

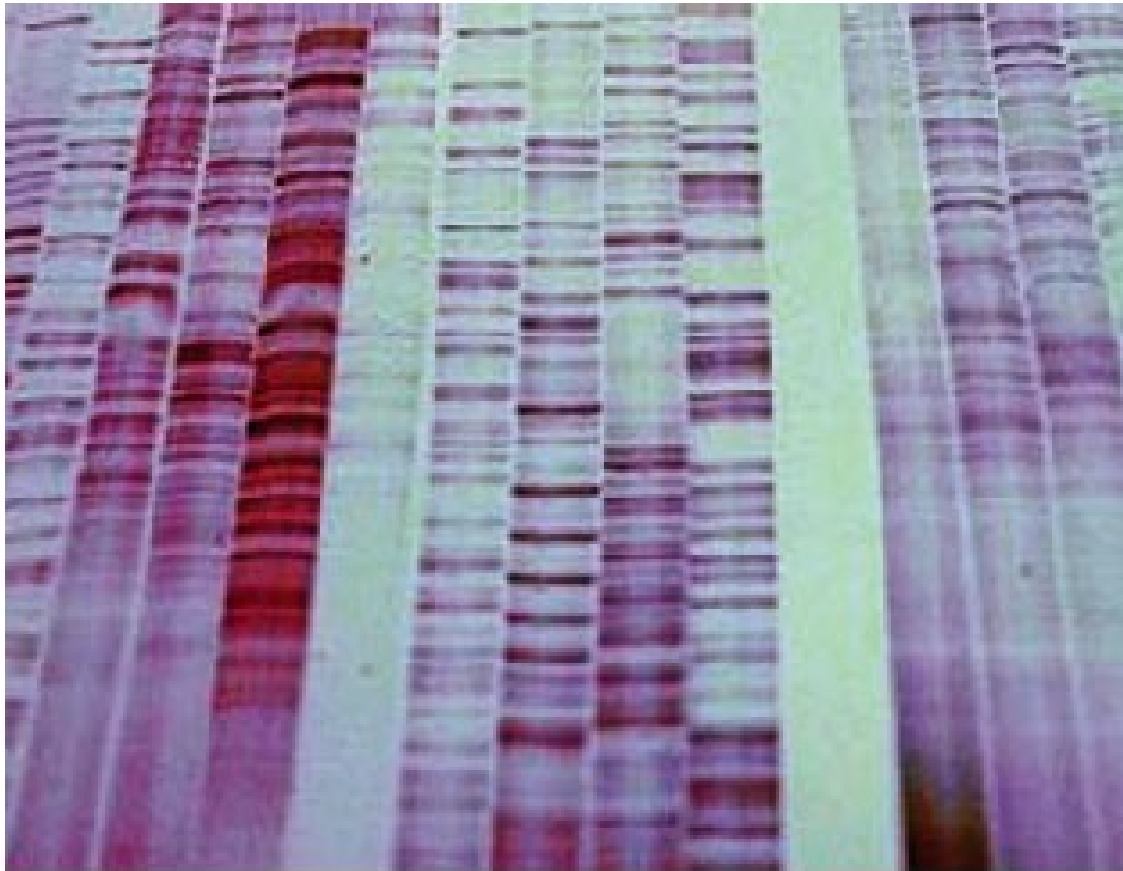
Introduction to next-generation sequencing

Simon van Heeringen
November 3, 2014

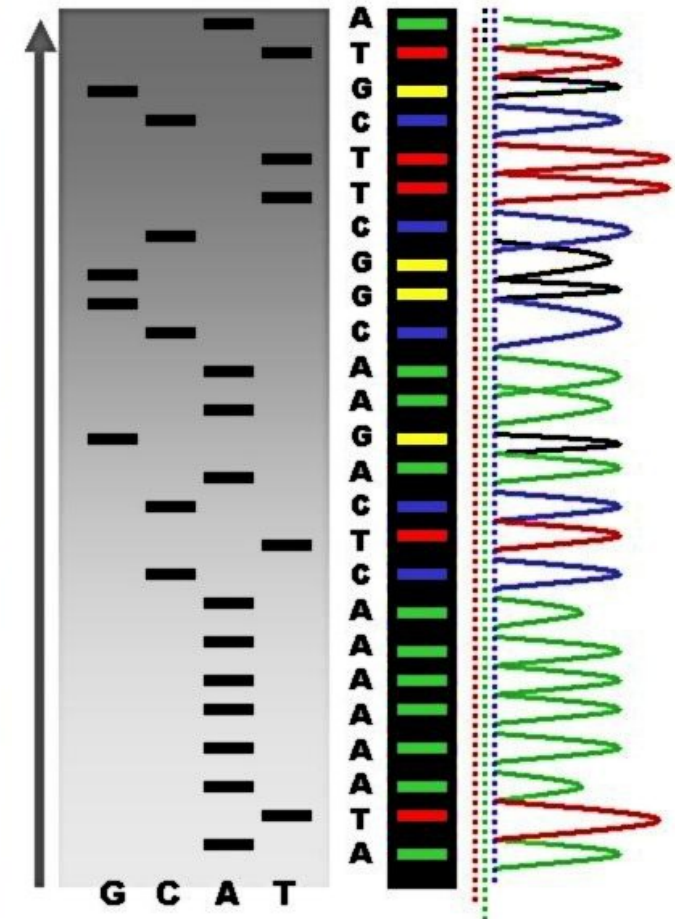
So, you want to do sequencing...?

3'Seq 3-seq 3P-seq AHT-ChIP-seq ARS-seq
ATAC-seq BOINC-seq BS-seq Bar-seq BisChIP-
seq Bru-seq Bubble-seq CAB-seq CAGE-seq
CHART-seq CLASH-seq CNV-seq CRE-seq
Capture-C-seq Cel-seq ChIA-PET-seq ChIP-seq
ChIRP-seq Chem-seq Chip-exo-seq Cir-seq
DMS-seq DNase-seq DNaseI-seq Dup-seq
FAIRE-seq FRAG-seq FRT-seq Frac-seq Freq-
seq GRO-seq GTI-seq HELP-seq HITS-KIN-seq
Hi-C-seq HiTS-Flip-seq IMS-MDA-seq IN-seq Ig-
seq Immuno-seq MeDIP-seq Methylo-seq Mu-seq
NET-seq NOME-seq Nascent-seq Novel-seq
Nucleo-seq PAI-seq PAR-Clip-seq PARS-seq

Sanger sequencing



Nature Methods, 2008



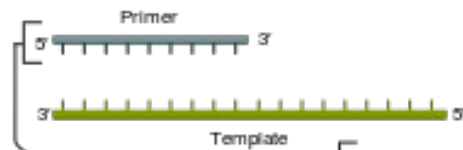
Wikipedia

CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTACACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
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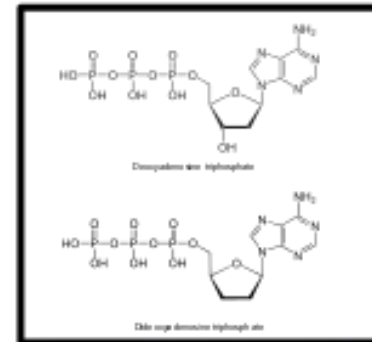
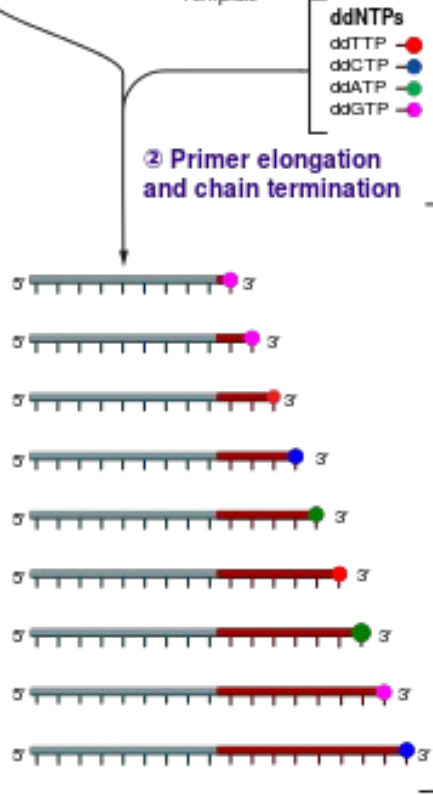
Sanger sequencing

① Reaction mixture

- Primer and DNA template
- DNA polymerase
- ddNTPs with flourochromes
- dNTPs (dATP, dCTP, dGTP, and dTTP)



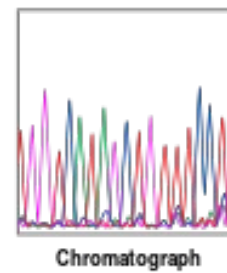
② Primer elongation and chain termination



③ Capillary gel electrophoresis separation of DNA fragments



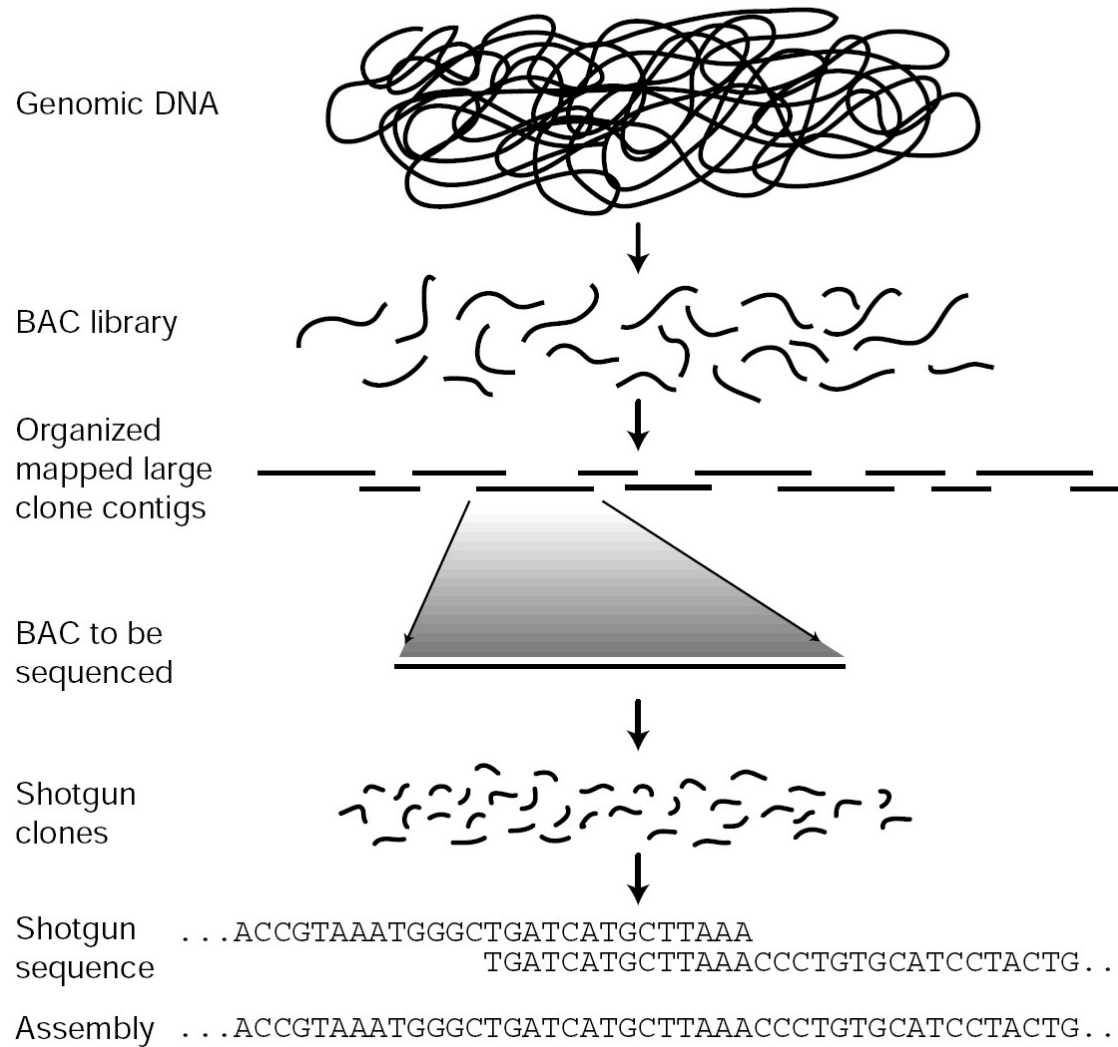
④ Laser detection of flourochromes and computational sequence analysis



CAAAATACTGATATATAACAATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTACACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCCGCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

Shotgun sequencing

Hierarchical shotgun sequencing



Nature, 2001

CAAAATACTGATATATACAAACATGAACGAAATGTCAGACAGTACATGAAAGGACAGAAAGCCCGACAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTCACCACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

2001: draft of the human genome



0101100111011010000111010000100000001010000110001100101001001000000011001000110000001100000011100100101100011001000110000001100
0110111101101110001000000111011001100001011011100010000001001000011001010110010101110010011010010110111001100111011001110110010101101110
1000010110111001101000011001010110010101110010011010010110111001100111011001010110111001000000011011100110001101101101011011000

“Next-generation” sequencing

CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTACCCACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

0101100111011010000111010000100000001010000110001100101001001000000011001000110000001100000011100100101100011001000110000001100
0110111101101110001000000111011001100001011011100010000001001000011001010110010101110010011010010110111001100111011001010110111
1000010110111001101000011001010110010101110010011010010110111001100111011001010110111001000000011011100110001101101101011011000

“Next-generation” sequencing

or

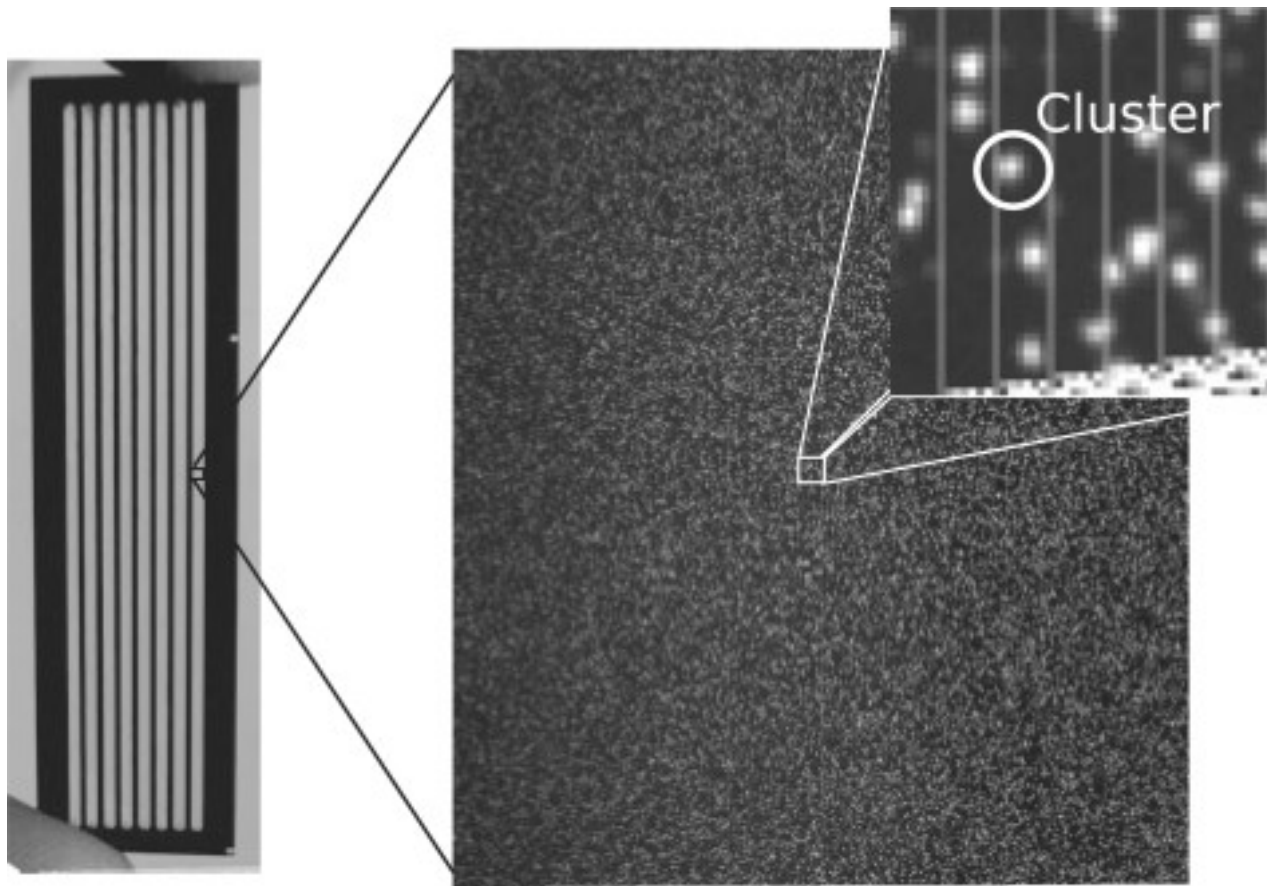
.. a whole lotta reads..

CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTACCCACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

Illumina sequencing



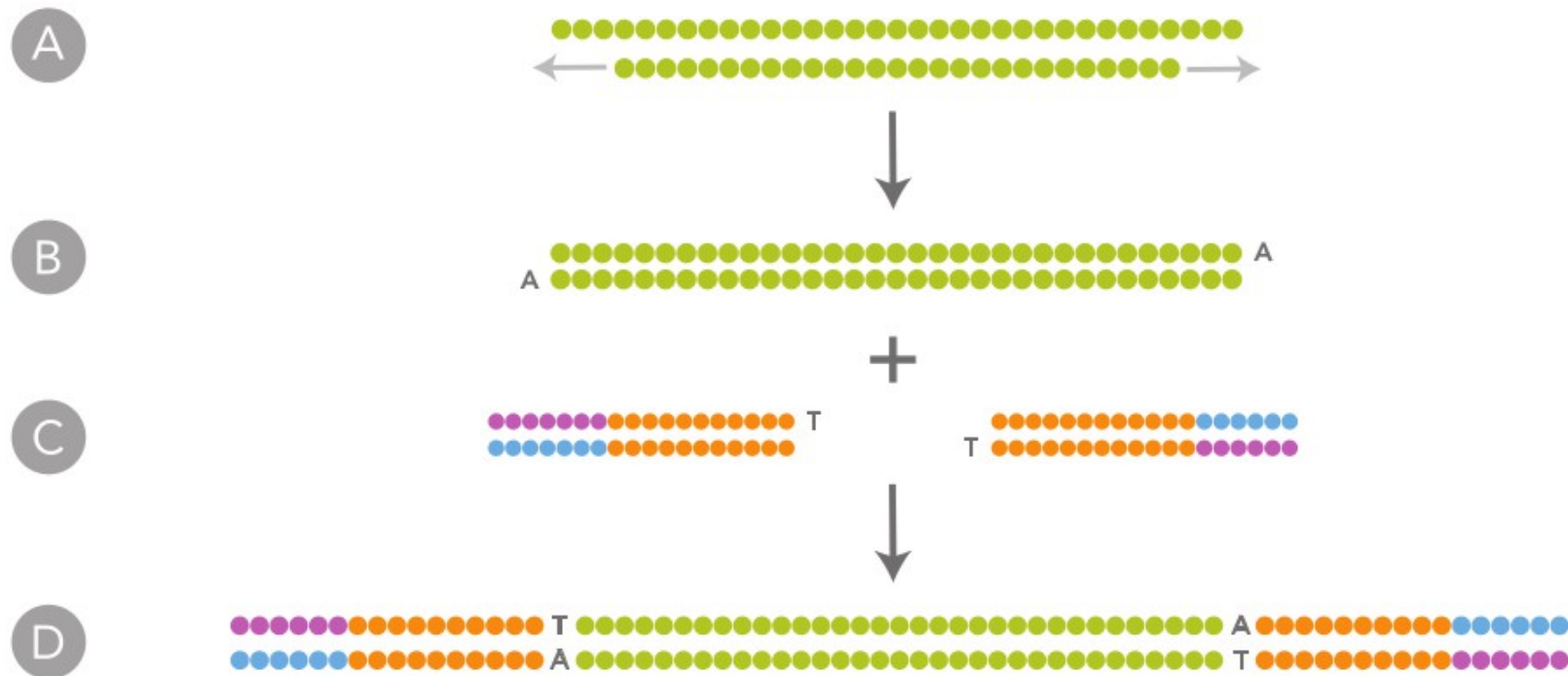
An Illumina flowcell



Whiteford et al., Bioinformatics, 2009

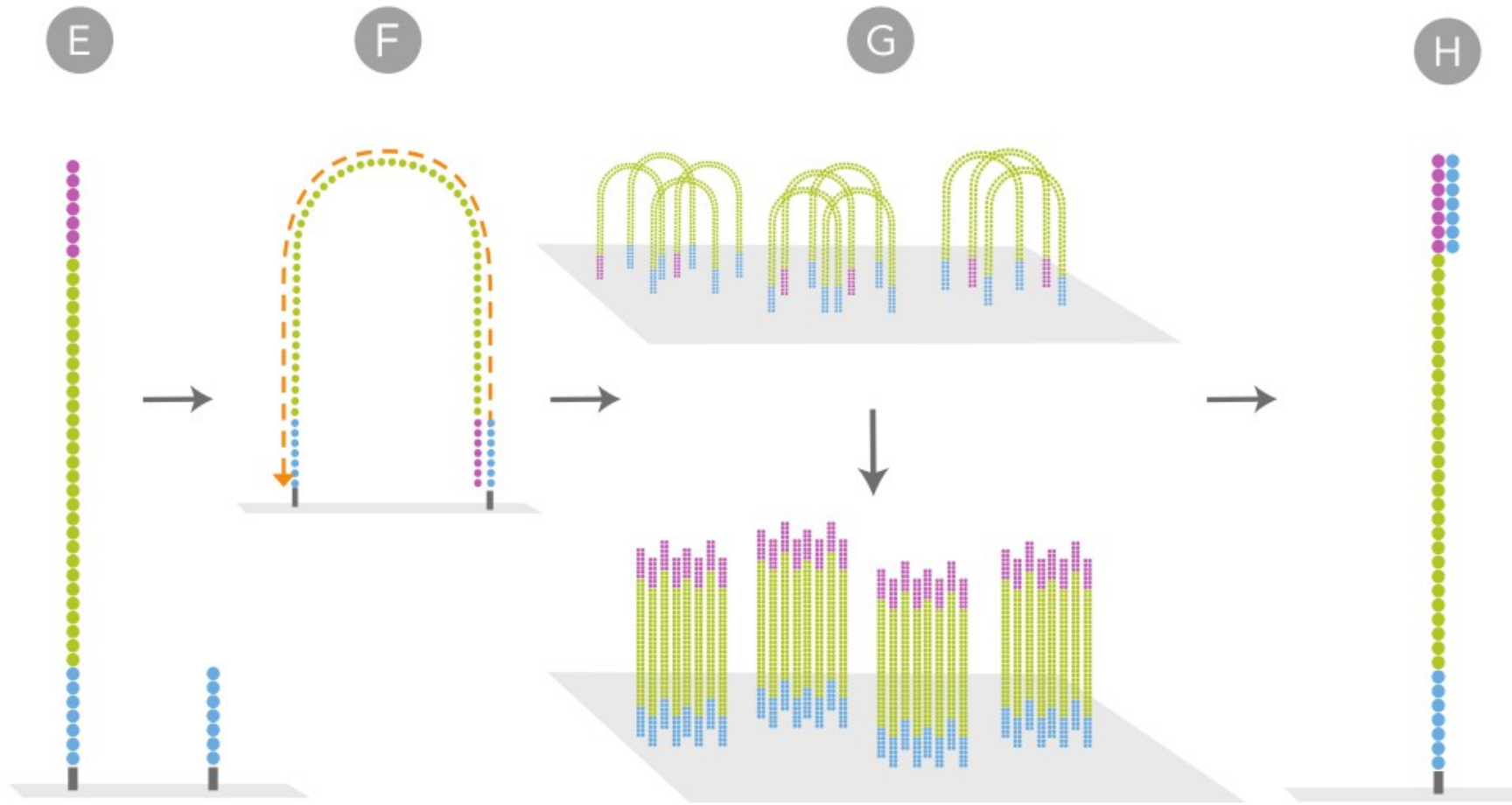
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GGGGATTACACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
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Sequencing by synthesis



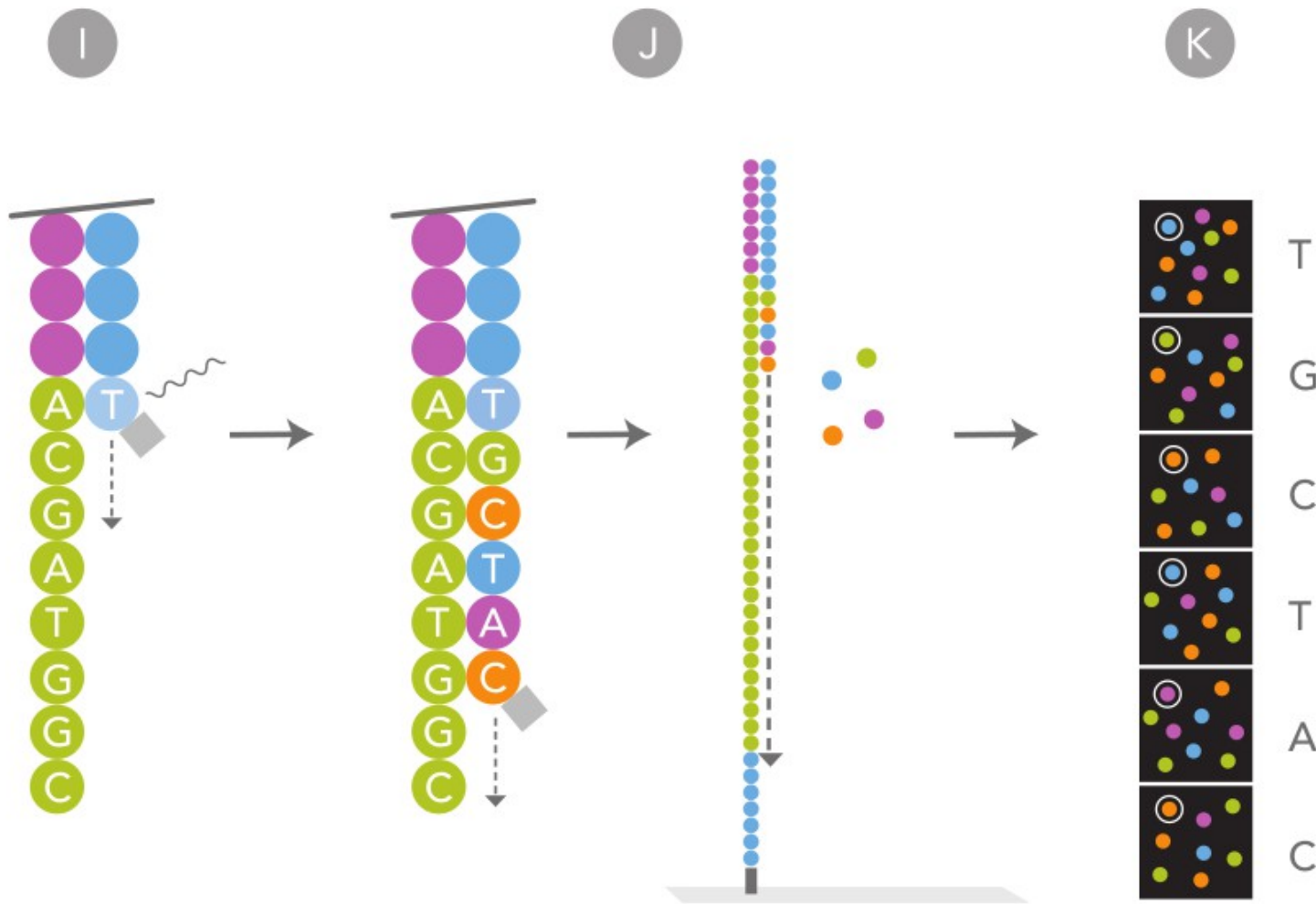
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Sequencing by synthesis



CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTCACCACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

Sequencing by synthesis



CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
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AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

Illumina sequencing systems



MiSeq

Focused power. Speed and simplicity for targeted and small genome sequencing.



NextSeq 500

Flexible power. Speed and simplicity for everyday genomics.



HiSeq 2500

Production power. Power and efficiency for large-scale genomics.



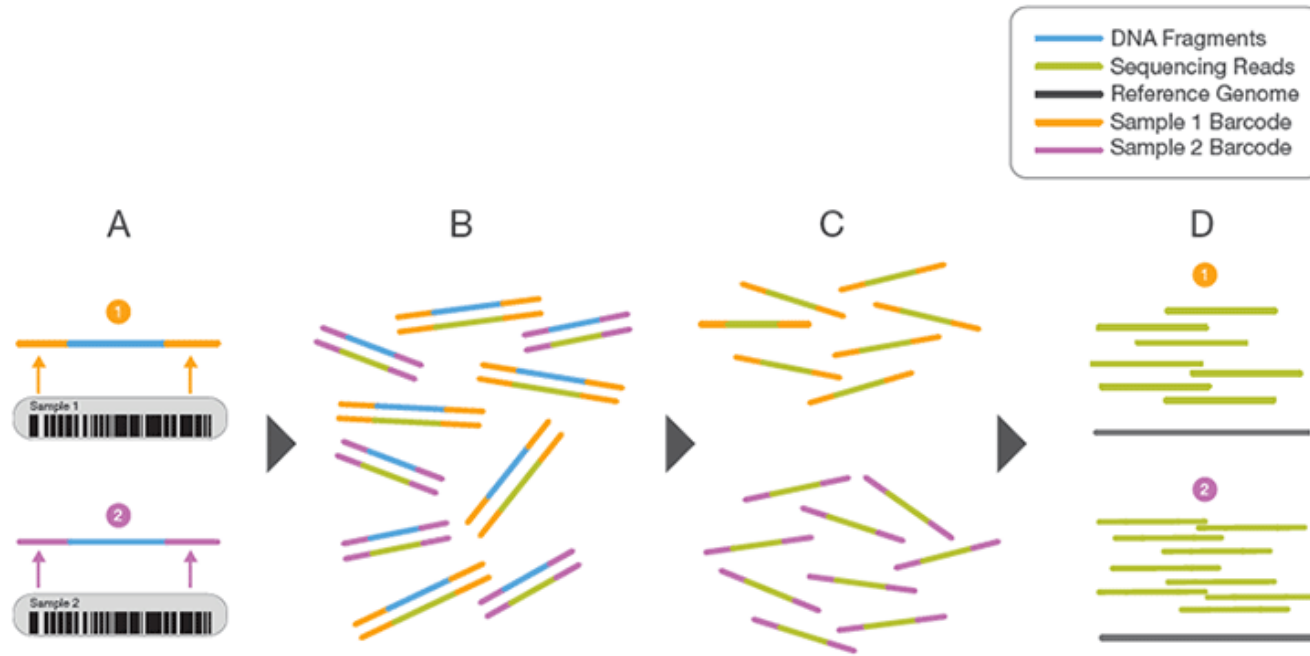
HiSeq X*

Population power. \$1,000 human genome and extreme throughput for population-scale sequencing.

Key applications	Small genome, amplicon, and targeted gene panel sequencing.	Everyday genome, exome, transcriptome sequencing, and more.		Production-scale genome, exome, transcriptome sequencing, and more.		Population-scale human whole-genome sequencing.
Run mode	N/A	Mid-Output	High-Output	Rapid Run	High-Output	N/A
Flow cells processed per run	1	1	1	1 or 2	1 or 2	1 or 2
Output range	0.3-15 Gb	20-39 Gb	30-120 Gb	10-300 Gb	50-1000 Gb	1.6-1.8 Tb
Run time	5-55 hours	15-26 hours	12-30 hours	7-60 hours	< 1 day - 6 days	< 3 days
Reads per flow cell†	25 Million‡	130 Million	400 Million	300 Million	2 Billion	3 Billion
Maximum read length	2 × 300 bp	2 × 150 bp	2 × 150 bp	2 × 250 bp	2 × 125 bp	2 × 150 bp

Multiplexing

Figure 2: Conceptual Overview of Sample Multiplexing



- Two representative DNA fragments from two unique samples, each attached to a specific barcode sequence that identifies the sample from which it originated.
- Libraries for each sample are pooled and sequenced in parallel. Each new read contains both the fragment sequence and its sample-identifying barcode.
- Barcode sequences are used to de-multiplex, or differentiate reads from each sample.
- Each set of reads is aligned to the reference sequence.

Illumina

CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTACACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

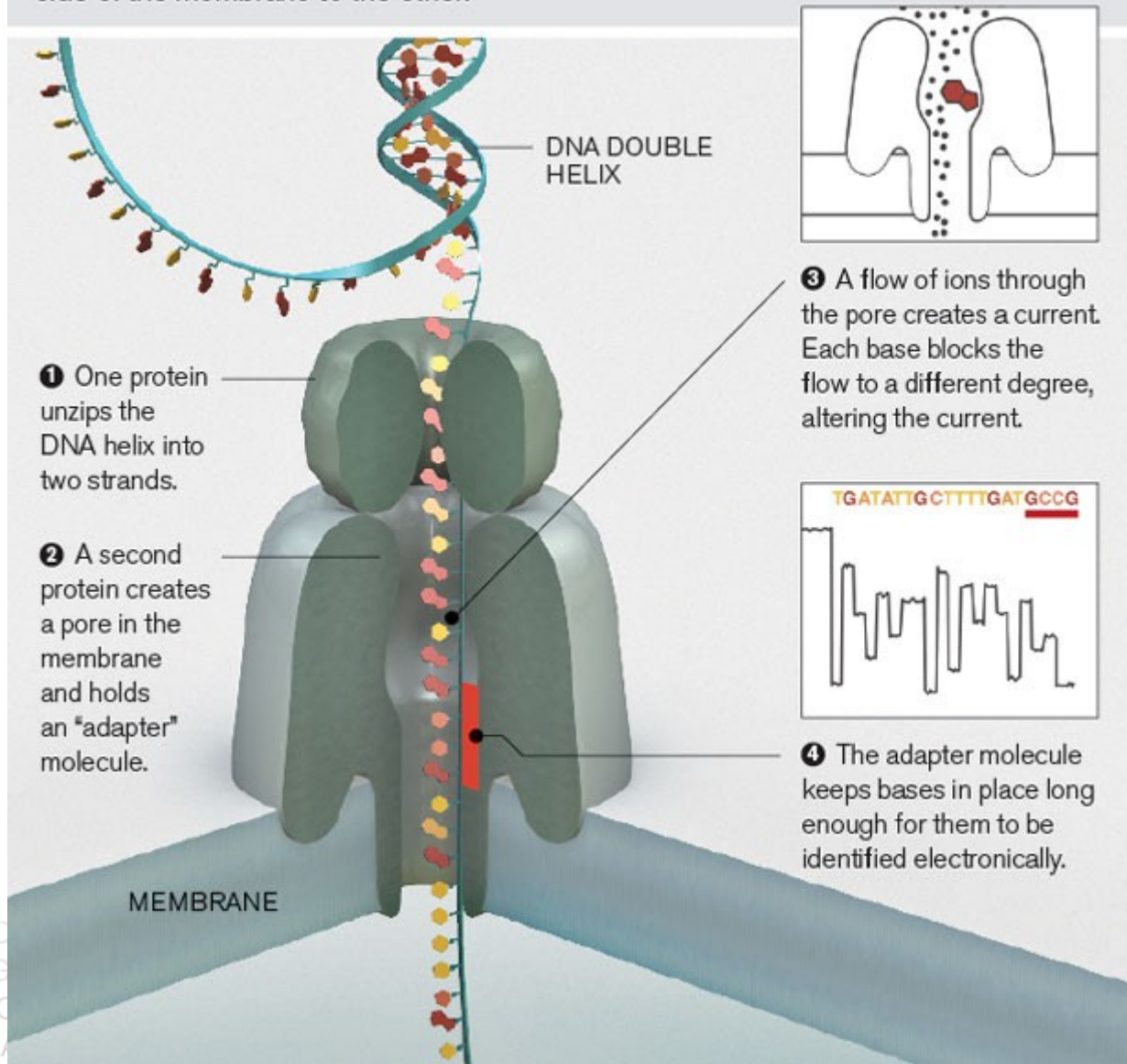
Other technologies...



<http://nextgenseek.com>

Nanopore sequencing

DNA can be sequenced by threading it through a microscopic pore in a membrane. Bases are identified by the way they affect ions flowing through the pore from one side of the membrane to the other.



The MinION



Pacific Biosciences



What's next?

- It's a rapidly evolving field
- Existing technologies continually improve
- Illumina started out as a small player 10 years ago (Solexa)
- Sequencing as a service?
- Benchtop (USB-sized?) sequencers?

The FASTQ format

```
@D256N5M1:31:C1B42ACXX:4:2305:3881:47
605
ACCCCCCACAGGGACCCTTGTCACGTCCCCCTAACTC
CCTGC
+
@?
@FDFFFDFFFDDBGIIIIIGDGHIG@GHIIGEF@@DFH
GGI
```

FASTQ quality scores

Phred quality score

$$Q_{\text{sanger}} = -10 \log_{10} p$$

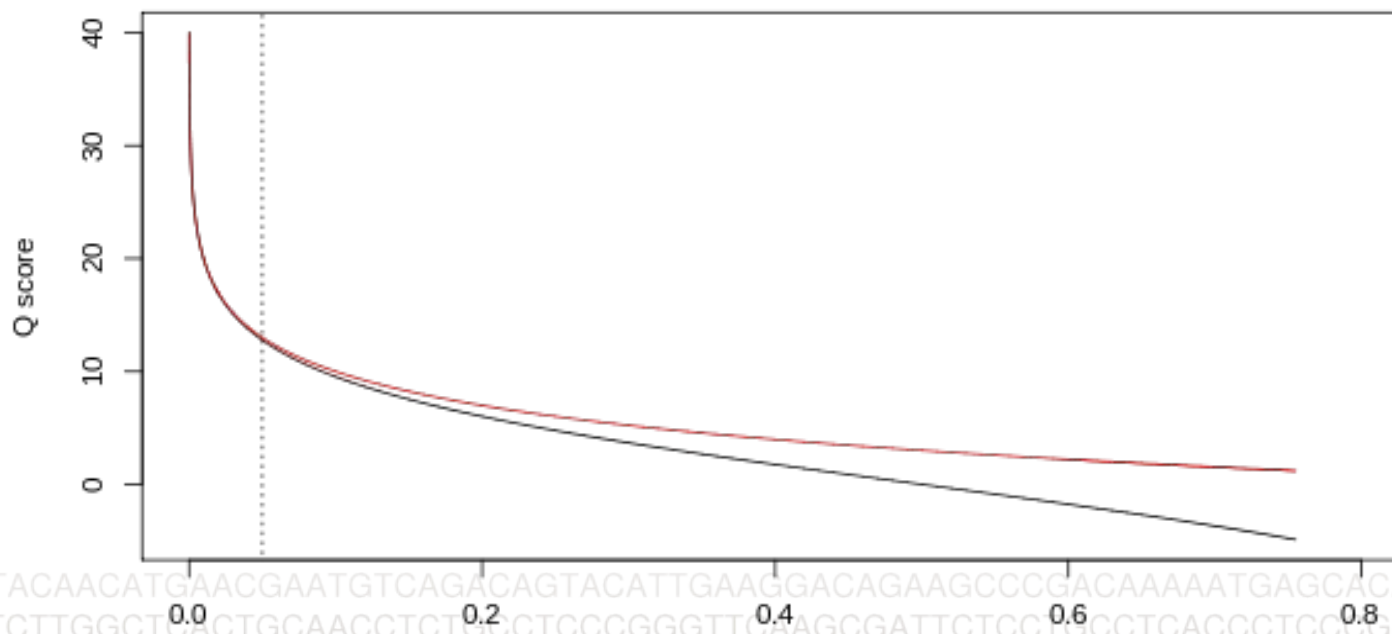
Q10	1 in 10	90%
Q20	1 in 100	99%
Q30	1 in 1000	99.9%
Q40	1 in 10000	99.99%

FASTQ quality scores

Phred quality score

$$Q_{\text{sanger}} = -10 \log_{10} p$$

Q10	1 in 10	90%
Q20	1 in 100	99%
Q30	1 in 1000	99.9%
Q40	1 in 10000	99.99%



FASTQ quality scores

Table 1 ASCII Characters Encoding Q-scores 0-40

Symbol	ASCII Code	Q-Score	Symbol	ASCII Code	Q-Score	Symbol	ASCII Code	Q-Score
!	33	0	/	47	14	=	61	28
"	34	1	0	48	15	>	62	29
#	35	2	1	49	16	?	63	30
\$	36	3	2	50	17	@	64	31
%	37	4	3	51	18	A	65	32
&	38	5	4	52	19	B	66	33
'	39	6	5	53	20	C	67	34
(40	7	6	54	21	D	68	35
)	41	8	7	55	22	E	69	36
*	42	9	8	56	23	F	70	37
+	43	10	9	57	24	G	71	38
,	44	11	:	58	25	H	72	39
-	45	12	;	59	26	I	73	40
.	46	13	<	60	27			

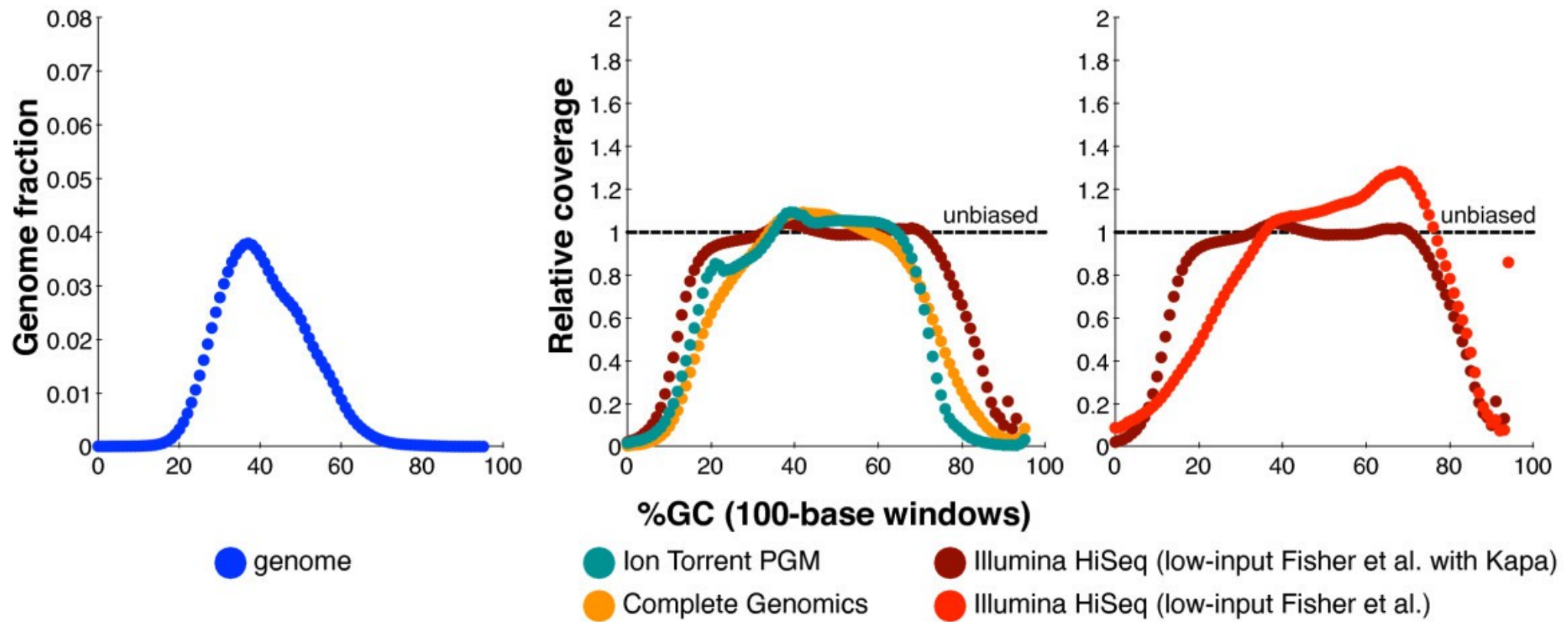
FASTQonfusion!

- Illumina:
 - CASAVA ≤ 1.3 Solexa
 - CASAVA 1.3 – 1.7 Illumina
 - CASAVA ≥ 1.8 Sanger (the “standard”)

Data quality and bias

- Sequencing errors
 - Different for different techniques!
- Amplification in sample prep => duplicate reads
- GC bias
 - Sample prep
 - Bridge amplification

GC bias



Ross et al, Genome Biology, 2013

Workflow



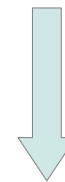
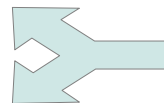
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ATATTGGGGTTTGGATAAACTGTAAAGCAGATTTGTCTTTCCTGAAACACT
TGTCAGTAACATTTTAAAAACAGTACAAGACATAATAGTGCCCATTTGGGC
ATAACTGTCAAATGAAACAATCATCAAGTGAATTGAGTTTTAGTAGGAAT
CAGGGGTCTTGCCTGACAGAAGTGGGATTAACAGGATTCTAAAAAAGCT
TTACAGATCGCCAAACCACAACAATAACAAAGGCATGGATAGGGATCC
TTAAAAAGCTTATCCCAATATGAATTGTTCCATATGGACCACTGTCAGAGG
GATTTTCCCGGGCTTAGGAAGGGGAGGAGCGAGCAAGACAGCCTACCTTTT
```

Identification of duplicates

Quantification

Peak calling

....



Mapping to reference



```
CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
GGGGATTACACGTTGGCCACGCTGGTCTGGAACCTCTATCCTCAAGTAATCCGCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC
```

Workflow



```
CTAGTGATTTATCATCTAGGCCAGTGAATACCAAGTGGGTGGCAACCCTACC
GAATGCTCGAGCGTTCATGCGAACGATCCGAGCGCATTTCGGCGCACGAC
CATGATGTGTAGGTAATGATTCTGAGACAAATTGCAATTGGTTTTTCATTTT
ATATTGGGGTTTGGATAAACTGTAAAGCAGATTTGTCTTTCCTGAAACACT
TGTCAGTAACATTTTAAAAACAGTACAAGACATAATAGTGCCCATTGGGC
ATAACTGTCAAATGAAACAATCATCAAGTGAATTGAGTTTTAGTAGGAAT
CAGGGGTCTTGCCTGACAGAAGTGGGATTAACAGGATTCTAAAAAAGCT
TTACAGATCGCCAAACCACAACAATAACAAAGGCATGGATAGGGATCC
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GATTTTCCCGGGCTTAGGAAGGGGAGGAGCGAGCAAGACAGCCTACCTTTT
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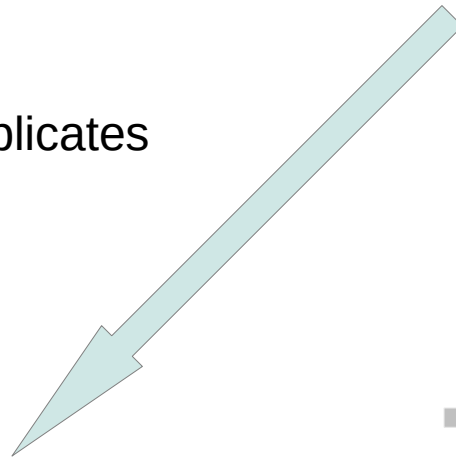
Identification of duplicates

Quantification

Peak calling

Assembly

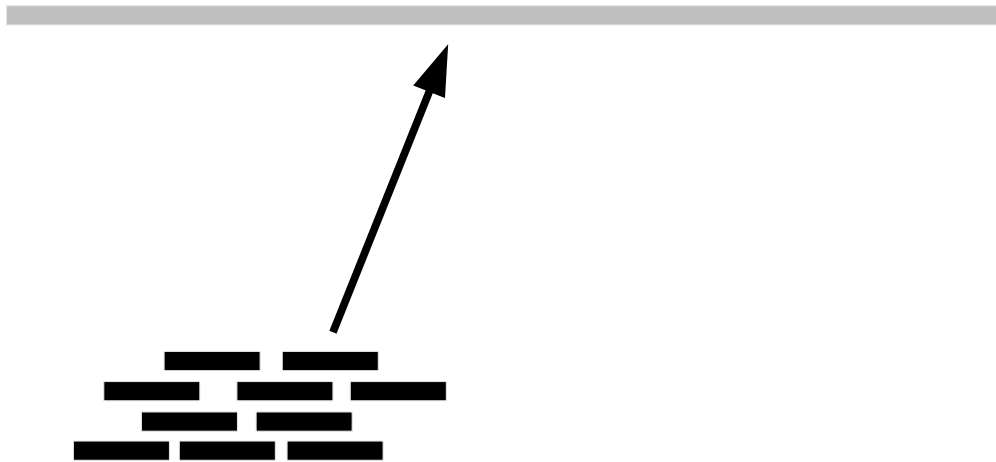
....



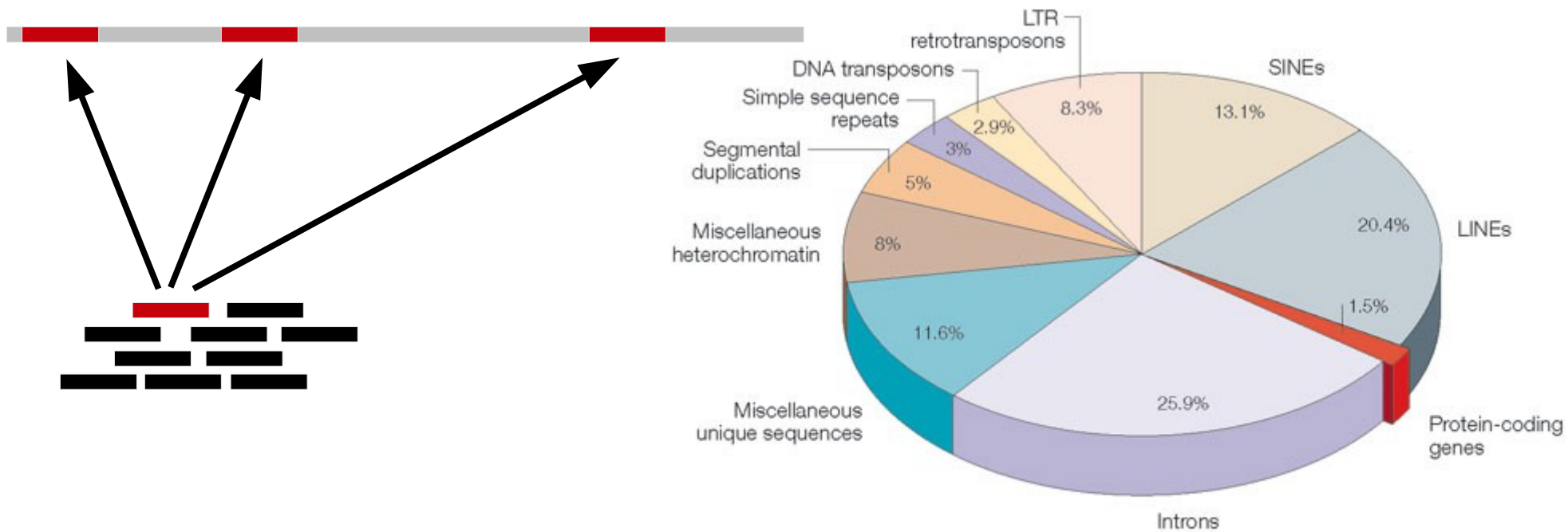
```
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GGGGATTACACGTTGGCCACGCTGGTCTGGAACCTCCTATCCTCAAGTAATCCGCCCGCCTCGGCCTCCCAAAGTGCAGGCGTGAGCCAC
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC
```

Mapping to reference

- Genome or transcriptome
 - Any other set of sequences



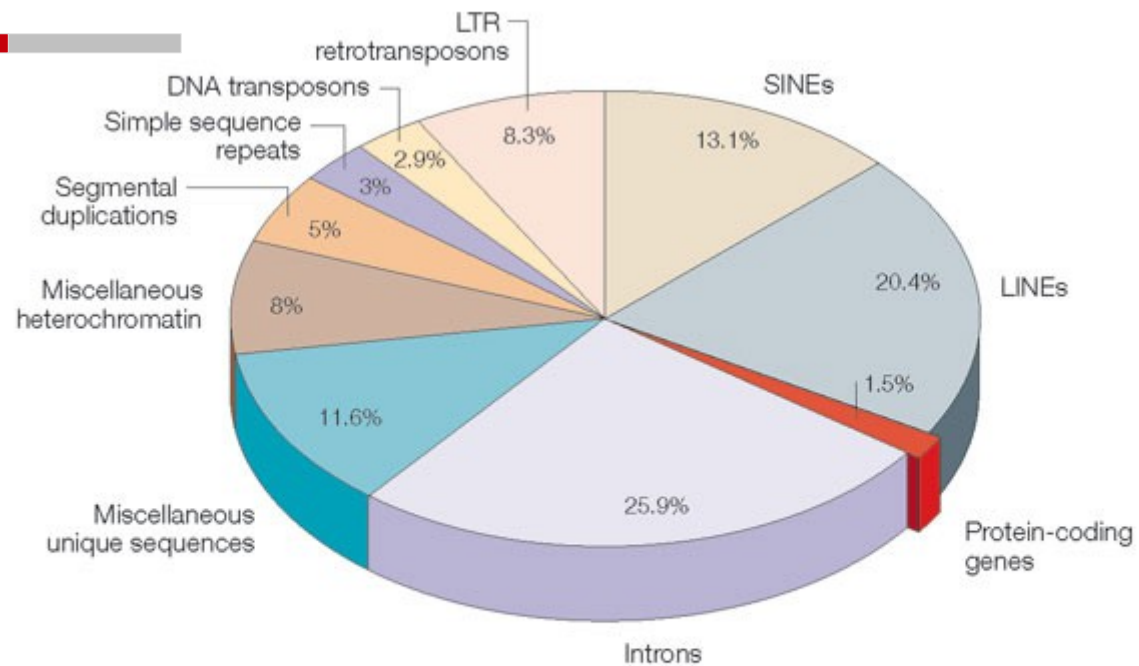
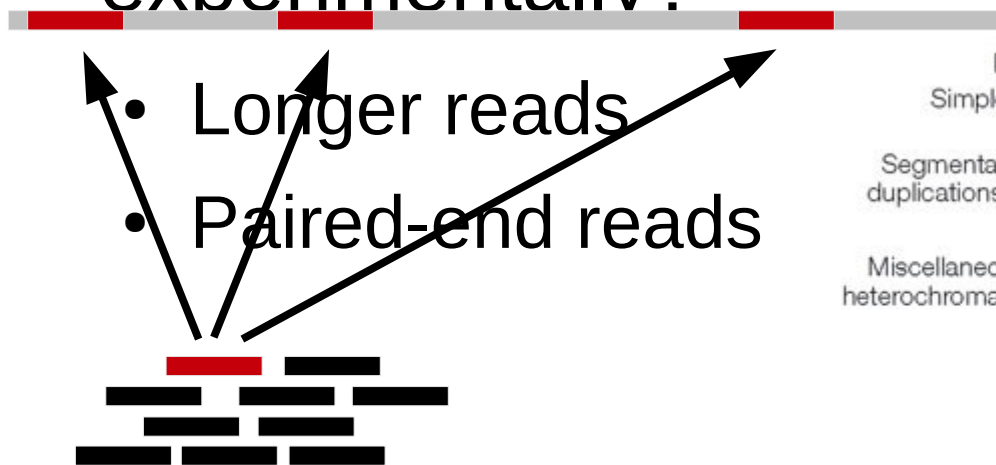
Repetitive sequence



CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTA
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCC
GGGGATTACACGTTGGCCACGCTGGTCTGGAACCTCCTAT
AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGAATC

Repetitive sequence

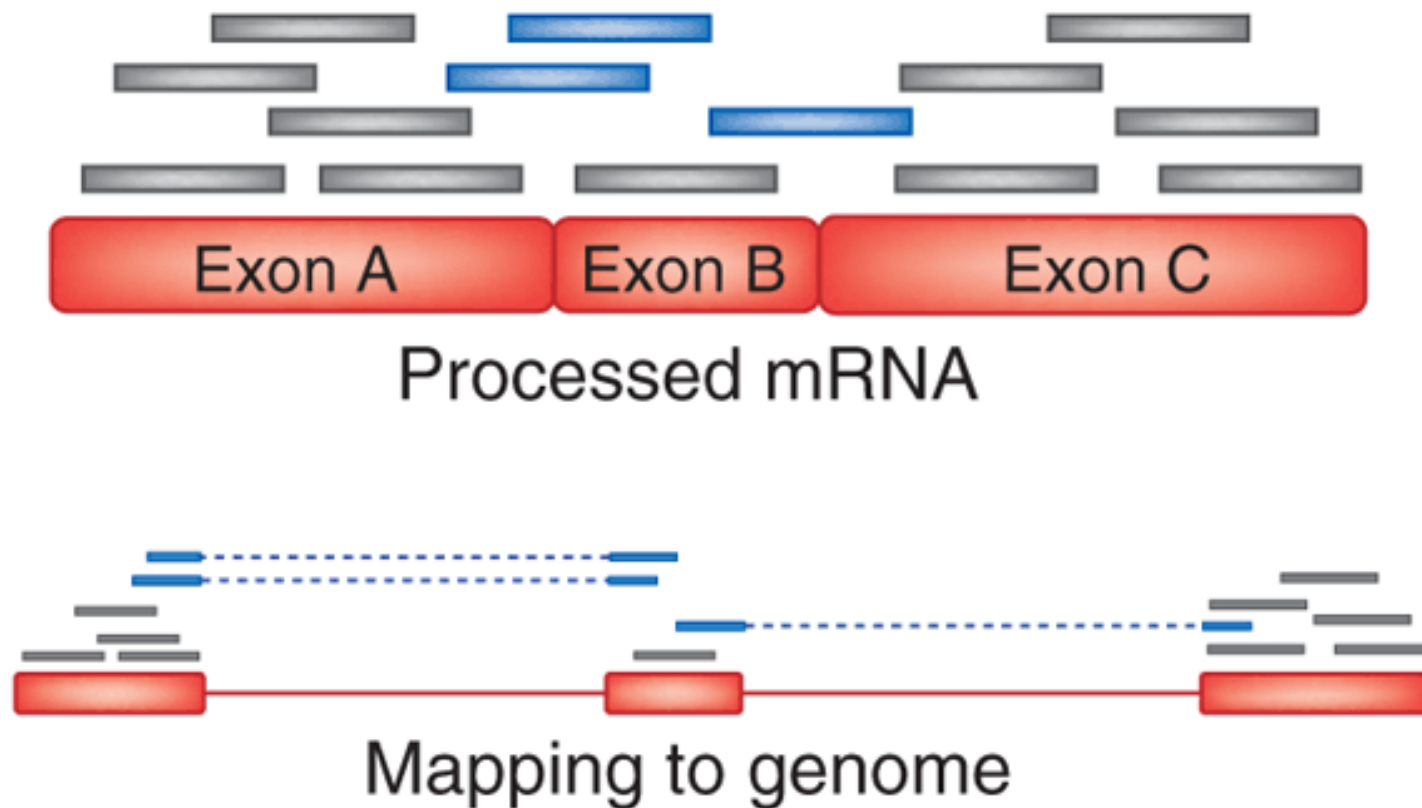
- How to deal with repeats during mapping?
- What can be done experimentally?



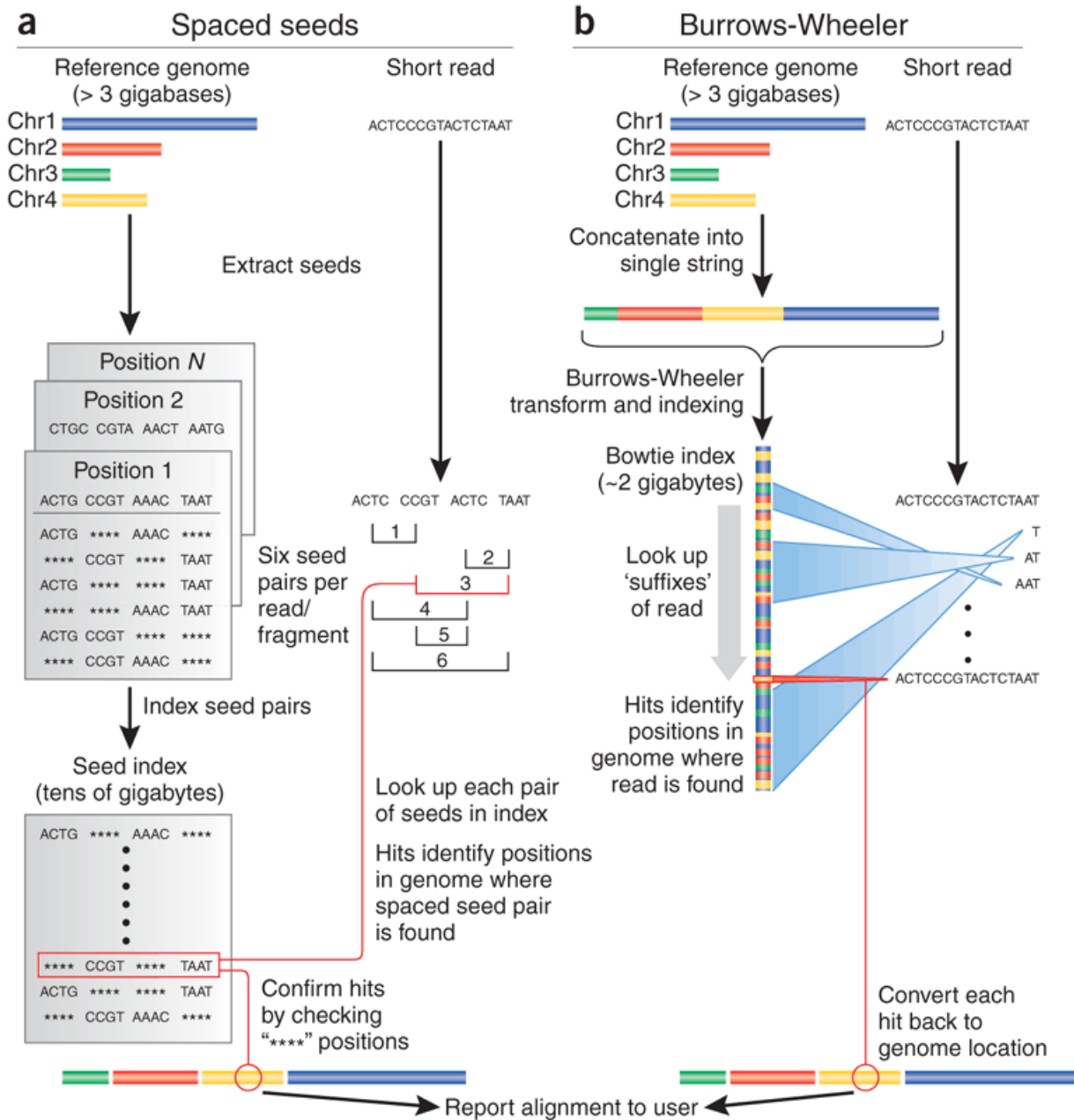
Difficulties

- Sequencing errors
- Errors in reference genome
- Polymorphisms
 - Insertions
 - Deletions
 - SNPs
- Spliced alignment

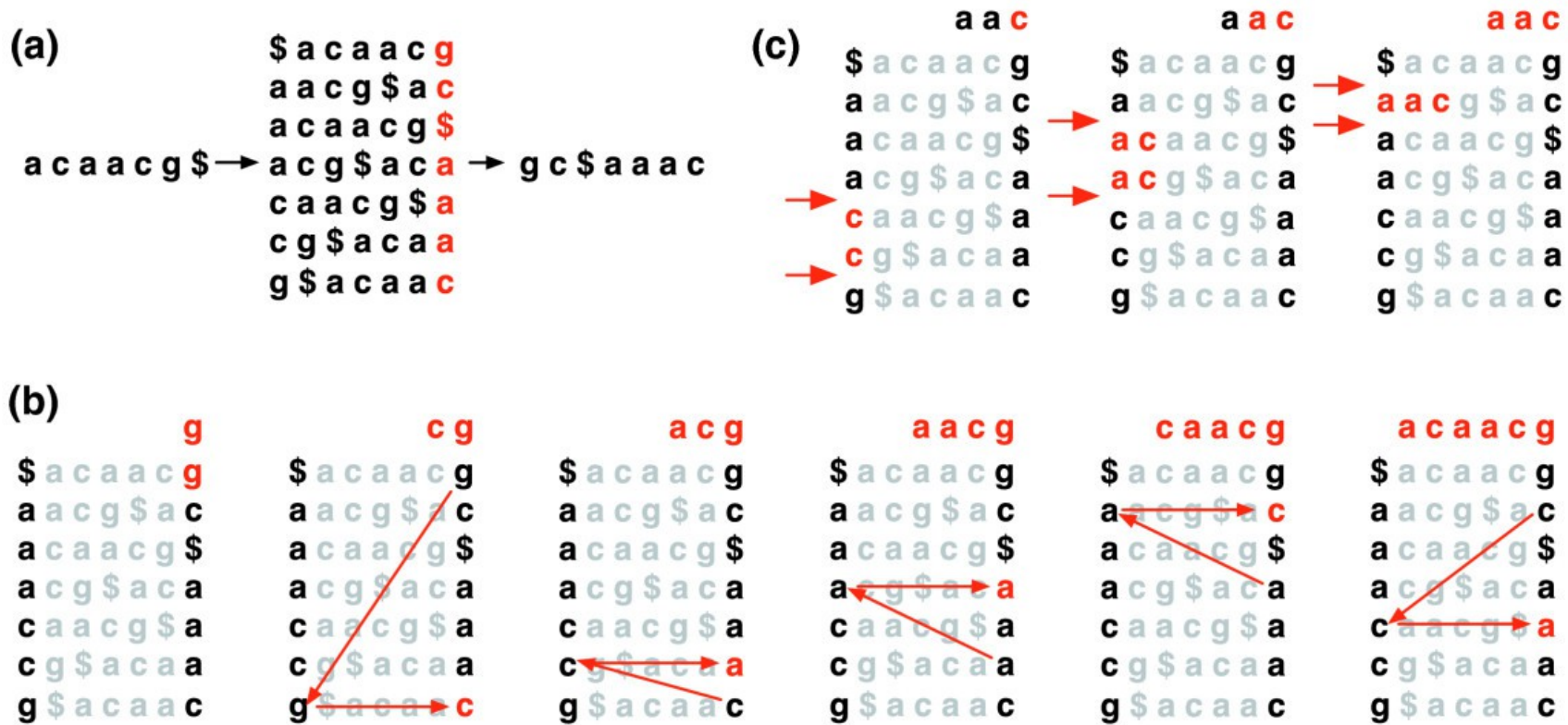
Mapping RNA-seq / spliced alignment



Approaches to mapping



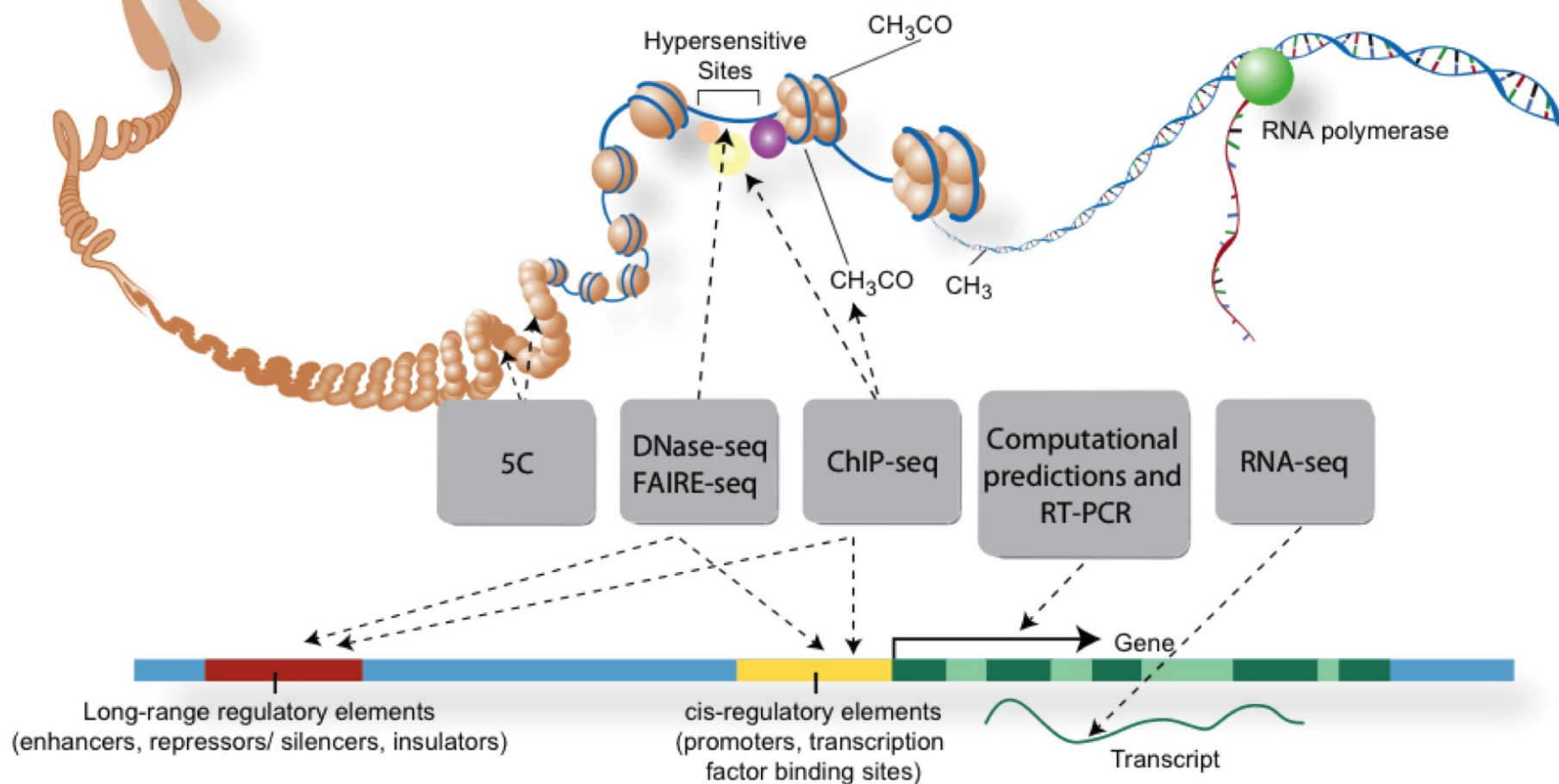
Burrows-Wheeler transform



Langmead et al, 2009

CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
GTGTAGTGGCACGATCTTGGCTCACTGCAACCTCTGCCTCCCGGGTTCAAGCGATTCTCCTGCCTCACCTCCCGAATAGCTGGGATTACAG
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AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

Next-gen seq applications



The ENCODE Consortium

CAAAATACTGATATATACAACATGAACGAATGTCAGACAGTACATTGAAGGACAGAAGCCCGACAAAAATGAGCACATAATGTATGATTCCCC
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AAAATGGTTATGGAGATCAAAATAAAGGTGGGGTCGGGAATCGACTGGGAAGAGACGTGATGAAACGTTTCTGGGACGATGAAAAGGGTCTC

Big Data

40 ZETTABYTES

[43 TRILLION GIGABYTES]
of data will be created by 2020, an increase of 300 times from 2005



Volume
SCALE OF DATA



It's estimated that
2.5 QUINTILLION BYTES
[2.5 TRILLION GIGABYTES]
of data are created each day



Most companies in the U.S. have at least
100 TERABYTES
[100,000 GIGABYTES]
of data stored



The FOUR V's of Big Data

From traffic patterns and music downloads to web history and medical records, data is recorded, stored, and analyzed to enable the technology and services that the world relies on every day. But what exactly is big data, and how can these massive amounts of data be used?

As a leader in the sector, IBM data scientists break big data into four dimensions: **Volume, Velocity, Variety and Veracity**

Depending on the industry and organization, big data encompasses information from multiple internal and external sources such as transactions, social media, enterprise content, sensors and mobile devices. Companies can leverage data to adapt their products and services to better meet customer needs, optimize operations and infrastructure, and find new sources of revenue.

By 2015
4.4 MILLION IT JOBS
will be created globally to support big data, with 1.9 million in the United States



As of 2011, the global size of data in healthcare was estimated to be

150 EXABYTES
[161 BILLION GIGABYTES]



30 BILLION
PIECES OF CONTENT
are shared on Facebook every month



Variety
DIFFERENT FORMS OF DATA



By 2014, it's anticipated there will be
420 MILLION
WEARABLE, WIRELESS
HEALTH MONITORS

4 BILLION+
HOURS OF VIDEO
are watched on YouTube each month



400 MILLION TWEETS
are sent per day by about 200 million monthly active users



The New York Stock Exchange captures
1 TB OF TRADE
INFORMATION
during each trading session



Velocity
ANALYSIS OF
STREAMING DATA



Modern cars have close to
100 SENSORS
that monitor items such as
fuel level and tire pressure

By 2016, it is projected there will be
18.9 BILLION
NETWORK
CONNECTIONS
— almost 2.5 connections per person on earth



1 IN 3 BUSINESS
LEADERS
don't trust the information they use to make decisions

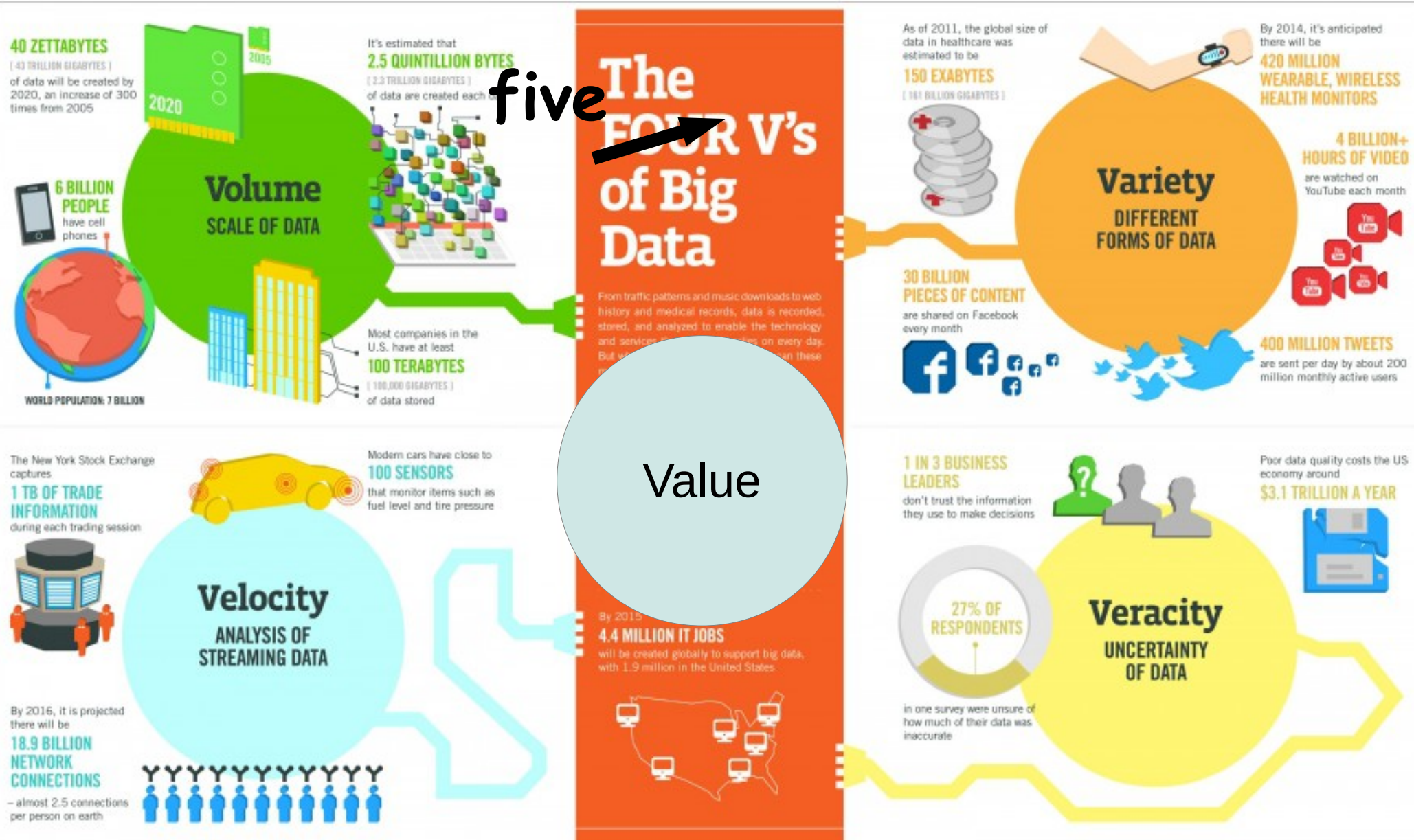


Veracity
UNCERTAINTY OF DATA

Poor data quality costs the US economy around
\$3.1 TRILLION A YEAR



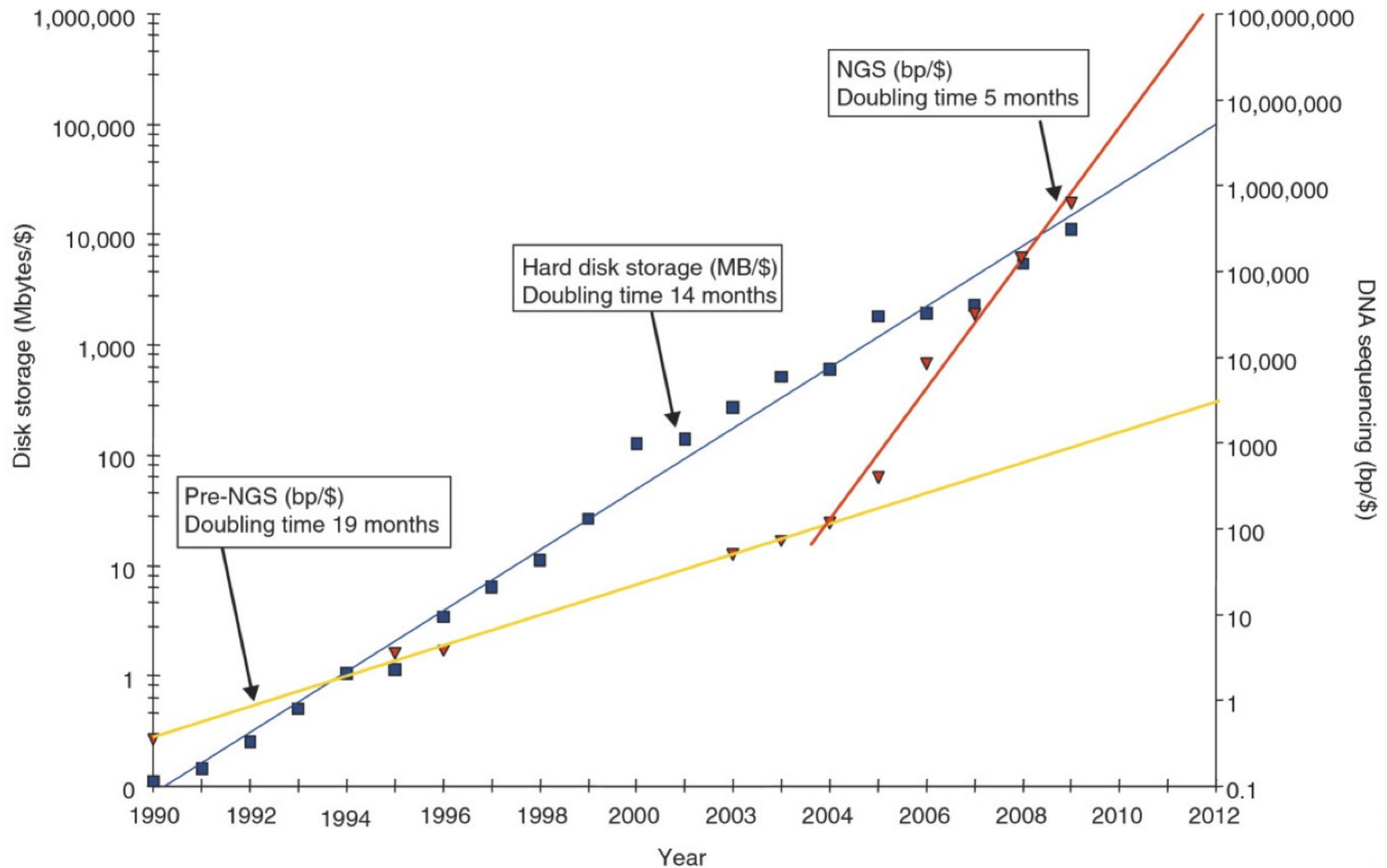
Big Data



Challenges

- Storage
 - From terabytes to petabytes
- Analysis
 - Computational resources
 - Memory
 - CPU
- Sharing
 - Bandwidth

NextGen Sequencing a Game-Changer



Lincoln Stein (via C. Titus Brown)

Opportunities

- A wealth of data in public databases
- Computational analysis:
 - Reproducible
 - Not dependent on lab / materials
- Cloud-based analysis
 - Amazon AWS
 - National HPC and e-Science resources
 - The Netherlands: SURFsara

ARTICLE

Received 9 Apr 2014 | Accepted 25 Jun 2014 | Published 24 Jul 2014

DOI: 10.1038/ncomms5498

OPEN

A highly abundant bacteriophage discovered in the unknown sequences of human faecal metagenomes

Bas E. Dutilh^{1,2,3,4}, Noriko Cassman^{3,†}, Katelyn McNair², Savannah E. Sanchez³, Genivaldo G.Z. Silva⁵, Lance Boling³, Jeremy J. Barr³, Daan R. Speth⁶, Victor Seguritan³, Ramy K. Aziz^{2,7}, Ben Felts⁸, Elizabeth A. Dinsdale^{3,5}, John L. Mokili³ & Robert A. Edwards^{2,4,5,9}

Research

Highly accessed

Open Access

DNA methylation age of human tissues and cell types

Steve Horvath

Correspondence: Steve Horvath shorvath@mednet.ucla.edu

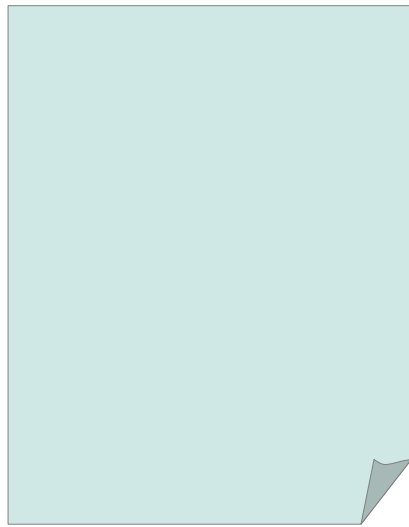
▼ Author Affiliations

Human Genetics, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, CA 90095, USA

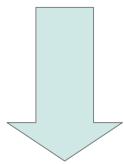
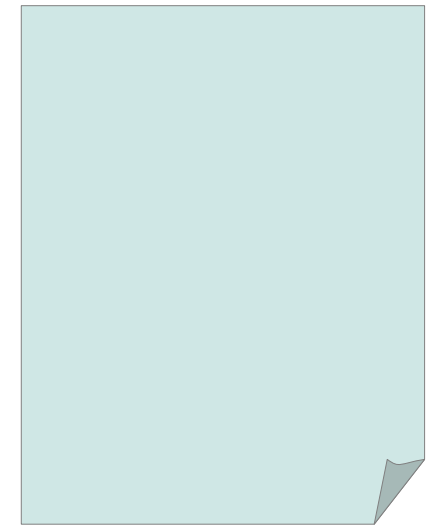
Biostatistics, School of Public Health, University of California Los Angeles, Los Angeles, CA 90095, USA

Human Genetics, Gonda Research Center, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, CA 90095-7088, USA

Bioinformatic analysis in a nutshell

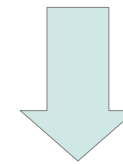


“Do something”



Mostly text:

- Sequences
- Genomic coordinates
- Etc.



Mostly text:

- Sequences
- Genomic coordinates
- Etc.

A side note...

Bioinformatician? Computational

Biologist? Data analyst? Data
curator? Database developer?

Statistician? Mathematical

Modeler? Software Developer?

Ontologist? Programmer?

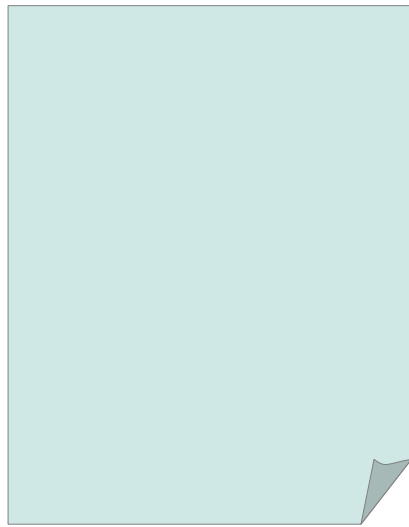
So you want to be a computational biologist? Loman & Watson, 2013



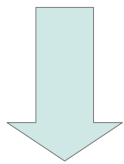
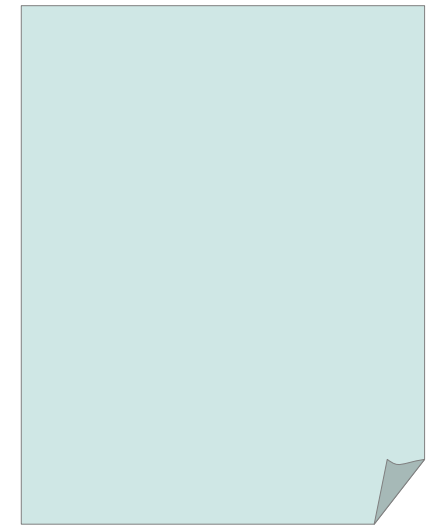
Computational Biology

- It's about the biology
- It's research
- The computer is just the tool used to answer interesting questions
- Iterative, collaborative process between wet-lab and dry-lab

Bioinformatic analysis in a nutshell

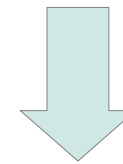


“Do something”



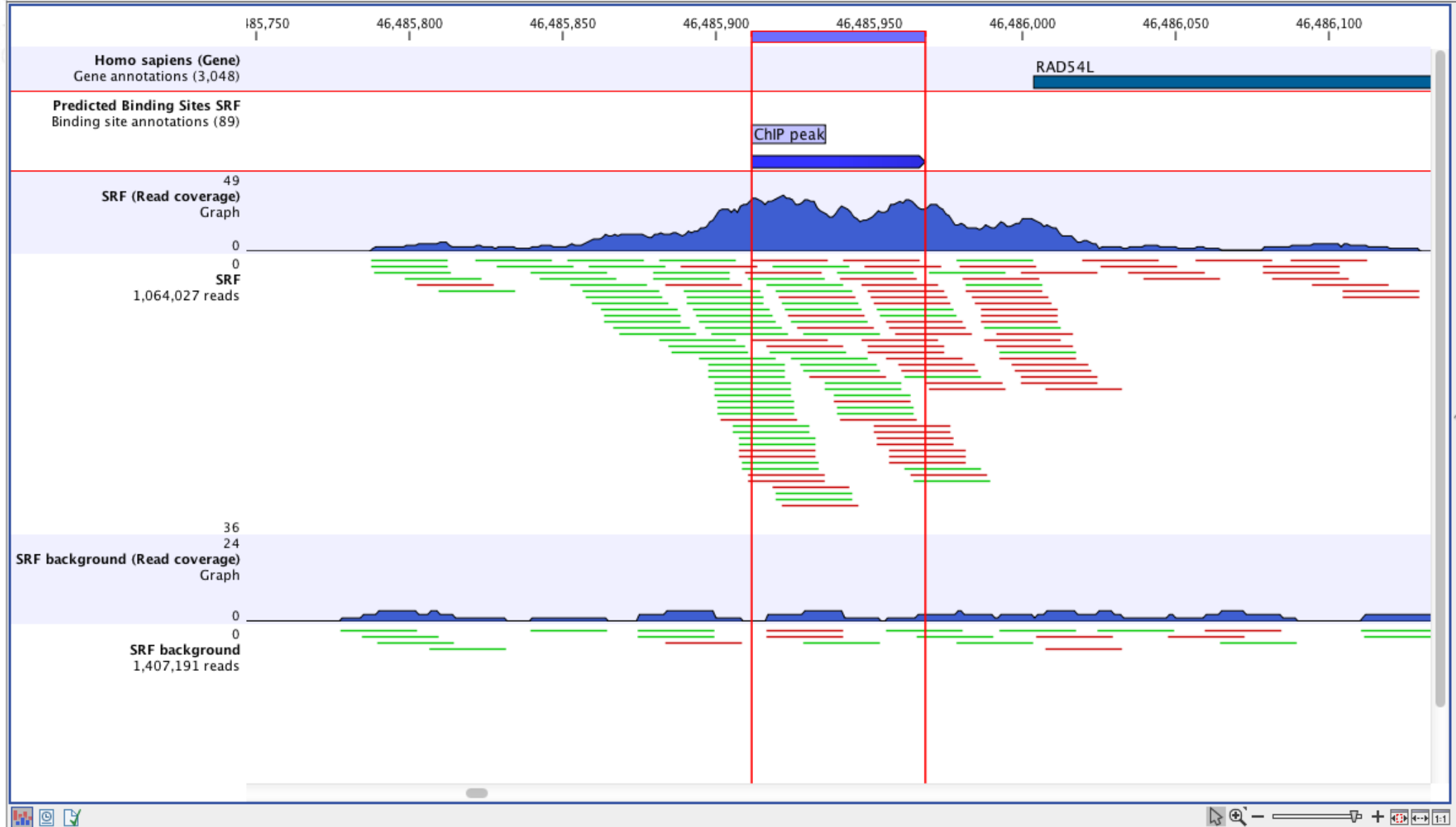
Mostly text:

- Sequences
- Genomic coordinates
- Etc.



Mostly text:

- Sequences
- Genomic coordinates
- Etc.



Chromosome	Region	Name	p-value	Score	FDR	note
chr1	45224888..45224947	ChIP peak	0.00	0.00	4.43E-108	# forward reads : 196, # reverse reads : 222, Region containing reads : 45224467..45225
chr1	45578365..45578433	ChIP peak	3.82E-6	3.82E-6	3.63E-6	# forward reads : 43, # reverse reads : 63, Region containing reads : 45577983..4557871
chr1	46024004..46024099	ChIP peak	9.20E-6	9.20E-6	6.18E-4	# forward reads : 18, # reverse reads : 13, Region containing reads : 46023761..4602445
chr1	46485912..46485968	ChIP peak	5.02E-14	5.02E-14	1.54E-34	# forward reads : 85, # reverse reads : 80, Region containing reads : 46485517..4648639
chr1	46855203..46855264	ChIP peak	0.00	0.00	3.36E-86	# forward reads : 162, # reverse reads : 160, Region containing reads : 46854793..46855
chr1	51539515..51539583	ChIP peak	4.56E-5	4.56E-5	4.10E-22	# forward reads : 53, # reverse reads : 44, Region containing reads : 51539124..5153995

► NCBI/BLAST Home

BLAST finds regions of similarity between biological sequences. [more...](#)

New DELTA-BLAST, a more sensitive protein-protein search

Go

BLAST Assembled RefSeq Genomes

Choose a species genome to search, or [list all genomic BLAST databases](#).

- | | | | |
|--|---|--|--|
| <input type="checkbox"/> Human | <input type="checkbox"/> Dog | <input type="checkbox"/> Fruit fly | <input type="checkbox"/> Arabidopsis |
| <input type="checkbox"/> Mouse | <input type="checkbox"/> Rabbit | <input type="checkbox"/> Honey bee | <input type="checkbox"/> Rice |
| <input type="checkbox"/> Rat | <input type="checkbox"/> Chimp | <input type="checkbox"/> Chicken | <input type="checkbox"/> Yeast |
| <input type="checkbox"/> Cow | <input type="checkbox"/> Guinea pig | <input type="checkbox"/> Zebrafish | <input type="checkbox"/> Neurospora crassa |
| <input type="checkbox"/> Pig | <input type="checkbox"/> Sheep | <input type="checkbox"/> Clawed frog | <input type="checkbox"/> Microbes |

Basic BLAST

Choose a BLAST program to run.

- | | |
|----------------------------------|--|
| nucleotide blast | Search a nucleotide database using a nucleotide query
<i>Algorithms: blastn, megablast, discontinuous megablast</i> |
| protein blast | Search protein database using a protein query
<i>Algorithms: blastp, psi-blast, phi-blast, delta-blast</i> |
| blastx | Search protein database using a translated nucleotide query |
| tblastn | Search translated nucleotide database using a protein query |
| tblastx | Search translated nucleotide database using a translated nucleotide query |

Specialized BLAST

Choose a type of specialized search (or database name in parentheses.)

- ☐ Make specific primers with [Primer-BLAST](#)
- ☐ Search [trace archives](#)
- ☐ Find [conserved domains](#) in your sequence (cds)
- ☐ Find sequences with similar [conserved domain architecture](#) (cdart)
- ☐ Search sequences that have [gene expression profiles](#) (GEO)
- ☐ Search [immunoglobulins and T cell receptor sequences](#) (IgBLAST)
- ☐ Screen sequence for [vector contamination](#) (vecscreen)
- ☐ [Align](#) two (or more) sequences using BLAST (bl2seq)
- ☐ Search [protein](#) or [nucleotide](#) targets in PubChem BioAssay
- ☐ Search [SRA by experiment](#)

Your Recent Results **New!**

 [All Recent results...](#)

News

[New gap costs available for PAM30 and PAM70](#)

The BLAST webpage now offers additional, more stringent, gap costs for PAM30 and PAM70.

Tue, 29 Jul 2014 13:00:00 EST

 [More BLAST news...](#)

Tip of the Day

[Integrating web PSI-BLAST with command line PSI-BLAST using the PssmWithParameters format](#)

This format of the PSSM can be directly used with other stand-alone Blast software tools, in particular as an input checkpoint file for blastpgp.

 [More tips...](#)

Data intensive biology *for everyone.*

Galaxy is an open, web-based platform for data intensive biomedical research. Whether on the free public server or your own instance, you can perform, reproduce, and share complete analyses.

Use Galaxy



Use project's free server or other public servers

Get Galaxy



Install locally or in the cloud or get Galaxy on SlipStream

Learn Galaxy



Screencasts, Galaxy 101, ...

Get Involved



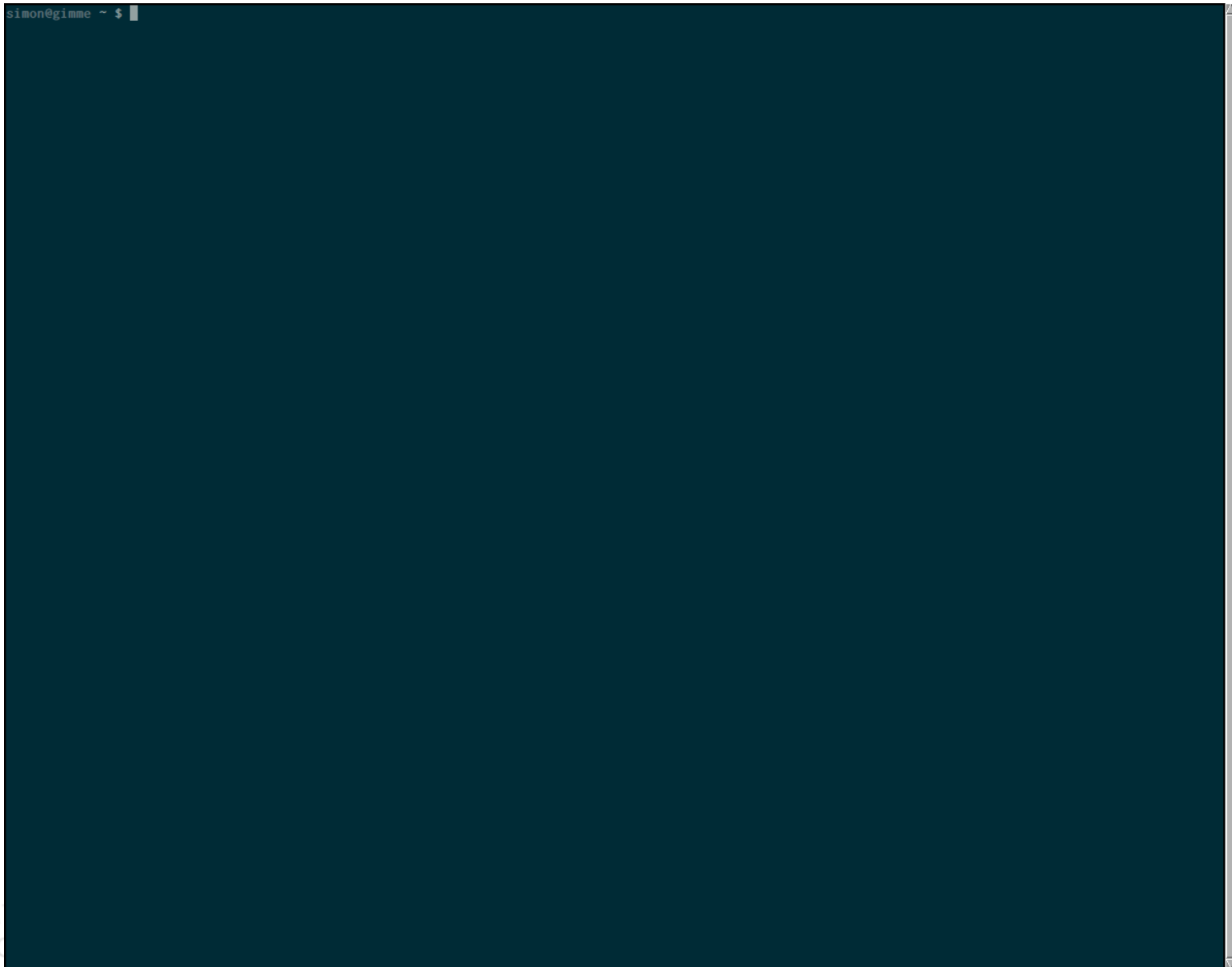
Mailing lists, Tool Shed, wiki

[Search all resources](#)

The Galaxy Team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.

An alternative

The command line



Why?

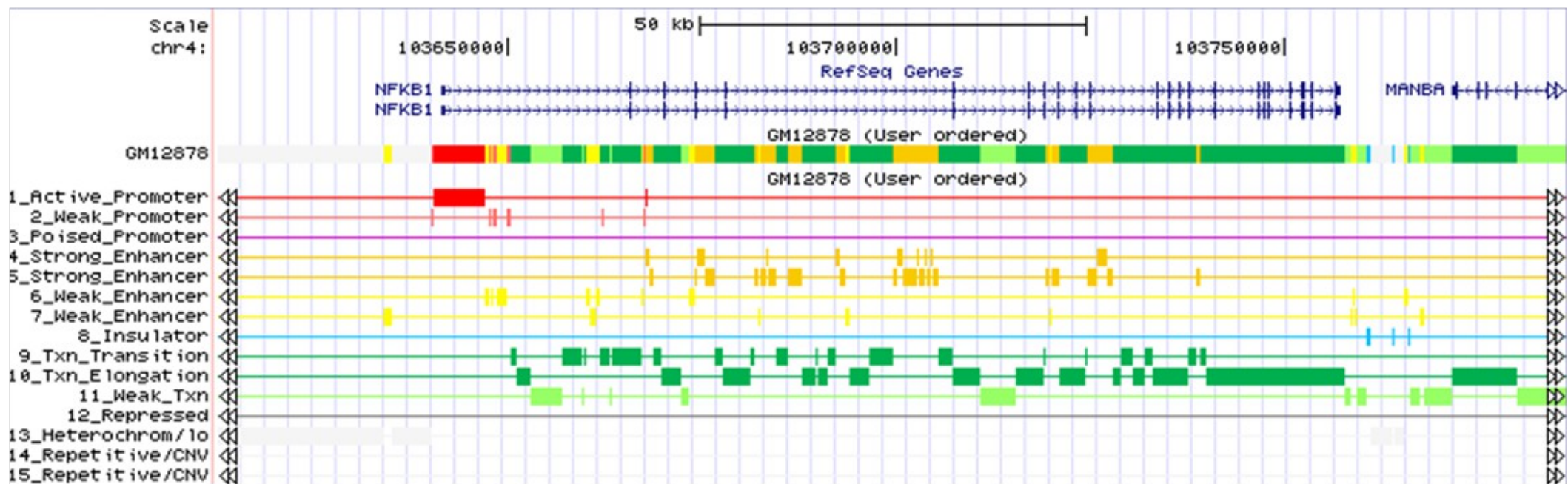


The command line

- Powerful
- Great control over what you're doing
- Run multiple jobs
 - at once
 - hundreds of 'em!
- Many computational tools don't have a GUI
- Reproducible, reusable research
 - (In theory...)



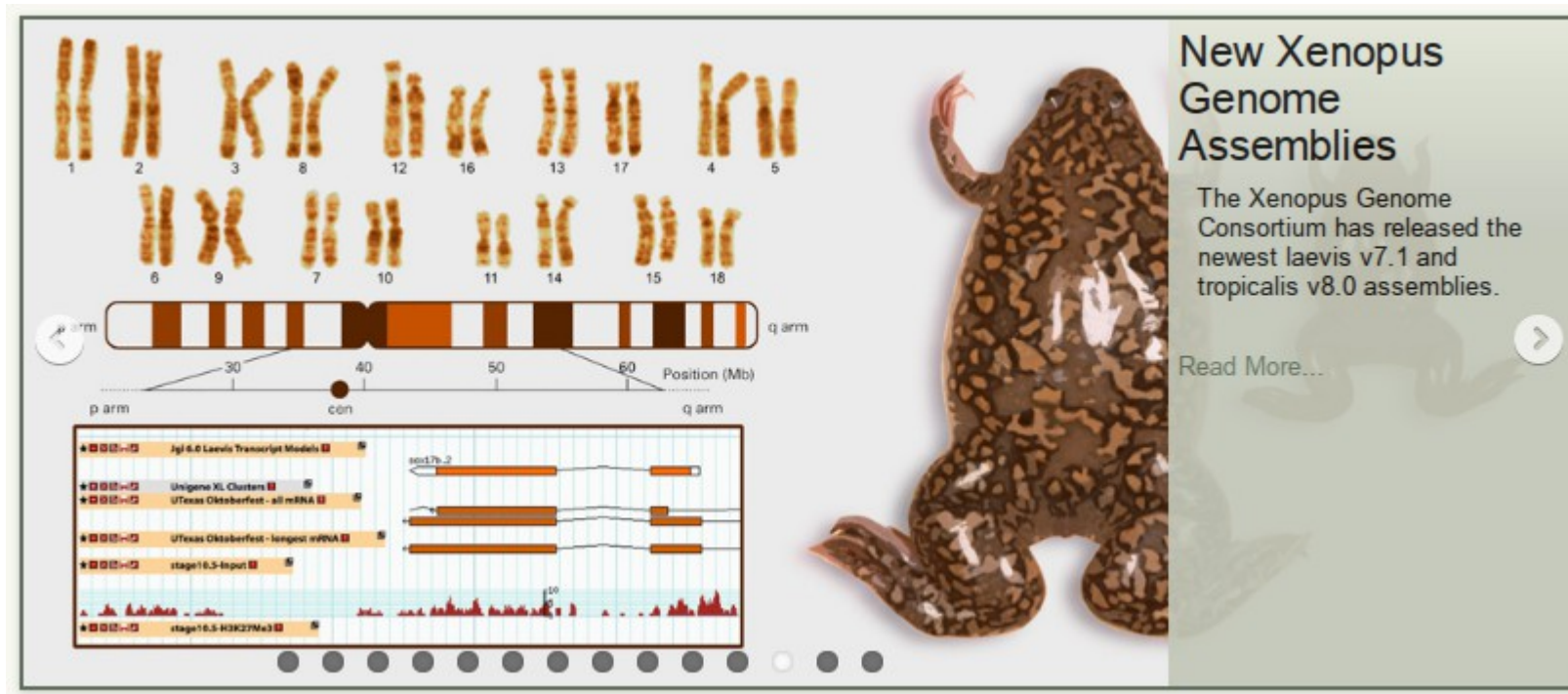
An example



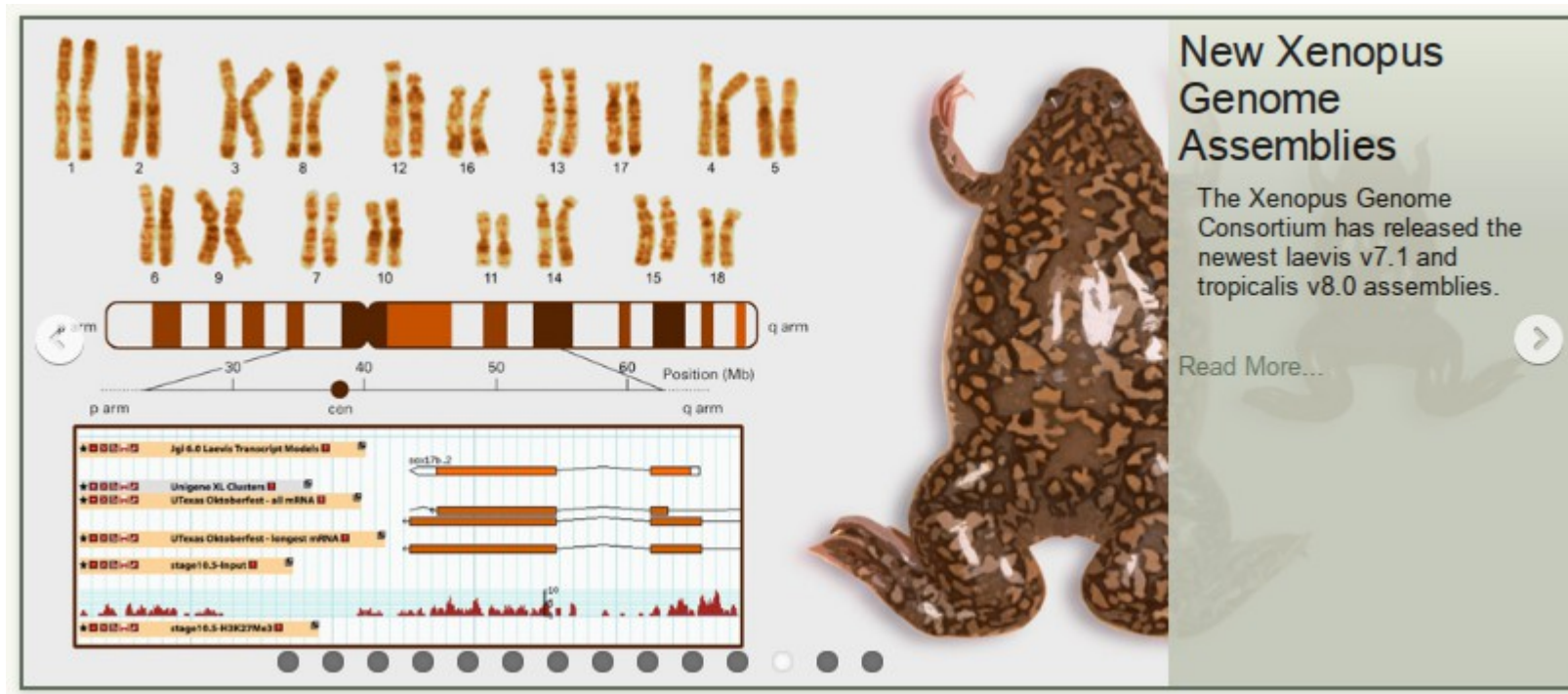
ChromHMM, Ernst & Kellis, 2012

An example

- Studying development in *Xenopus tropicalis*
- 10 different assays in 5 different stages of development
- Next-gen sequencing data
- Analysis steps:
 - 1) Mapping + “peak-calling”
 - 2) Combine data in ChromHMM and learn model
 - 3) Run analyses and make figures



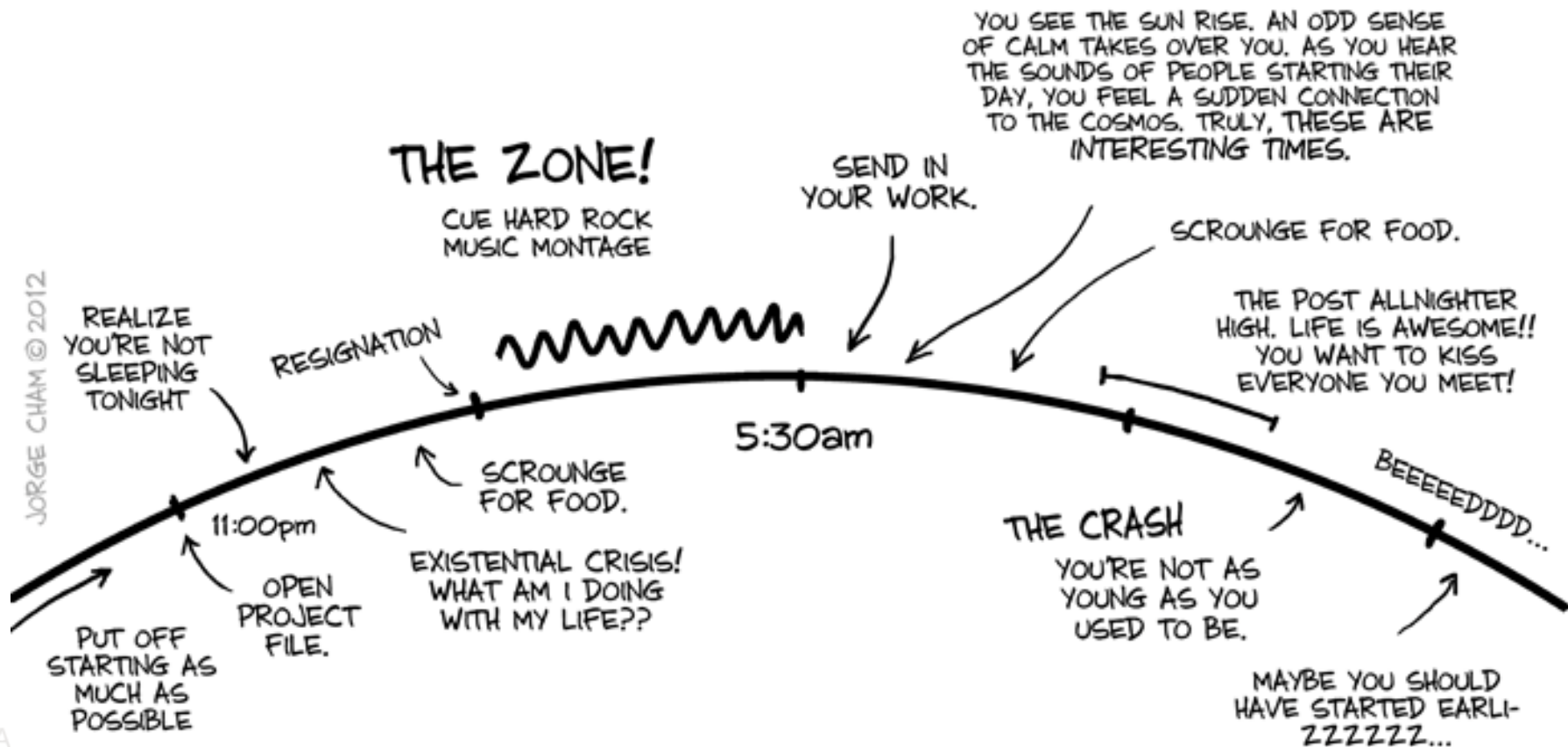
- Analysis steps:
 - 1) Mapping + “peak-calling”
 - 2) Combine data in ChromHMM and learn model
 - 3) Run analyses and make figures



- Analysis steps:
 - 1) **Mapping** + “peak-calling”
 - 2) Combine data in ChromHMM and learn model
 - 3) Run analyses and make figures

That's one unhappy PhD student..

THE ALLNIGHTER



Instead...

```
$ sed -i 's/JGI_7.1/JGI_8.0/' config.txt  
  
$ ./run_analysis.sh config.txt
```

- Change configuration file
- Start script
- ~~Go home and watch Netflix~~ Continue with new, exciting analysis

Other advantages

- Load-whole-file versus streaming
- Data doesn't always all fit into memory
- A lot of biological data is just text
- Can be processed line by line

General considerations

- Understand your goals
- Step-by-step, don't try to do it all at once
- Try to break your own scripts
- Choose appropriate methods and tools

The background of the image is a dark, atmospheric scene. It features a heavy, grey, and textured cloud formation that appears to be part of a storm or late evening sky. Below the clouds, the dark, silhouetted outlines of hills or mountains are visible against a slightly lighter, though still dark, horizon. The overall color palette is dominated by deep blues, greys, and blacks, creating a somber and mysterious mood.

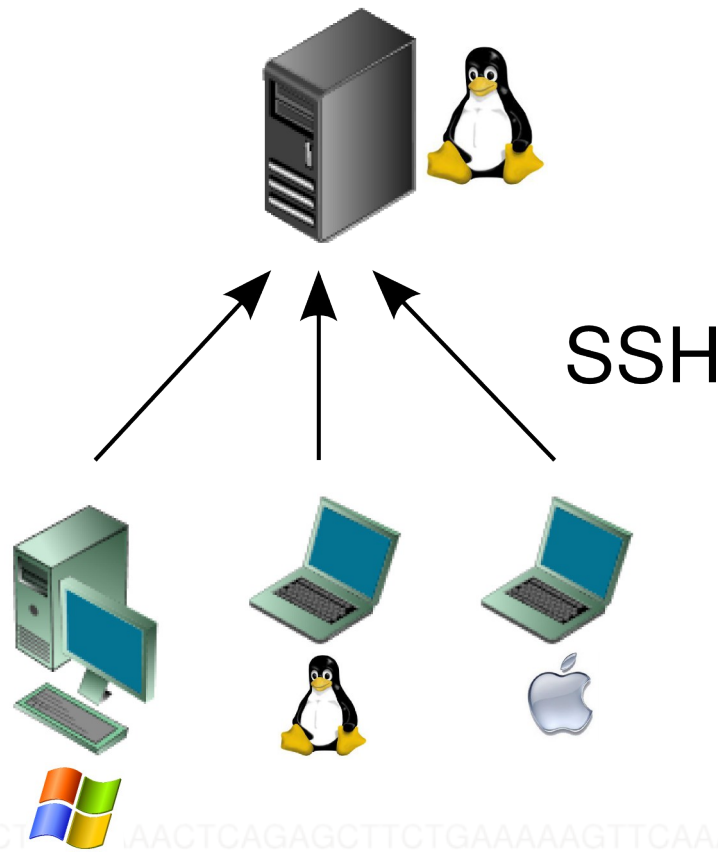
TRUST NO ONE

Today

- Familiarize yourself with the Linux command line
- Next-gen file formats
 - FASTQ
 - BAM
- Mapping (?)

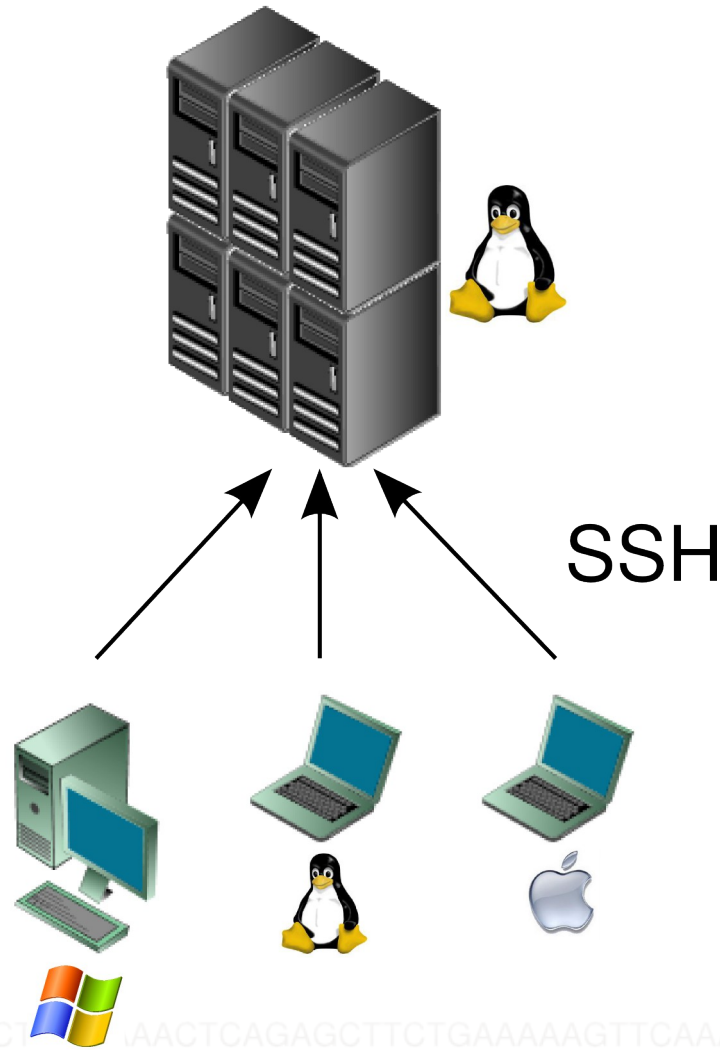
SSH (Secure Shell)

- Connect to server
- Clients available for every OS
- Perform analysis remotely

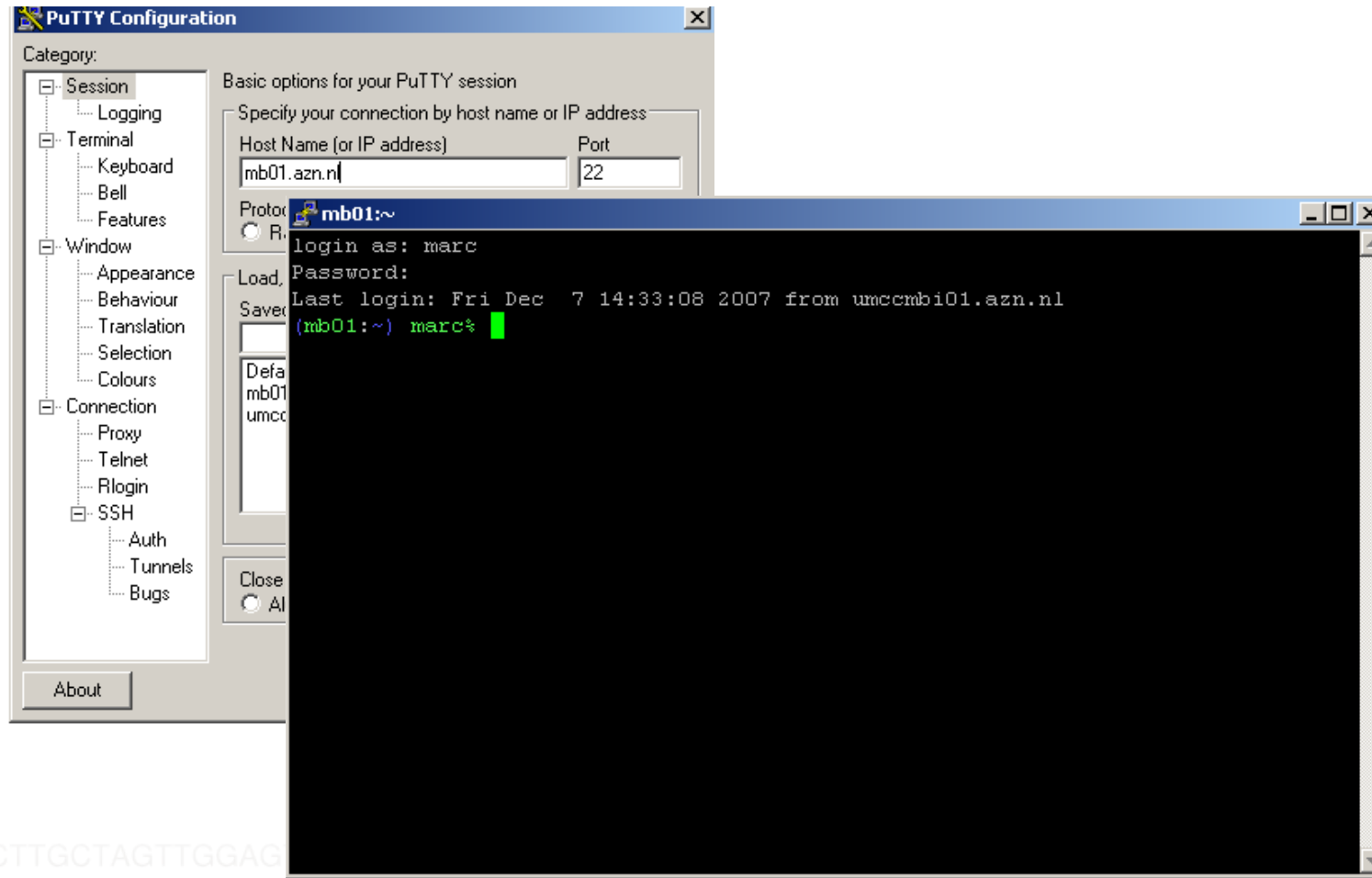


SSH (Secure Shell)

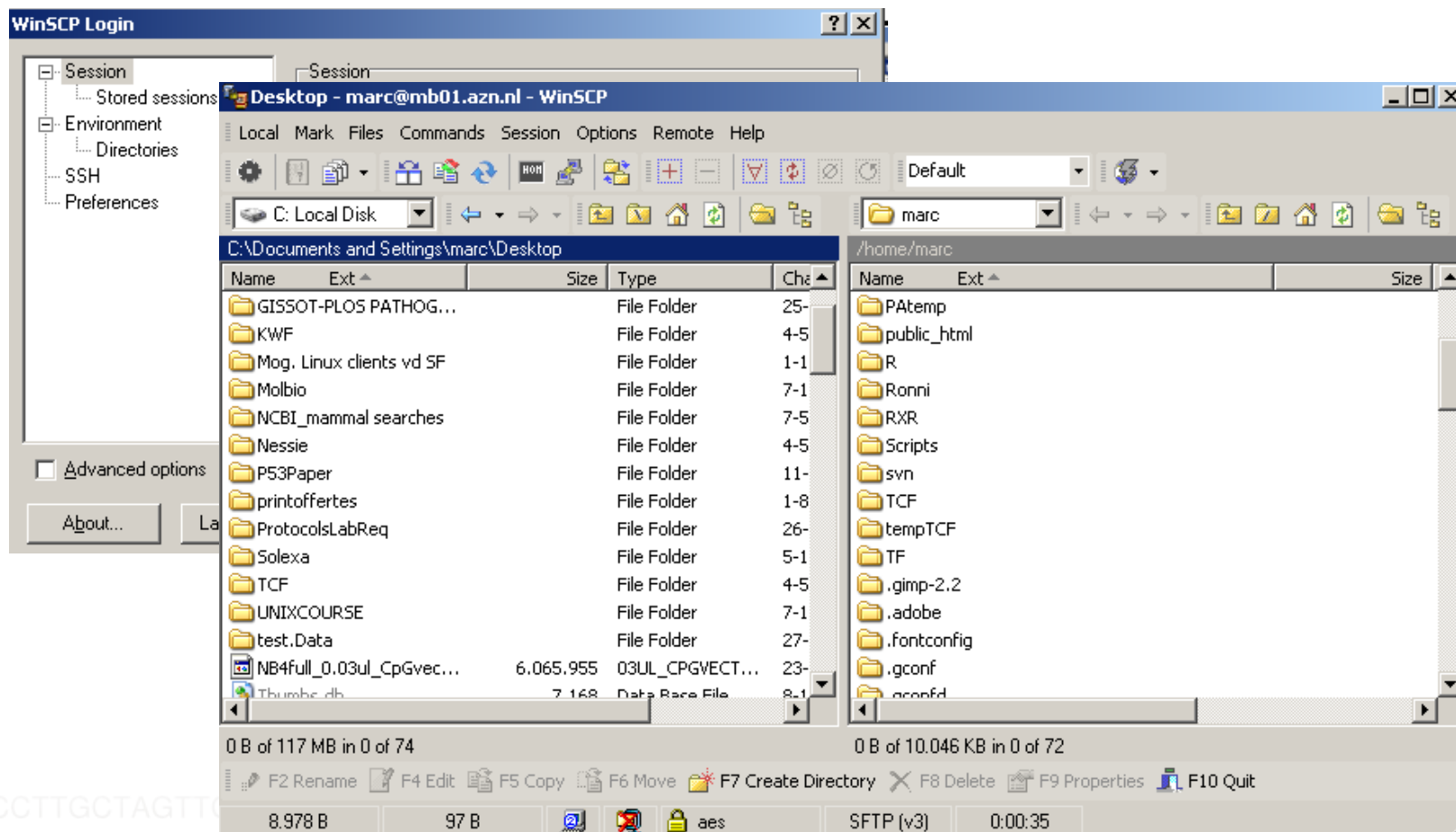
- Connect to server
- Clients available for every OS
- Perform analysis remotely
- Server can have lots of memory and CPU power



For Windows: PuTTY



For Windows: WinSCP



Server IP addresses

- 23.20.162.10
- 54.80.42.82
- 23.20.67.155
- 54.198.43.176
- 54.81.15.78
- 54.242.170.57
- 54.80.155.251