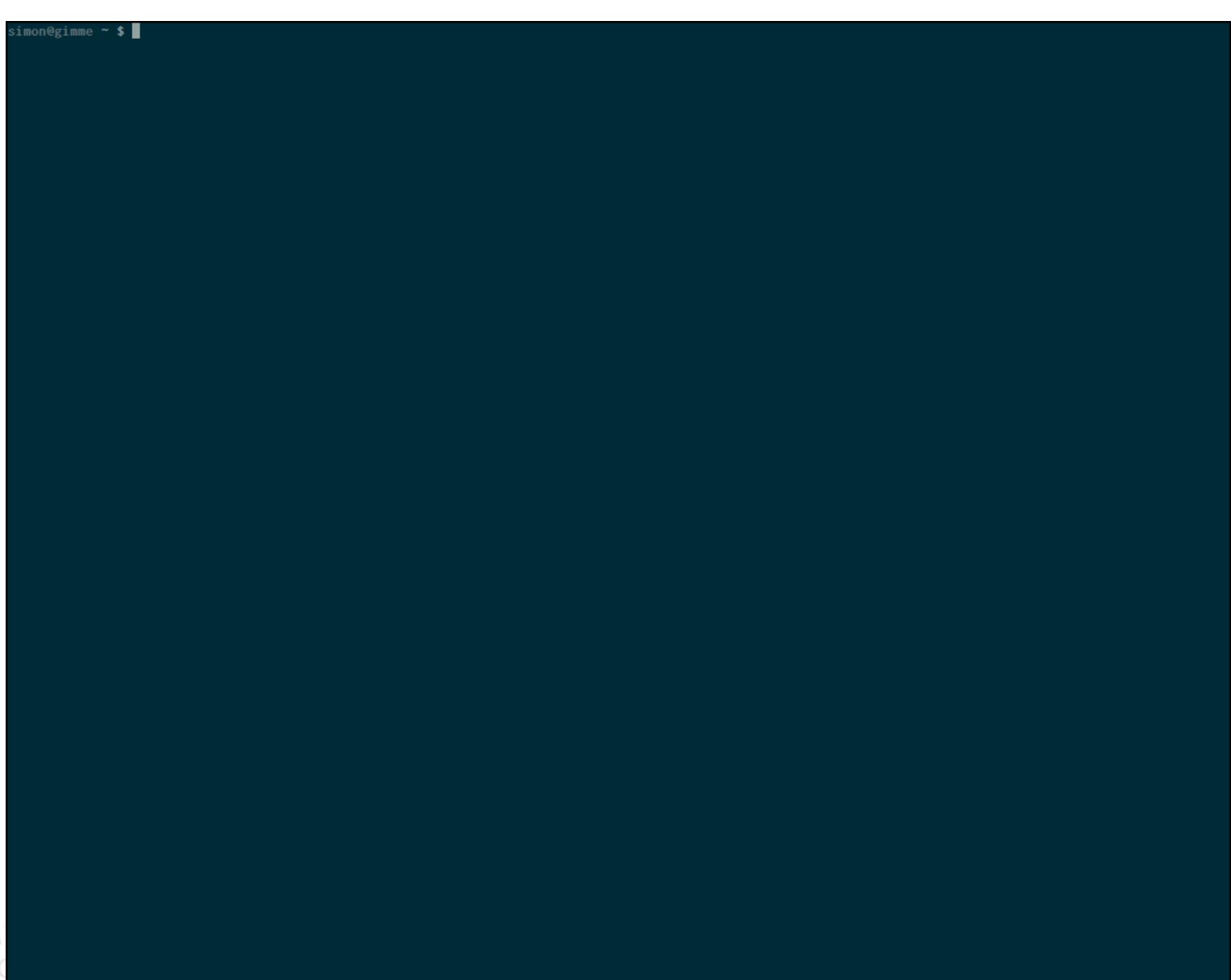
Search all resources

The Galaxy Team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.

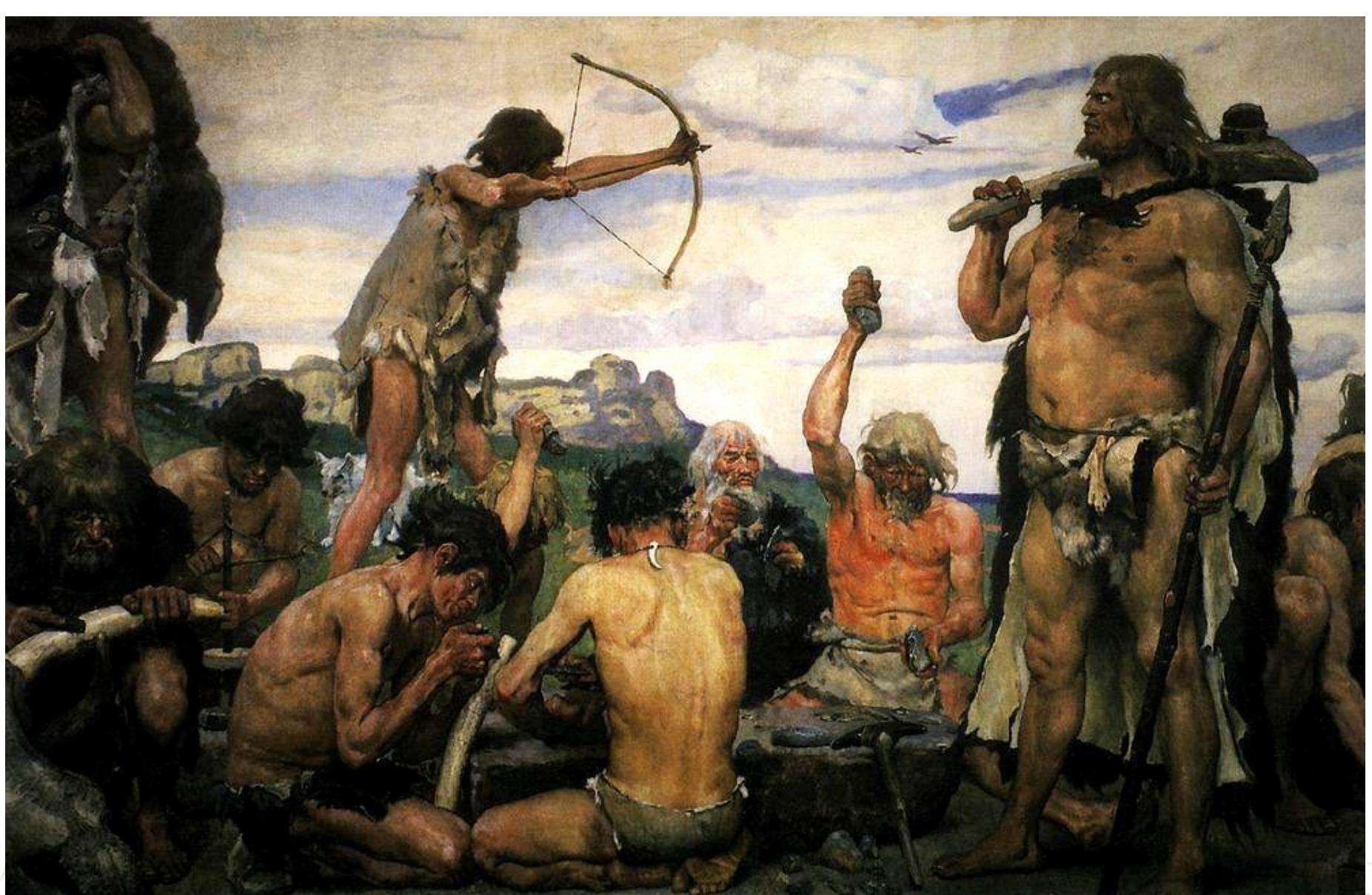
<http://galaxyproject.org>

An alternative

The command line



Why?



Viktor M. Vasnetsov - Wikipedia

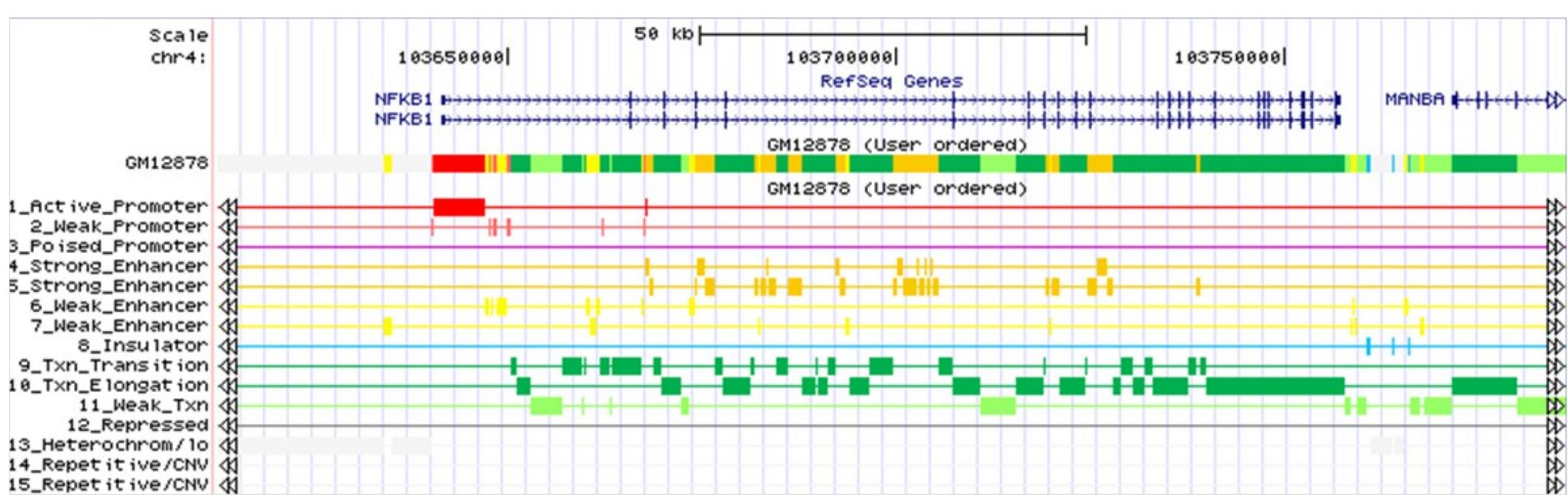
The command line

- Powerful
- Great control over what you're doing
- Run multiple jobs
 - at once
 - hundreds of 'em!
- Many computational tools don't have a GUI
- Reproducible, reusable research
 - (In theory...)

The command line



An example

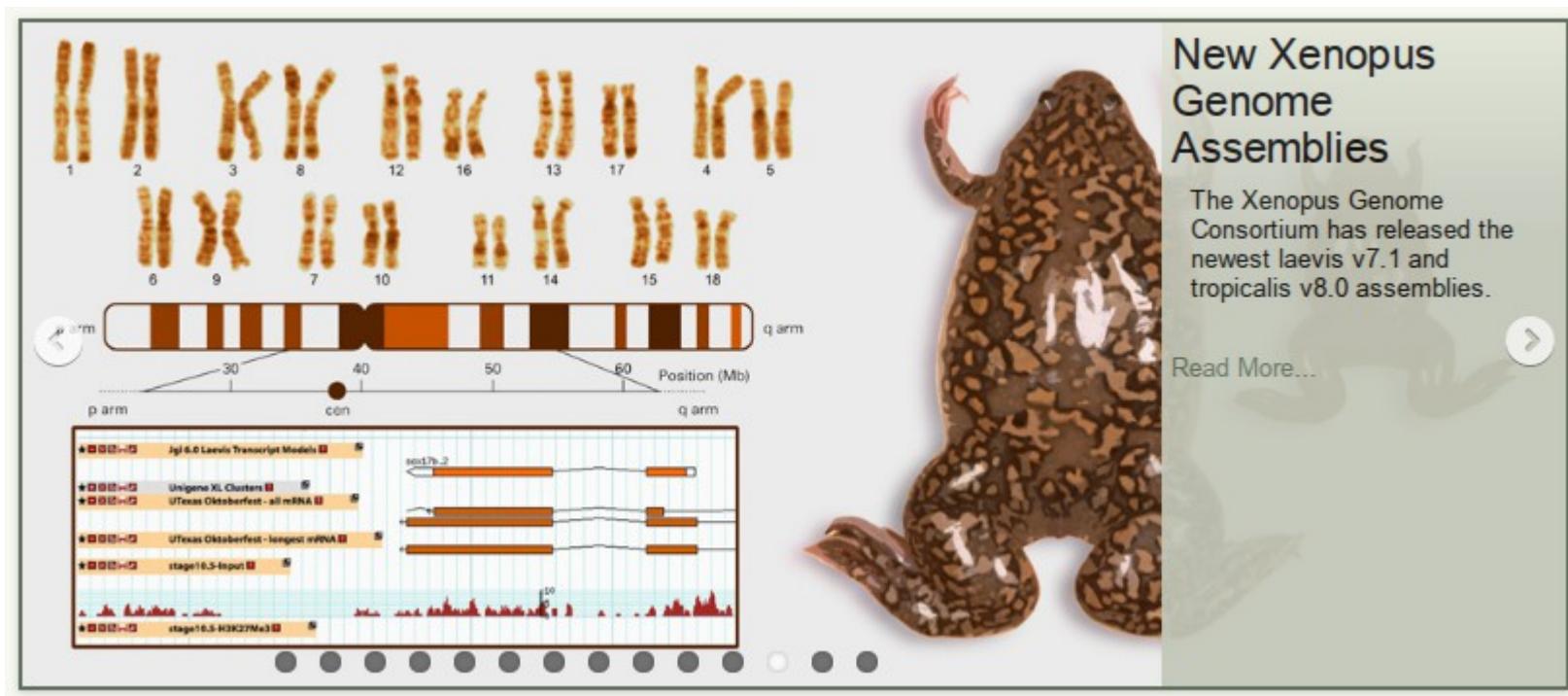


ChromHMM, Ernst & Kellis, 2012

CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
GGGGATTCAACCACGTTGGCCACGCTGGTCTGGAACTCCTCAAGTAATCCGCCCGCTCGGCCTCCAAAGTGCAGGCGTGAGCCAC
AAAAATGGTTATGGAGATCAAAATAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

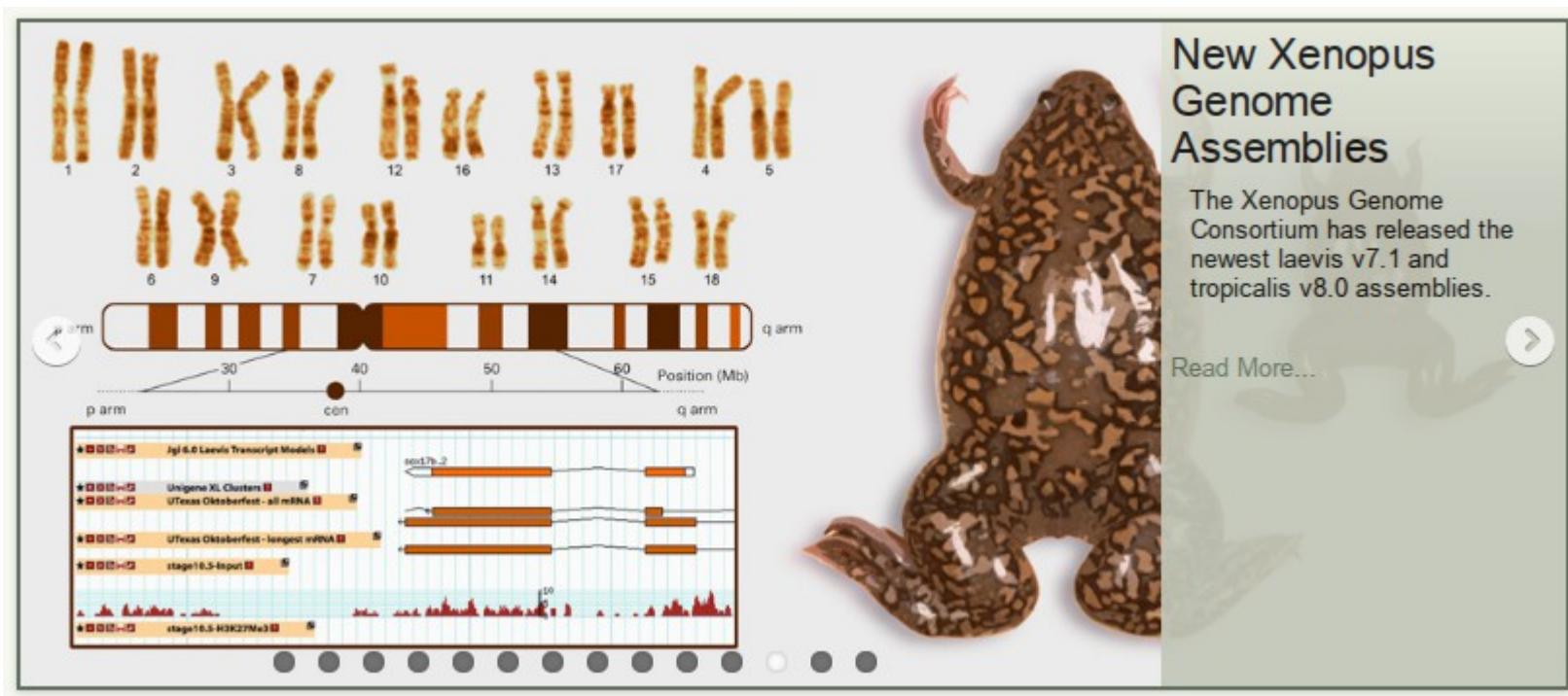
An example

- Studying development in *Xenopus tropicalis*
- 10 different assays in 5 different stages of development
- Next-gen sequencing data
- Analysis steps:
 - 1) Mapping + “peak-calling”
 - 2) Combine data in ChromHMM and learn model
 - 3) Run analyses and make figures



- Analysis steps:
 - 1) Mapping + “peak-calling”
 - 2) Combine data in ChromHMM and learn model
 - 3) Run analyses and make figures

CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
 GTGTAGTGGCACGATCTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
 GGGGATTCAACCACGTTGGCCACGCTGGACTCCTATCCTCAAGTAATCCGCCGCTCGGCCTCCAAAGTGCAGGGGTGAGGCCAC
 AAAATGGTTATGGAGATCAAAATAAGGTGGGGTGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

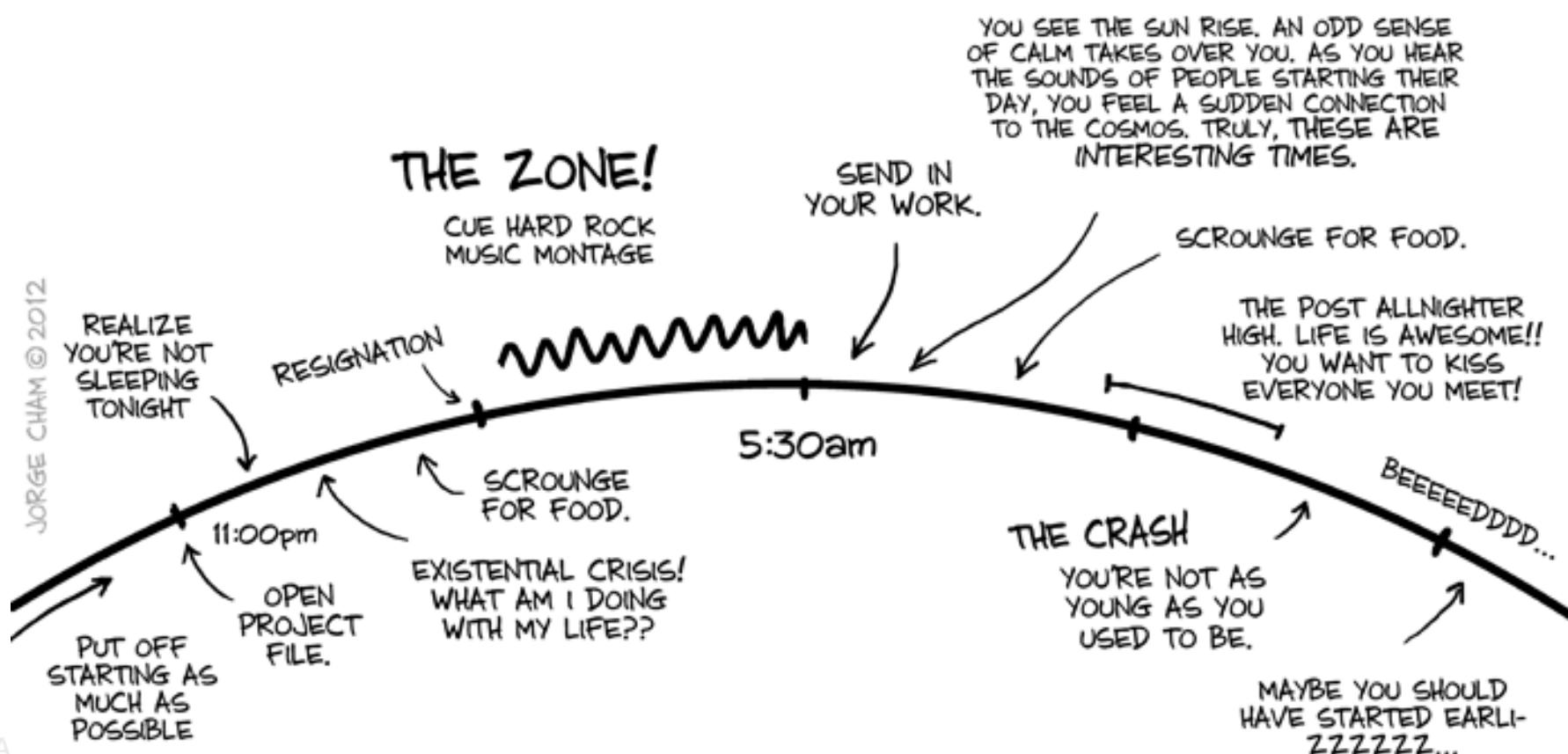


- Analysis steps:
 - 1) **Mapping** + “peak-calling”
 - 2) Combine data in ChromHMM and learn model
 - 3) Run analyses and make figures

CAAAATACTGATATATAACATGAACGAATGTCAGACAGTACATTGAAGGGACAGAAGGCCGACAAAAATGAGCACATAATGTATGATTCCCC
 GTGTAGTGGCACGATCTGGCTCACTGCAACCTCTGCCTCCGGTTCAAGCGATTCTCCTGCCTCACCCCTCCGAATAGCTGGGATTACAG
 GGGGATTCAACCACGTTGGCCACGCTGGACTCCTATCCTCAAGTAATCCGCCGCTGGCCTCCAAAGTGCAGGCGTGAGCCAC
 AAAATGGTTATGGAGATCAAAATAAGGTGGGGTGGGAATCGACTGGGAAGAGACGTGATGAAACGTTCTGGGACGATGAAAAGGGTCT

That's one unhappy PhD student..

THE ALLNIGHTER



CAAAATA
GTGTAGT
GGGGAT
AAAAATGG

WWW.PHDCOMICS.COM

GATTCCCC
GATTACAG
TGAGCCAG
AGGGTCTG

Instead...

```
$ sed -i 's/JGI_7.1/JGI_8.0/' config.txt  
  
$ ./run_analysis.sh config.txt
```

- Change configuration file
- Start script
- ~~Go home and watch Netflix~~ Continue with new, exciting analysis

Other advantages

- Load-whole-file versus streaming
- Data doesn't always all fit into memory
- A lot of biological data is just text
- Can be processed line by line

General considerations

- Understand your goals
- Step-by-step, don't try to do it all at once
- Try to break your own scripts
- Choose appropriate methods and tools

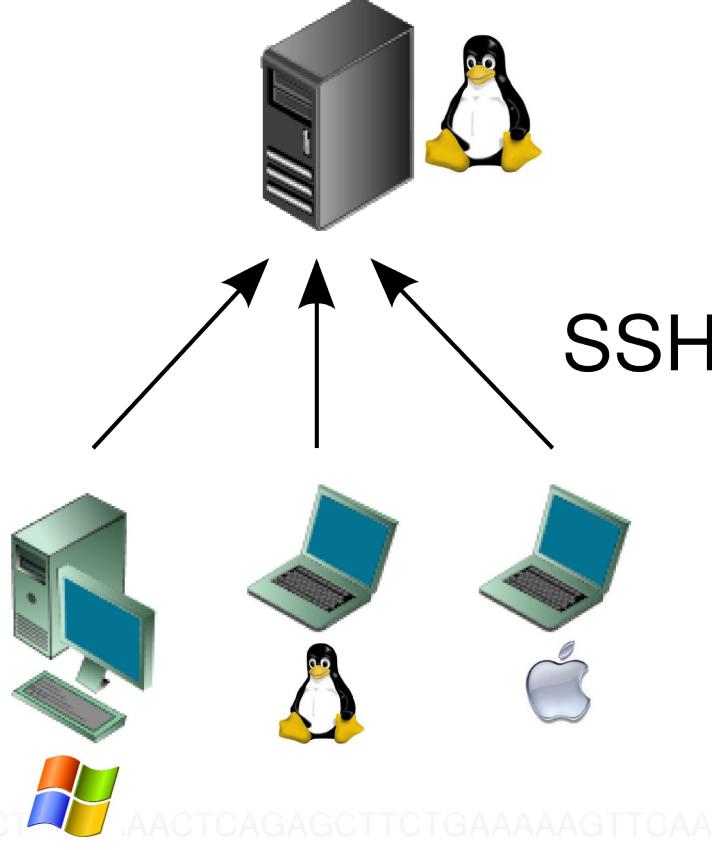
TRUST NO ONE

Today

- Familiarize yourself with the Linux command line
- Next-gen file formats
 - FASTQ
 - BAM
- Mapping (?)

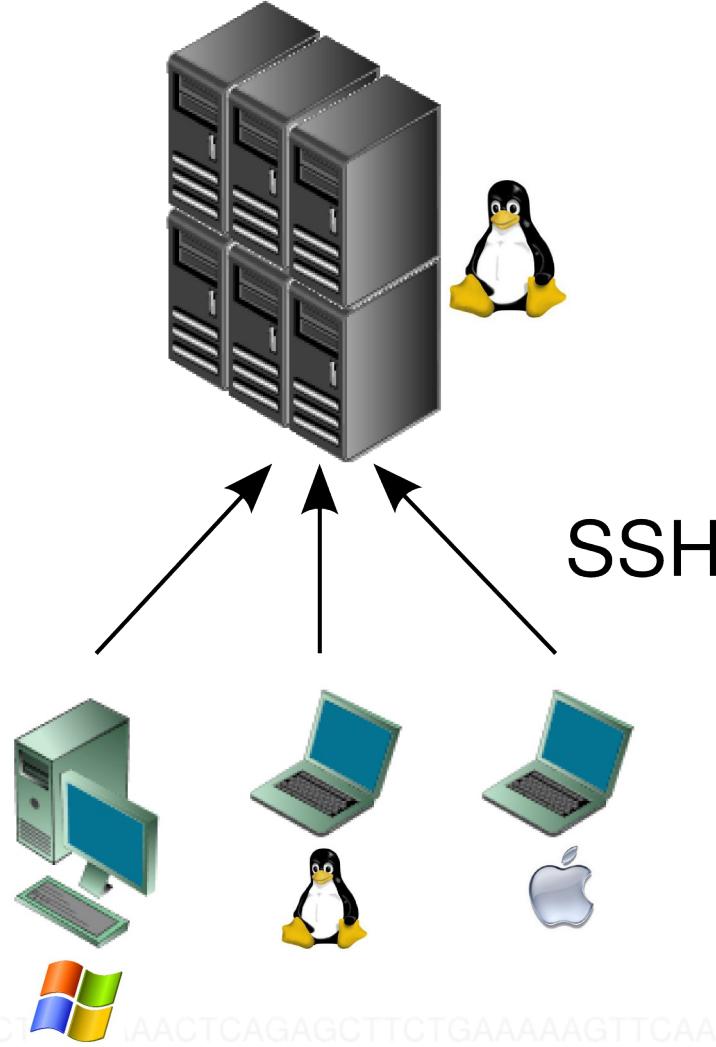
SSH (Secure Shell)

- Connect to server
- Clients available for every OS
- Perform analysis remotely

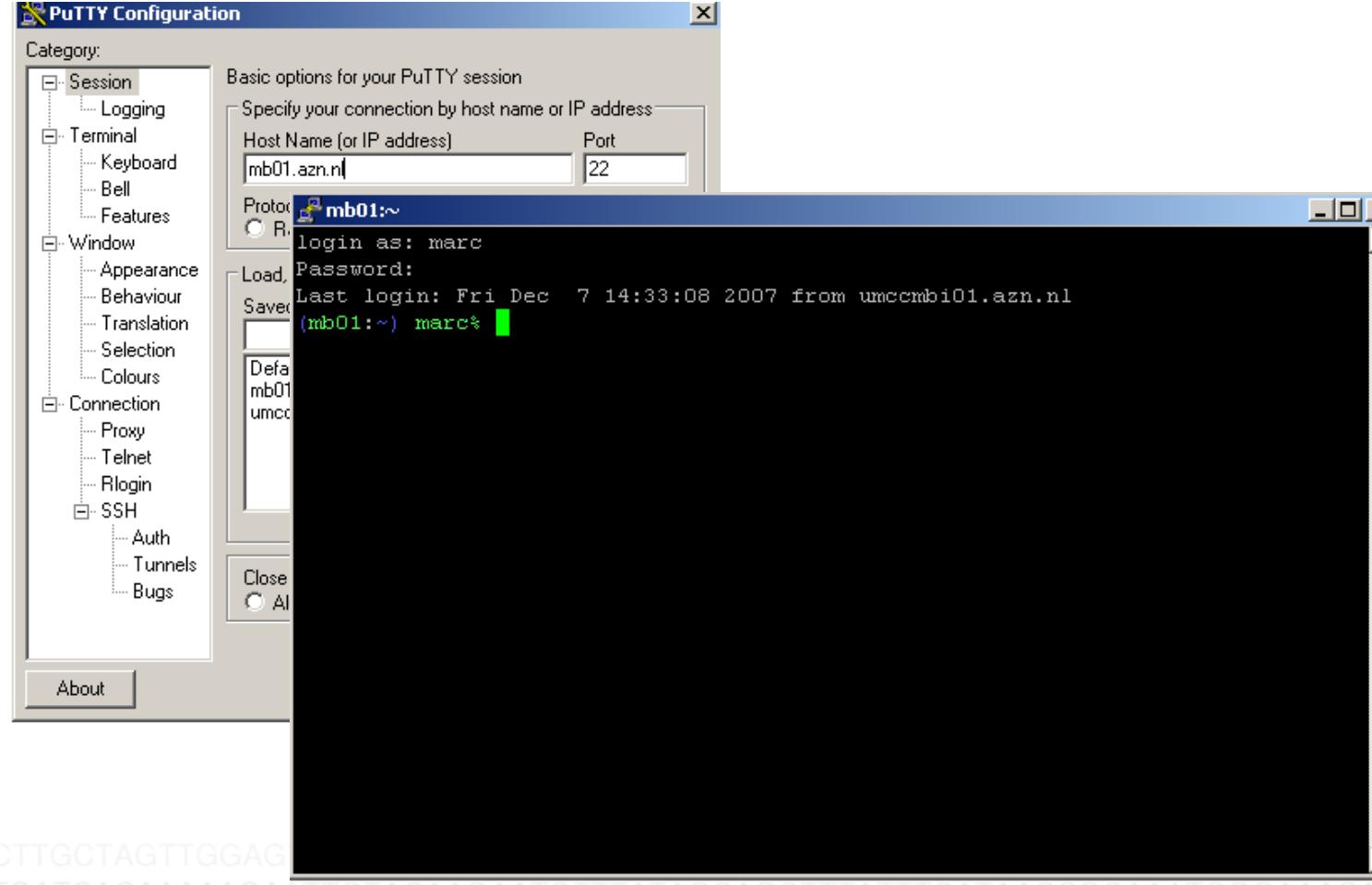


SSH (Secure Shell)

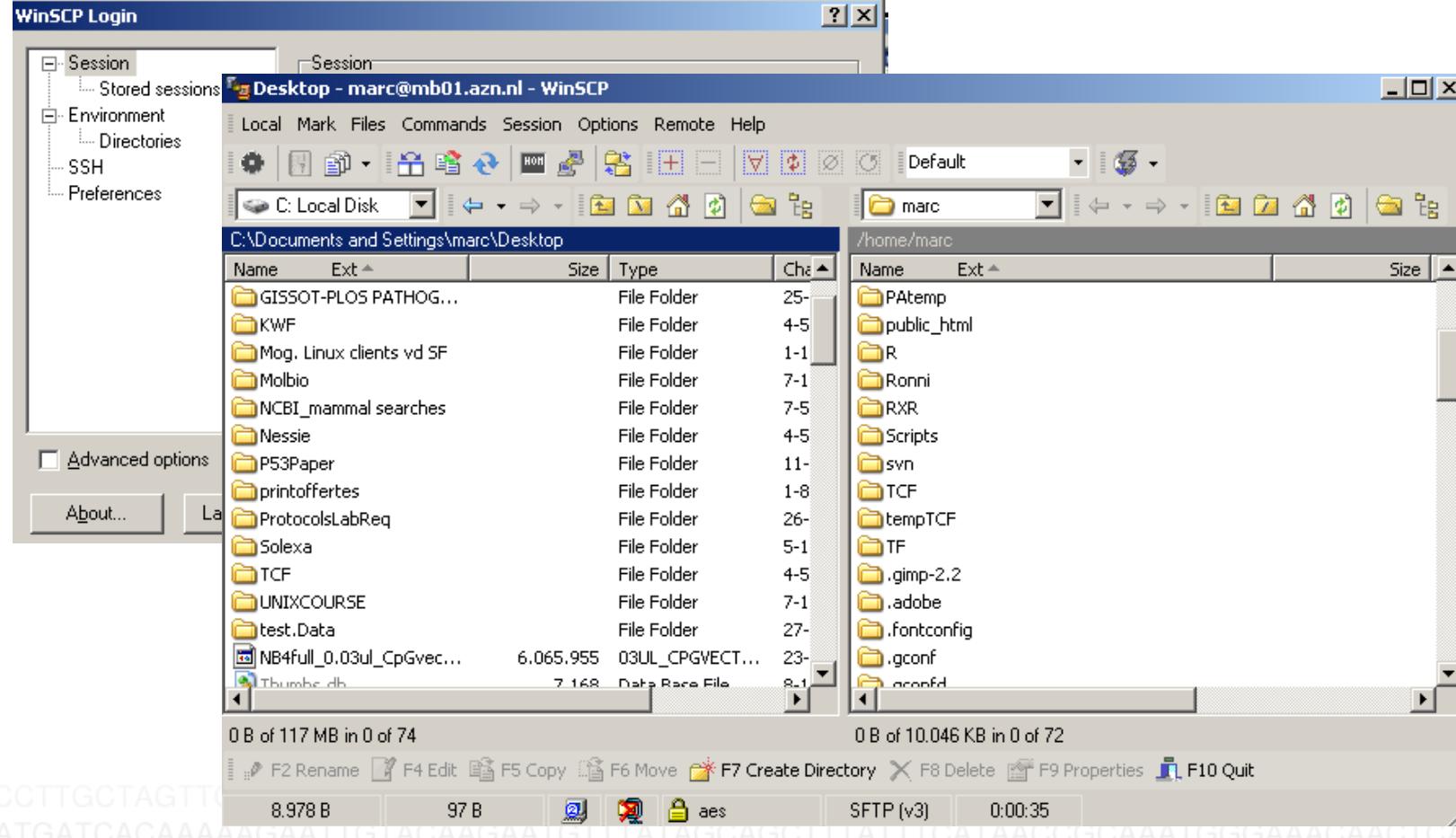
- Connect to server
- Clients available for every OS
- Perform analysis remotely
- Server can have lots of memory and CPU power



For Windows: Putty



For Windows: WinSCP



Server IP addresses

- 23.20.162.10
- 54.80.42.82
- 23.20.67.155
- 54.198.43.176
- 54.81.15.78
- 54.242.170.57
- 54.80.155.251